

模型引导的药物开发在抗体偶联药物领域的应用

吴白杨¹, 王 凌², 姜 静^{1,2*}

(1. 滨州医学院药学院, 山东 烟台 264003; 2. 荣昌生物制药(烟台)股份有限公司, 山东 烟台 264006)

摘要: 抗体偶联药物 (antibody drug conjugates, ADC) 作为抗肿瘤治疗的前沿技术, 近年来取得了显著进展。ADC 通过连接子将高活性小分子毒素与高特异性抗体进行偶联, 不仅能够实现对肿瘤细胞的精准打击, 同时降低了药物的全身毒性, 进而扩大了治疗的有效性和安全性窗口。然而, 由于 ADC 分子设计的复杂性, 其疗效和安全性受多种因素影响。模型引导的药物开发 (model informed drug development, MIDD) 是一种通过数学和统计模型进行建模和模拟, 对药物研发进行定量分析和决策指导的方法。这种方法为新药研发提供强大的工具支持。通过 MIDD 整合 ADC 相关的多方面数据和信息, 有助于理解 ADC 的复杂机制、药代动力学和药效学等作用特征, 为优化 ADC 研发流程和临床转化决策提供独特见解。本文将介绍 MIDD 和 ADC 的基本概念, 并浅析 MIDD 在 ADC 研发不同阶段的应用案例, 旨在为 ADC 的发展提供有益参考。

关键词: 模型引导的药物开发; 抗体偶联药物; 建模与模拟; 药代动力学/药效动力学; 定量系统药理学; 生理药代动力学; 群体药代动力学

中图分类号: R914

文献标识码: A

文章编号: 0513-4870(2025)02-0288-12

Application of model informed drug development in the field of antibody drug conjugates

WU Bai-yang¹, WANG Ling², JIANG Jing^{1,2*}

(1. School of Pharmacy, Binzhou Medical University, Yantai 264003, China; 2. RemeGen Co., Ltd., Yantai 264006, China)

Abstract: Antibody drug conjugates (ADC) have emerged as a cutting-edge technology in anti-tumor treatment, making significant strides in recent years. ADC couple a highly active small molecule toxin payload to highly specific antibodies through a linker, enabling precise targeting of tumor cells while reducing systemic toxicity, thereby expanding the therapeutic window. However, due to the complexity of ADC molecule design, its efficacy and safety are influenced by various factors. Model-informed drug development (MIDD) is a powerful tool that utilizes various mathematical models for modeling and simulation to conduct quantitative analysis, guiding drug development and decision-making. By integrating multi-faceted data and information using mathematical models, it is possible to gain insights into the complex mechanisms, pharmacokinetics, and pharmacodynamics of ADC, providing unique perspectives for optimizing ADC development processes and clinical translation decisions. This review will introduce the basic concepts of MIDD and ADC and discuss application cases of MIDD in different stages of ADC development, aiming to provide beneficial references for the advancement of ADC.

Key words: model informed drug development; antibody drug conjugate; modeling and simulation; pharmacokinetics/pharmacodynamics; quantitative systems pharmacology; physiologically based pharmacokinetics; population pharmacokinetics

收稿日期: 2024-08-14; 修回日期: 2024-11-19.

基金项目: 山东省自然科学基金项目 (R2021MH220); 山东省泰山产业领军人才项目.

*通讯作者 E-mail: j.jiang@bzmc.edu.cn

DOI: 10.16438/j.0513-4870.2024-0787

抗体偶联药物 (antibody drug conjugates, ADC) 是一种新型抗肿瘤药物, 与传统化疗药物相比, 其结合了单克隆抗体的高靶向性和细胞毒素的高活性, 在实现精准高效杀伤肿瘤细胞的同时降低对正常组织的毒副作用。ADC 药物在某些难治性肿瘤中的治疗效果显著优于传统单克隆抗体, 被誉为肿瘤治疗领域的“魔法子弹”^[1]。目前, 全球已有 15 款 ADC 成功获批上市 (表 1), 上百种 ADC 正在积极进行临床评估, 充分展现出 ADC 在肿瘤治疗领域的巨大潜力^[2]。

ADC 研发管线正在不断被扩大, 大量资源和时间被投入到分子筛选及优化、评估疗效与安全性、实现临床转化以及临床方案设计等关键环节中。鉴于 ADC 药物独特的结构和复杂的作用机制, 仅依赖传统实验方法可能难以甚至无法全面理解各个组成部分对药物疗效及安全性的影响。模型引导的药物研发 (model informed drug development, MIDD) 通过建模与模拟

(modeling and simulation, M&S) 技术整合生理学、药理学和疾病过程等信息通过数学框架进行定量研究, 进而指导新药研发和决策^[3-5]。随着 MIDD 在新药研发领域的不断发展, 研究人员意识到将其应用于 ADC 研发将有助于实现这一目标^[6,7]。MIDD 为整合 ADC 不同研究阶段、不同维度的数据信息提供了一个定量框架, 进而为药物研发过程中“继续/停止”决策提供了重要的参考依据。这不仅有助于改善现有数据不足的困境, 还能有效提高研发效率, 降低药物研发失败的风险^[8,9]。本文将简要介绍 MIDD 和 ADC 的基本概念, 并分析 MIDD 在 ADC 研发不同阶段的应用案例, 旨在为 ADC 的发展提供有益的参考与启示。

1 MIDD 在 ADC 的应用背景

1.1 MIDD 的概念及发展历程 MIDD 是通过 M&S 技术整合生理学、药理学及疾病过程等信息, 进而指导新药研发和决策的定量研究方法。近年来, 随着对

Table 1 List of approved of antibody drug conjugates (ADC). HER2: Human epidermal growth factor receptor 2; Trop-2: Tumor associated antigen 2; BCMA: B-cell maturation antigen; EGFR: Epidermal growth factor receptor; MMAE: Monomethyl muristatin E; PE38: Pseudomonas exotoxin; Dxd: Deruxtecan; PBD: Pyrrolbenzodiazepine; DM: Dexamethasone methylation

Drug name	Trade name	First approved	Indication	Target antigen	Payload	Linker
Gemtuzumab ozogamicin	Mylotarg	2000	CD33-positive acute myeloid leukemia	CD33	Calicheamicins	Cleavable linker acid-labile hydrazone
Brentuximab vedotin	Adcetris	2001	CD30-positive Hodgkin lymphoma and relapsed systemic anaplastic large cell lymphoma	CD30	MMAE	Cleavable linker maleimidocapmyl valine citrulline
Trastuzumab emtansine	Kadcyla	2013	HER2-positive metastatic breast cancer	HER2	DM1	Non-cleavable linker based thioether
Inotuzumab ozogamicin	Besponsa	2017	Relapsed or refractory B-cell acute lymphoblastic leukemia	CD22	Calicheamicins	Cleavable linker acid-labile hydrazone
Moxetumomab pasudotox	Lumoxiti	2018	Relapsed or refractory hairy cell leukemia	CD22	PE38	Cleavable linker mc-vc-PABC
Polatuzumab vedotin	Polivy	2019	Relapsed or refractory diffuse large B-cell lymphoma	CD79B	MMAE	Cleavable linker maleimidocapmyl valine citrulline
Enfortumab vedotin	Padcev	2019	Locally advanced or metastatic urothelial cancer	Nectin-4	MMAE	Cleavable linker maleimidocapmyl valine citrulline
Trastuzumab deruxtecan	Enhertu	2019	Unresectable or metastatic HER2-positive breast cancer	HER2	Dxd	Cleavable linker based tetrapeptide
Sacituzumab govitecan	Trodelyv	2020	Triple-negative breast cancer with relapsed or refractory metastatic disease	Trop-2	SN38	Cleavable carbonate linker
Belantamab mafodotin	Blenrep	2020	Relapsed or refractory multiple myeloma	BCMA	MMAF	Non-cleavable linker maleimidocapryl
Cetuximab sarotalocansodium	Akalux	2020	Unresectable locally advanced or recurrent head and neck cancer	EGFR	IRDye700DX	Non-cleavable linker
Loncastuximab tesirine	Zynlonta	2021	Relapsed or refractory large B-cell lymphoma	CD19	PBD	Cleavable linker valine-alanine
Disitamab vedotin	Aidixi	2021	The integration of a stomach/stomach esophagus adenocarcinoma, breast cancer, urothelial carcinoma	HER2	MMAE	Cleavable linker MC-Val-Cit-PAB
Tisotumab vedotin	Tivdak	2021	Recurrent or metastatic cervical cancer	Tissue factor	MMAE	Cleavable linker valine-citrulline
Mirvetuximab soravtansine	Elahere	2022	FR α -positive platinum-resistant ovarian cancer	FR α	DM4	Cleavable disulfide bond linker

MIDD的认知和价值的不断深化, MIDD已广泛应用于药物研发的各个阶段, 涵盖药物发现、临床前研究、临床开发、监管评估以及药物全生命周期管理, 对药物研发决策的证据支持和指导具有重要意义。模型分析与实测研究过程通常呈现出“学习与确认”循环形式, 即通过已有数据信息建立模型, 预测相关研究结果, 进一步通过后续实测数据验证模型分析结果的可靠性及判断后续研究方向, 并随着研发过程的推进对模型进行不断更新和完善(图1)^[10-13]。

MIDD应用多方面模型工具, 包括群体药代动力学(population pharmacokinetics, Pop-PK)、生理药理学(physiologically based pharmacokinetics, PBPK)、药动学/药效学(pharmacokinetics/pharmacodynamics, PK/PD)、暴露-反应关系(exposure-response, E-R)、基于模型的荟萃分析(model-based meta-analysis, MBMA)及定量系统药理学(quantitative systems pharmacology, QSP)等已经在药物研发的多个环节取得了成功应用, 例如适应症与剂量选择、非临床与临床转化研究以及临床药理学研究等方面^[14]。

目前, 这些工具在美国FDA批准的药物中的使用比例高达90%以上, 已经成为药物研发领域不可或缺的关键工具。全球众多监管机构包括美国食品药品监督管理局(Food and Drug Administration, FDA)、国家药品监督管理局药品审评中心(Center for Drug Evaluation, CDE)、欧洲药品管理局(European Medicines Agency, EMA)等均发布了多项涉及MIDD的指南和药品申报要求, 这些指导原则旨在鼓励并指导制

药企业运用MIDD, 以进一步提升药物的商业、科学和临床价值^[15-20]。2018年, FDA在处方药使用者方案第6次修订版(PDUFA VI)正式认定MIDD为高效和有效药物开发的重要推动因素, 鼓励药物开发人员和美国FDA审查人员能够共同参与讨论药物开发中使用MIDD工具, 有助于药品不同开发环节的研发人员和审评人员就剂量选择、临床试验模拟、机制化安全性评估等方面的决策更早达成共识^[15]。CDE于2020年发布的《模型引导的药物研发技术指导原则》将有助于提升国内制药工业界的实践能力, 促进制药行业的技术进步和创新^[17]。为了进一步指导我国创新药物临床研究阶段剂量探索和优化, 并提供可参考的技术标准, CDE于2024年7月25日起草了《模型引导的创新药物剂量探索和优化技术指导原则(征求意见稿)》^[20]。

1.2 ADC的结构、作用机制及特征 相较于传统药物, ADC具有独特的结构设计和复杂的作用机制。ADC通常由靶向特异性抗原的单克隆抗体、化学连接子及有效载荷(小分子毒素)三部分组成, 每个抗体分子上平均偶联的小分子毒素数量称为抗体药物比(drug antibody ratio, DAR)。ADC的作用机制(图2)是通过抗体部分特异性地识别并结合到目标细胞表面的抗原, 随后通过内吞作用进入细胞内部。在细胞内, ADC经溶酶体作用后连接子断裂, 释放出细胞毒素, 这些毒素能够结合至微管蛋白、DNA或拓扑异构酶, 进而诱导细胞死亡。同时, 部分ADC可以与内吞体的新生儿Fc受体(neonatal Fc receptor, FcRn)结合, 导致ADC循环至细胞外。一些疏水性小分子毒素还可通过细胞扩散,

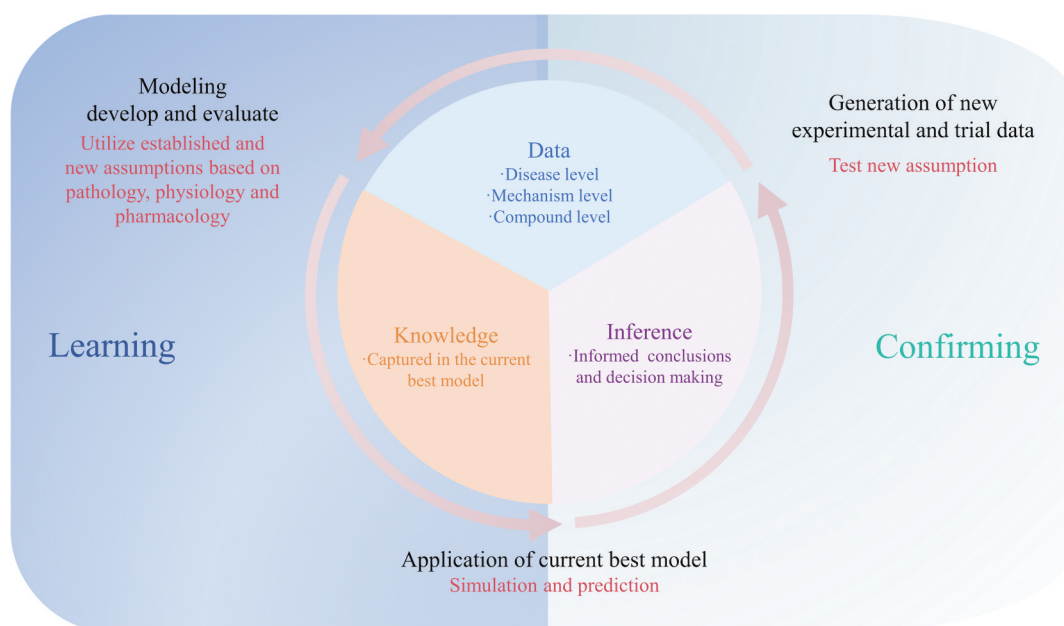


Figure 1 A quantitative framework flow of "Learn and Confirm Cycle". Image adapted with permission from reference^[13]. Copyright © John Wiley and Sons Ltd.

对邻近肿瘤细胞产生杀伤活性,称为旁观者效应^[21]。

ADC的分子设计使其同时结合了单克隆抗体药物和小分子毒素药物两者的PK和PD特征。ADC通常经过静脉给药,从分子量大小和空间体积来看,小分子毒素药物的相对分子质量为裸抗体的1/150。因此,ADC表现出了诸多与抗体药物类似的PK特征,如靶点介导的药物清除、FcRns受体循环作用及非特异性蛋白酶降解等。然而,ADC在代谢和清除途径过程中同时具有抗体药物和小分子毒素特征,即在低剂量时呈非线性,高剂量时呈线性。ADC进入体内后,小分子毒素通过酶解或化学反应逐渐从ADC药物上解离下来,进一步增加了ADC药物在体内的复杂性,导致在进行PK研究时需要大量的分析物进行分析检测,例如总抗体浓度、结合抗体浓度、游离小分子浓度等,以便更好地了解ADC药物的体内过程^[22-25]。

因此,ADC药物的PK/PD之间的关系不仅需要综合考虑各种组成成分(抗体、连接子和小分子毒素)的特征,还需要考虑它们之间的相互作用。ADC的PK特征取决于抗体和小分子毒素在体内的动态变化,PD特征通常涉及ADC与作用靶细胞之间的相互作用程度及其产生的效应。PK/PD整合分析有助于理解ADC在体内的行为,评估药物浓度与疗效和毒性之间的关系,为临床决策提供科学依据。

2 MIDD在ADC领域的应用

截至目前,已有不少学术机构及医药公司发表了

与ADC药物相关的模型研究案例(表2)^[26-66]。通过整合在研发过程中生成的数据,研究者们构建了数学模型框架来解决各个研发阶段的目标和问题。随着M&S技术的进步,这些模型研究不仅仅依赖于传统的经验/机械模型,同时还结合了ADC的作用机制及其在肿瘤内的分布特征进行建模(图3)^[67]。下文通过将MIDD在ADC研发及临床阶段已有的应用参考案例归纳为分子筛选及设计优化、加速临床转化、评价药物-药物相互作用(drug-drug interaction, DDI)及临床剂量探索及优化。

2.1 分子筛选及设计优化 在药物研发的早期阶段,筛选和优化适合临床研究的ADC分子是关键任务。为了更高效地完成这一任务,研究人员需设计和开发合适的模型,并确保将模型输出结果有效地分享给团队成员,这对于药物研发的决策制定具有重大意义。

Maass等^[33]构建了描述ADC细胞水平作用机制PK模型,该模型纳入了可能影响ADC在细胞内处理和有效载荷释放的相关参数。同时基于流式细胞术和荧光成像法对关键参数建立了定量方法,如抗体与抗原的结合和解离常数、ADC内吞速率和降解速率等。通过比较不同参数设置下细胞内释放的小分子毒素药物的曲线下面积(AUC)变化进行敏感性分析,有助于了解细胞内处理步骤与设计参数之间的相互影响。结果表明ADC的内吞速率和小分子毒素药物的外排速率是影响药物在细胞中暴露量关键因素。Wada等^[30]

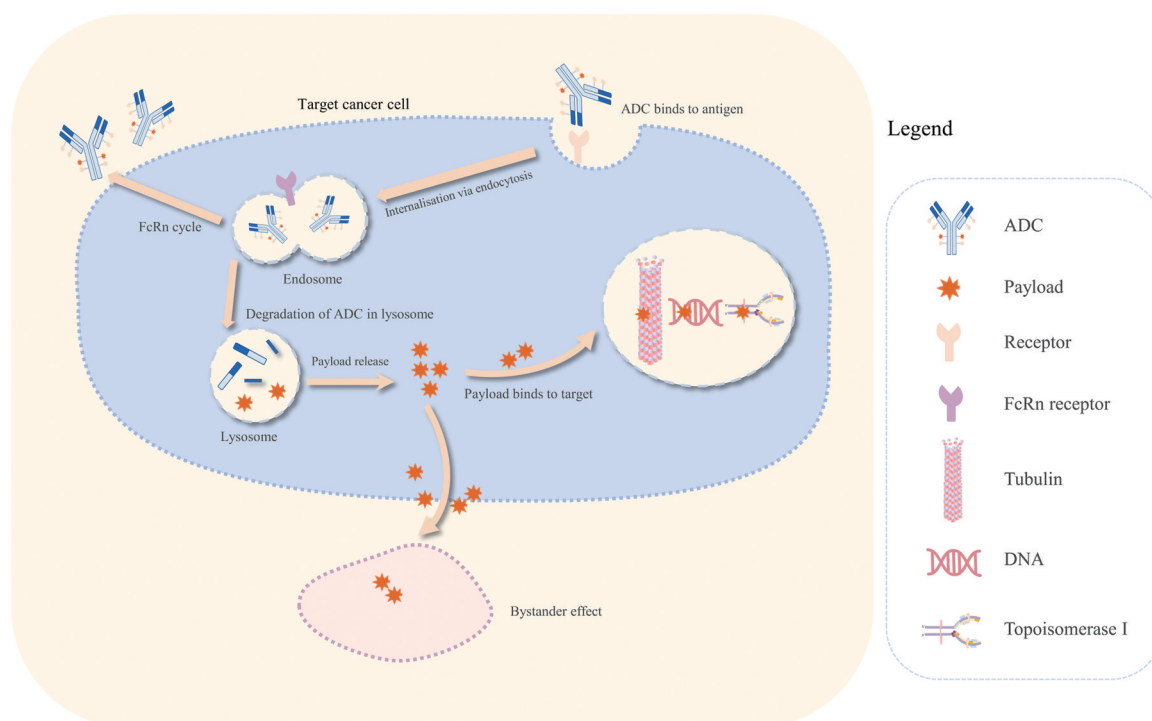


Figure 2 Mechanism action of antibody drug conjugates. FcRn: Neonatal Fc receptor

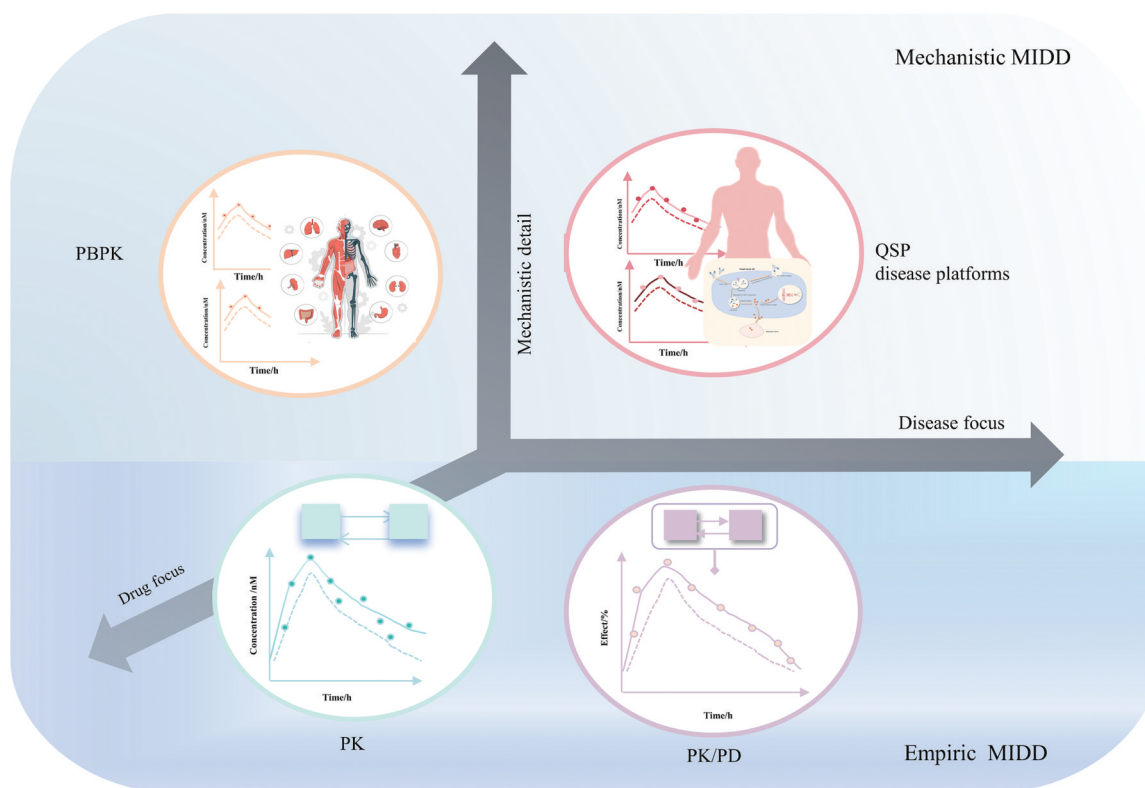


Figure 3 Schematic overview of model informed drug development approaches of ADC. MIDD: Model informed drug development

Table 2 List of prominent models for ADC. PK: Pharmacokinetics; PD: Pharmacodynamics; Pop-PK: Population pharmacokinetics; E-R: Exposure-response; PBPK: Physiologically based pharmacokinetic; QSP: Quantitative systems pharmacology; IVIVC: *In vitro-in vivo* correlation; DDI: Drug-drug interaction; MMAF: Monomethyl auristatin F; STEAP1: Six-transmembrane epithelial antigen of the prostate

Model	ADC-modeled	Data source	Reference
Bench to bedside translation of ADC using a multiscale mechanistic PK/PD model: a case study with brentuximab-vedotin	Brentuximab-vedotin	<i>In vitro/in vivo/clinical</i>	[26]
On translation of ADC efficacy from mouse experimental tumors to the clinic: a PK/PD approach	T-DM1 and 5T4-mc-MMAF	<i>In vivo/clinical</i>	[27]
A priori prediction of tumor payload concentrations: preclinical case study with an auristatin-based anti-5T4 ADC	5T4-mc-MMAF	<i>In vitro/in vivo</i>	[28]
A mechanistic PK model elucidating the disposition of T-DM1, an ADC for treatment of metastatic breast cancer	T-DM1	<i>In vivo</i>	[29]
Mechanistic PK/PD modeling of <i>in vivo</i> tumor uptake, catabolism, and tumor response of trastuzumab maytansinoid conjugates	T-DM1 and T-SPP-DM1	<i>In vitro/in vivo</i>	[30]
A mechanistic tumor penetration model to guide ADC design	General ADC	<i>In vitro/in vivo</i>	[31]
PBPK modeling as a tool to predict drug interactions for ADC	ADC based vc-MMAE	<i>In vivo/clinical</i>	[32]
Determination of cellular processing rates for a trastuzumab-maytansinoid ADC highlights key parameters for ADC design	Trastuzumab-maytansinoid	<i>In vitro</i>	[33]
Evolution of ADC tumor disposition model to predict preclinical tumor PKs of T-DM1	T-DM1	<i>In vitro/in vivo</i>	[34]
Preclinical to clinical translation of ADCs using PK-PD modeling: a retrospective analysis of inotuzumab ozogamicin	Inotuzumab ozogamicin	<i>In vitro/in vivo/clinical</i>	[35]
Multiscale modeling of ADCs: connecting tissue and cellular distribution to whole animal PKs and potential implications for efficacy	T-DM1	<i>In vitro/in vivo</i>	[36]
Quantitative characterization of <i>in vitro</i> bystander effect of ADC	Trastuzumab-vc-MMAE	<i>In vitro</i>	[37]
Development and translational application of an integrated, mechanistic model of ADC PKs	anti-STEAP1-vc-MMAE	<i>In vitro/in vivo/clinical</i>	[38]

			Continued
Model	ADC-modeled	Data source	Reference
Application of a PK/PD modeling and simulation-based strategy for clinical translation of ADCs: a case study T-DM1	T-DM1	<i>In vivo</i> /clinical	[39]
A mechanism-based PK/PD model for hematological toxicities induced by ADCs	Brentuximab vedotin and T-DM1	<i>In vivo</i>	[40]
Pop-PK of brentuximab vedotin in patients with CD30-expressing hematologic malignancies	Brentuximab vedotin	Clinical	[41]
Measurement and mathematical characterization of cell-level PKs of ADCs: a case study with trastuzumab-vc-MMAE	Trastuzumab-vc-MMAE	<i>In vivo</i>	[42]
Platform model describing PK properties of vc-MMAE ADC	ADC based vc-MMAE	Clinical	[43]
Development of a translational PBPK model for ADC: a case study with T-DM1	T-DM1	<i>In vitro</i> / <i>in vivo</i> /clinical	[44]
Computational transport analysis of ADC bystander effects and payload tumoral distribution: implications for therapy	Trastuzumab-vc-MMAE and T-DM1	<i>In vitro</i> / <i>in vivo</i>	[45]
Establishing IVIVC for ADC efficacy: a PK/PD modeling approach	19 different ADCs	<i>In vitro</i> / <i>in vivo</i>	[46]
A "dual" cell-level systems PK-PD model to characterize the bystander effect of ADC	Trastuzumab-vc-MMAE	<i>In vitro</i>	[47]
A cell-level systems PK/PD model to characterize <i>in vivo</i> efficacy of ADCs	Trastuzumab-vc-MMAE	<i>In vitro</i> / <i>in vivo</i>	[48]
PK/PD modeling to support the re-approval of gemtuzumab ozogamicin	Gemtuzumab ozogamicin	Clinical	[49]
Antibody coadministration as a strategy to overcome binding-site barrier for ADCs: a quantitative investigation	Trastuzumab-vc-MMAE and T-DM1	<i>In vitro</i> / <i>in vivo</i>	[50]
An agent-based systems pharmacology model of the ADC kadcyla to predict efficacy of different dosing regimens	T-DM1	<i>In vitro</i> / <i>in vivo</i>	[51]
Evaluation of quantitative relationship between target expression and ADC exposure inside cancer cells	Trastuzumab-vc-MMAE	<i>In vitro</i>	[52]
PBPK model-informed drug development for polatuzumab vedotin: label for DDI without dedicated clinical trials	Polatuzumab vedotin	<i>In vitro</i> / <i>in vivo</i> /clinical	[53]
Pop-PK of brentuximab vedotin in adult and pediatric patients with relapsed/refractory hematologic malignancies: model-informed hypothesis generation for pediatric dosing regimens	Brentuximab vedotin	Clinical	[54]
Evolution of the systems PK-PD model for ADCs to characterize tumor heterogeneity and <i>in vivo</i> bystander effect	Trastuzumab-vc-MMAE	<i>In vitro</i> / <i>in vivo</i>	[55]
Mechanistic modeling of intra-tumor spatial distribution of ADC: insights into dosing strategies in oncology	General ADC	<i>In vivo</i> /clinical	[56]
Pop-PK of trastuzumab deruxtecan in patients with HER2-positive breast cancer and other solid tumors	T-Dxd	Clinical	[57]
E-R relationships in patients with HER2-positive metastatic breast cancer and other solid tumors treated with trastuzumab deruxtecan	T-Dxd	Clinical	[58]
Pop-PK of belantamab mafodotin, a BCMA-targeting agent in patients with relapsed/refractory multiple myeloma	Belantamab mafodotin	Clinical	[59]
Simulating the selection of resistant cells with bystander killing and antibody co-administration in heterogeneous HER2-positive tumors	Trastuzumab-vc-MMAE and T-DM1	<i>In vitro</i> / <i>in vivo</i>	[60]
Towards a platform QSP model for preclinical to clinical translation of ADCs	T-DM1 and T-Dxd	<i>In vitro</i> / <i>in vivo</i> /clinical	[61]
PK and PD of ADC administered <i>via</i> subcutaneous and intratumoral routes	Trastuzumab-vc-MMAE	<i>In vivo</i>	[62]
Pop-PK of patritumab deruxtecan in patients with solid tumors	Patritumab deruxtecan	Clinical	[63]
Development of a generalized PK model to characterize clinical PK of MMAE-based ADCs	MMAE based ADC	<i>In vivo</i> /clinical	[64]
Quantitative evaluation of trastuzumab deruxtecan PK and PD in mouse models of varying degrees of HER2 expression	T-DXd	<i>In vitro</i> / <i>in vivo</i> /clinical	[65]
Optimizing solid tumor treatment with ADC using agent-based modeling: considering the role of a carrier dose and payload class	T-DM1 and T-Dxd	<i>In vitro</i> / <i>in vivo</i>	[66]

通过PK/PD模型比较了两种不同连接子的ADC在肿瘤内小分子毒素浓度及肿瘤药效方面的差异性。模拟结果表明,连接子的不同会影响小分子毒素的释放特

性,连接子为二硫键的T-SPP-DM1比连接子为硫醚键的trastuzumab emtansine (T-DM1)可能具有更快的小分子毒素释放特性,这可能导致肿瘤内细胞毒性药物

的浓度更高。Shah 等^[46]开发了一种基于 19 种不同 ADC 药物的 PK/PD 模型, 并通过肿瘤静态浓度 (tumour static concentration, TSC) 建立体外-体内相关性 (*in vitro-in vivo* correlation, IVIVC)。模拟结果表明, 这些 ADC 药物在体内的 TSC 与体外的 TSC 呈线性正相关。因此, 这种合理的 IVIVC 方法能够利用体外药效数据来预测体内药效情况, 为体内药效实验设计提供参考。Singh 等^[37]开发了用以表征 ADC 的旁杀伤效应的体外 PD 模型, 该模型结合了两种不同的细胞分布模型来反映共培养系统中抗原高表达和低表达的细胞群体。这项研究提供了一个定量旁杀伤效应的框架, 有助于筛选出具有最佳旁杀伤能力的新型 ADC。Ait-Oudhia 等^[40]基于 T-DM1 和 Brentuximab vedotin 开发了评估 ADC 血液毒性的 PK/PD 模型。模型模拟结果表明, 小分子毒素的释放速率是影响 ADC 血液学毒性的一个关键因素, 因此调整小分子毒素释放速率是改善血液毒性, 优化 ADC 治疗窗口的有效途径。

MIDD 不仅对影响 ADC 药物性能的多个因素进行了参数化定量分析, 而且还利用有限的药物分子的数据为药物分子的筛选和优化提供了独特的见解。即借助模型预测, 研究人员能够在早期阶段掌握不同结构组合 (如连接子稳定性、抗体内吞速率、小分子毒素外排效率及血液毒性等) 对药物疗效与安全性的影响, 为更有效地筛选和优化 ADC 药物设计提供不同路径, 进而发掘更安全有效的药物分子。

2.2 加速临床转化 如何正确地转化临床前的研究结果, 预测临床疗效, 以提高临床转化率一直是 ADC 开发过程中面临的挑战。较低的临床转化率的主要原因之一在于难以将临床前的 E-R 关系正确地转化应用于临床中。此外, ADC 药物具有复杂的作用机制及 PK 行为, 肿瘤组织中的药物浓度与血浆中的药物浓度在大多数情况下呈现不平衡状态, 肿瘤对 ADC 的摄取量并不等于其发挥疗效的药量。因此, 如果仅依赖传统经验模型来推测 ADC 药物浓度与疗效之间的关系可能并不可靠, 临床前和临床肿瘤生长抑制 (tumor growth inhibition, TGI) 的结果存在较大的差异。因此, 开发合理的模型用以描绘 ADC 肿瘤内暴露量与疗效之间的关系并转化到临床是临床前研究的核心任务。

Shah 等^[26]首次建立了基于 brentuximab vedotin 的机制 PK/PD 模型, 该模型描述了 ADC 在全身暴露过程并纳入了关键肿瘤生物指标 (如肿瘤大小、抗原表达水平及抗原内化等), 同时成功地基于临床前的研究对药物的临床效果进行了预测。基于这一模型基础, Singh 等^[39]等进一步开发了基于机制 PK/PD 模型转化

ADC 临床前至临床的通用策略, 并在 T-DM1 的研发过程中进行验证。首先, 该策略根据 T-DM1 临床前荷瘤小鼠研究建立肿瘤处置 PK/PD 模型; 随后, 将食蟹猴的 PK 参数通过异速放大至人体, 以预测药物在人体的 PK 行为; 最后将预测的人体 PK 数据、小鼠 PK/PD 模型估算的疗效参数与临床上观察到的乳腺癌体积和生长参数结合, 转化为人体临床 PK/PD 模型。最后, 该研究利用转化后的 PK/PD 模型, 进行了临床试验模拟, 即预测了 T-DM1 的无进展生存率 (progression-free survival, PFS) 和客观缓解率 (objective response rate, ORR)。模型模拟的人类表皮生长因子受体 2 (human epidermal growth factor receptor 2, HER2) 1+ 和 3+ 人群的 PFS 率结果与三个不同临床试验中观察到的相当。Scheuher 等^[61]建立了一个多尺度的 QSP 模型, 更多的机制细节被纳入模型。该模型使用 T-DM1 和 trastuzumab deruxtecan (T-DXd) 进行模型的开发、校准和验证。该模型与上述 Singh 等^[39]的临床转化策略相似, 但融入了更详细的机制性描述, 如 FcRn 介导的抗体循环。它同样描述了 ADC、抗体和小分子毒素在肿瘤内外的处置情况, 包括与肿瘤外部的目标结合。该模型被转化为人类, 并进行了虚拟临床试验模拟, 成功地预测了 T-DM1 和 T-DXd 在 HER2 阳性转移性乳腺癌治疗中的 PFS 反应, 包括基于 HER2 抗原表达差异的疗效区别。总的来说, 该模型是向 ADC 的平台 QSP 模型和策略迈进的一步, 它整合了多种类型的数据和知识来预测 ADC 的疗效。

这些研究提供了一个经过验证、可重复的临床转化策略 (图 4), 它不仅成功整合了临床前和临床数据, 还能通过模拟不同给药方案来预测临床试验的结果。这一策略为其他 ADC 的临床转化研究提供了重要参考。

2.3 评价 DDI DDI 是指在使用两种或两种以上药物时, 由于药物之间的相互作用或与机体的反应, 导致药物的效果、持续时间或性质发生不同程度的改变的现象。在考察 ADC 的 DDI 时需要考虑大分子抗体及小分子毒素药物的相互作用。虽然 ADC 的抗体部分与细胞色素 P450 同工酶 (CYP450) 的关联较小, 但其降解产物中的游离小分子毒素, 如 monomethyl auristatin E (MMAE) 可能会被 CYP 和转运蛋白代谢和排泄。研究表明, MMAE 既是 CYP3A 和 P-糖蛋白的底物, 也是 CYP3A 的抑制剂。尽管循环中未结合的 MMAE 水平较低, 但由于其具有高活性和低暴露量的特性, 其 DDIs 仍可能通过调节重要消除途径发生。因此, 评估与游离小分子毒素药物相关的 DDI 潜力是支持 ADC 临床开发的重要风险内容。Chen 等^[32]首次通

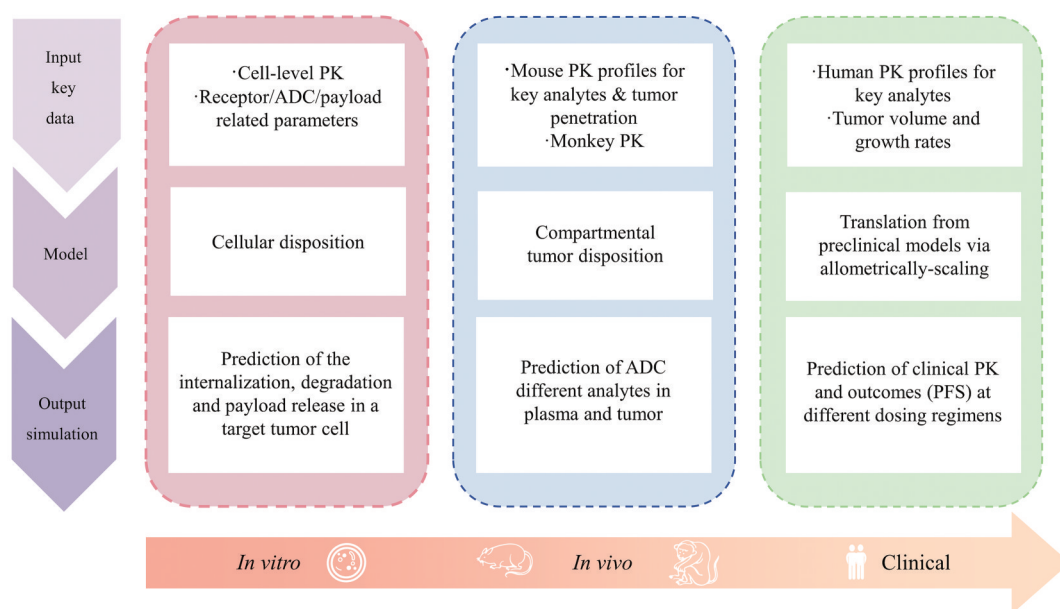


Figure 4 ADC clinical translational modeling and simulation strategy. PFS: Progression free survival

过PBPK模型对含有vc-MMAE的ADC进行DDI预测评估。该模型结合“自上而下”和“自下而上”两种建模方法,利用抗CD22-vc-MMAE的临床前和临床数据进行建模。在确认相关参数后,预测brentuximab vedotin的PK中MMAE的暴露量,并将其与brentuximab vedotin的临床试验PK中MMAE暴露量进行比较和验证。最后通过验证模型预测brentuximab vedotin与咪达唑仑(CYP3A底物)、酮康唑(CYP3A抑制剂)和利福平(CYP3A诱导剂)之间的DDI,其AUC及药物最大浓度结果均与临床DDI研究观测值接近。研究还表明,对于具有相同连接子(vc)及相同的小分子毒素(MMAE)的ADC,无论单克隆抗体部分的设计如何,其PK特征都相似。Samineni等^[53]进一步优化了上述PBPK模型,并成功将其应用于预测polatuzumab vedotin(CD79b-vc-MMAE)DDI的风险结果。此外,该模型的预测成果已成功获得监管部门的认可,它不仅能够有效替代传统的临床药物DDI试验,还为药物说明书的编写提供了科学的指导依据。

2.4 临床剂量探索及优化 ADC设计的初衷是将高活性小分子毒素药物与抗体结合,以降低药物毒性并扩大治疗窗口。然而,越来越多的临床数据显示,与传统药物相比,ADC的治疗窗仍相对较窄。许多ADC药物因毒性过大或风险/获益比过高而在临床开发过程中受阻。关键临床试验的剂量探索及优化是影响ADC研发成功与否的重要因素之一,若未进行充分的剂量探索,可能导致上市的剂量并非最佳剂量,无法为患者提供疗效最大化、安全性风险最小化的治疗,甚至可能导致治疗失败和药物退市^[68]。

Mylotarg (gemtuzumab ozogamicin, GO) 是全球首个经FDA批准上市用于治疗CD33阳性急性髓性白血病(acute myeloid leukemia, AML)的ADC药物。然而,该药物在验证性III期试验中却未能获得明显的临床效益,且出现了严重的肝损伤和高死亡率,导致研究提前终止研究并于2010年撤市。尽管GO退出了市场,但由于AML患者的结局持续不良,人们仍对其保持浓厚的研究兴趣。随后,使用不同给药方案(Mylo-France-1、ALFA-0701及AML-19)的临床试验结果发现,通过增加给药频率以较低剂量给药(分级剂量)可以显著提高GO疗效,同时降低毒性,但这些试验缺乏PK数据。Fostvedt等^[49]基于GO过去的8项临床研究数据,通过PK/PD模型进行分析,以支持这些新的给药方案的安全性和有效性。该研究通过成年患者数据进行Pop-PK建模,预测关键III临床试验(ALFA-0701)中GO总抗体的暴露量;随后通过模型描述预测的GO总抗体的暴露量与疗效及安全性之间的关系,并桥接外推至儿童患者;另外建立半机制模型预测不同治疗方案对血小板和粒细胞的抑制作用情况。模型模拟结果表明,与最初治疗方案相比(9 mg·m⁻²,第1和15天给药),新的给药方案中GO(3 mg·m⁻²,第1、4和7天给药)与阿糖胞苷和多柔比星联合化疗方案的临床效果最佳。根据以上结果,FDA于2017年重新批准了GO上市,用于治疗新诊断和复发的成人AML患者以及2~17岁儿童患者的复发AML。

T-DXd成为首个获得FDA批准用于治疗非小细胞肺癌(non-small cell lung cancer, NSCLC)的ADC药物,其剂量优化探索策略充分展现了MIDD的应用优

势。Yin 等^[57]的研究首次公布了 T-DXd 在 HER2 阳性转移性乳腺癌和其他实体瘤患者中的 Pop-PK 特征。这项研究涵盖了 5 个临床试验的患者数据 ($n = 679$), 探索了每三周一次 (Q3W) 给药方案在不同剂量水平 ($0.8 \sim 8.0 \text{ mg} \cdot \text{kg}^{-1}$) 下的情况。研究采用了隔室模型描述了人体血清中 T-DXd 和游离药物 Dxd 的变化, 并为后续的 E-R 分析估算了 PK 参数。此外, 评估相关协变量 (种族、年龄、肿瘤大小等) 对药物暴露的影响。研究结果表明, 除了体重和血清白蛋白水平有显著影响外, 其他协变量等对 T-DXd 和游离 Dxd 的暴露影响均不超过 20%。随后, 该团队^[58]通过两项临床试验患者数据 (DESTINY-Breast01 和 J101) 通过 E-R 分析进行随机剂量探索研究。分析结果表明, T-DXd 的 AUC 与临床 ORR 之间存在显著的统计关联, 同时安全性终点与 T-DXd 和游离 Dxd 的暴露显著相关。预测结果表明, 当 T-DXd 剂量从 $5.4 \text{ mg} \cdot \text{kg}^{-1}$ Q3W 增加至 $6.4 \text{ mg} \cdot \text{kg}^{-1}$ Q3W 时, ORR 和不良反应事件均有所增加。该研究通过比较不同剂量下的获益/风险比, 为探究 $5.4 \text{ mg} \cdot \text{kg}^{-1}$ Q3W 剂量方案的有效性和安全性提供了依据。这些成功的模型案例表明, 在 ADC 的临床试验中, 利用 M&S 进行临床剂量优化不仅有助于深入分析人体内暴露及安全有效性数据, 还能补充数据证据, 进而降低临床研究过程中的不确定性^[69,70]。

3 结语与展望

近年来, M&S 技术在定量理解 ADC 方面取得了快速发展, MIDD 为 ADC 的开发提供了独特的价值。本文讨论了不同模型案例在 ADC 研发阶段的不同应用, 旨在 MIDD 在 ADC 领域的应用提供有益的参考和启示。这些模型案例通过整合多元化的数据和知识, 对 ADC 的特性实现精准捕捉。同时, 模型在进行连续的验证和优化后, 能够在“假设”场景中生成新的数据, 从而有效应对传统实验方法的挑战, 为 ADC 开发周期内的各项决策提供了关键的支持和指导。

虽然目前已开发多种模型应用于 ADC 的研发关键环节, 但 MIDD 在 ADC 领域的应用仍面临一系列挑战。例如, 在早期研发阶段, 高质量数据往往较为缺乏, 这可能导致的模型在预测结果方面存在偏差; 肿瘤异质性和微环境的复杂性会对 ADC 疗效产生影响, 但当前建模工作尚未完全充分体现这些因素; ADC 的毒性建模需要更多的研究和数据支持, 这使得目前对 ADC 治疗指数的预测仍存在困难; ADC 在免疫肿瘤学治疗的建模和定量预测仍需进一步探索^[71]。此外, 随着 ADC 研发技术的不断进步, 下一代 ADC 可能会出现新的结构设计, 如多特异性抗体、非细胞毒性载荷和多载荷系统等, 现有模型需进行进一步发展和适应。

因此, 研究者可以通过利用 QSP、疾病进展等新型模型技术平台来持续优化和改进模型^[72-74], 从而提高其准确性和实用性。另一方面, 随着研究者们利用 M&S 软件持续进行深度学习和数据积累, 有望出现专门针对 ADC 药物研发的模型预测功能模块。这不仅极大地简化模型开发流程, 还优化了模型参数的调整过程, 为 ADC 研发提供更有益的支持。

现阶段, 一些国外知名制药企业, 如基因泰克和辉瑞, 已经率先将 M&S 工具融入 ADC 的研发过程中, 极大地提高了研发效率。我国目前在 ADC 研发方面的活动已占据全球超过半数比例, 在研发市场上展示出充沛的活力和巨大的潜力。如果能构建以模型为基础的 ADC 研发模式, 并在研发初期就开始运用 M&S 工具, 有望充分挖掘 MIDD 在 ADC 研发中的巨大潜力, 进一步提升研发效率并节省研发资源。

作者贡献: 吴白杨负责论文框架设计、撰写与资料收集; 王凌参与论文设计与修改; 姜静负责论文选题指导与资源支持。

利益冲突: 所有作者均声明无利益冲突。

References

- [1] Tarantino P, Carmagnani Pestana R, Corti C, et al. Antibody-drug conjugates: smart chemotherapy delivery across tumor histologies [J]. *CA Cancer J Clin*, 2022, 72: 165-182.
- [2] Li J, Wu YL, Ma CC, et al. Research progress on antibody-drug conjugates [J]. *West China J Pharm Sci* (华西药科学杂志), 2023, 38: 586-592.
- [3] Zheng QS. Model-informed drug development: principles and case studies [J]. *Chin J New Drugs* (中国新药杂志), 2023, 32: 1946-1952.
- [4] Li J, Yang JB, Wang YZ. Application of model-informed drug development in new drug research and development [J]. *Chin J Clin Pharmacol Ther* (中国临床药理学与治疗学), 2020, 25: 1-8.
- [5] Wei CM, Gao LL, He RR, et al. Model-informed methods support regulatory decision-making for innovative drugs in China [J]. *China Food Drug Adm Mag* (中国食品药品监管), 2024, 4: 14-19.
- [6] Haraya K, Tsutsui H, Komori Y, et al. Recent advances in translational pharmacokinetics and pharmacodynamics prediction of therapeutic antibodies using modeling and simulation [J]. *Pharmaceuticals* (Basel), 2022, 15: 508-539.
- [7] Singh AP, Shin YG, Shah DK. Application of pharmacokinetic-pharmacodynamic modeling and simulation for antibody-drug conjugate development [J]. *Pharm Res*, 2015, 32: 3508-3525.
- [8] Li C, Chen SC, Chen Y, et al. Impact of physiologically based pharmacokinetics, population pharmacokinetics and pharmacoki-

- netics/pharmacodynamics in the development of antibody-drug conjugates [J]. *J Clin Pharmacol*, 2020, 60: 105-119.
- [9] Hedrich WD, Fandy TE, Ashour HM, et al. Antibody-drug conjugates: pharmacokinetic/pharmacodynamic modeling, preclinical characterization, clinical studies, and lessons learned [J]. *Clin Pharmacokinet*, 2018, 57: 687-703.
- [10] Madabushi R, Seo P, Zhao L, et al. Review: role of model-informed drug development approaches in the lifecycle of drug development and regulatory decision-making [J]. *Pharm Res*, 2022, 39: 1669-1680.
- [11] Zhou Z, Liu B. Model-informed drug development: development history and application thinking [J]. *Chin J Clin Pharmacol (中国临床药理学杂志)*, 2021, 37: 772-776, 787.
- [12] Liu DY, Wang K, Ma GL, et al. The value of quantitative pharmacology research in new drug development and general considerations [J]. *Chin J Clin Pharmacol Ther (中国临床药理学与治疗学)*, 2018, 23: 961-973.
- [13] EFPIA MID3 Workgroup, Marshall SF, Burghaus R, et al. Good practices in model-informed drug discovery and development: practice, application, and documentation [J]. *CPT Pharmacometrics Syst Pharmacol*, 2016, 5: 93-122.
- [14] Sun ZY, Jia LR, Li DL, et al. Research progress and application of quantitative pharmacology [J]. *J Shenyang Pharm Univ (沈阳药科大学学报)*, 2018, 35: 431-436.
- [15] PDUFA reauthorization performance goals and procedures for fiscal years 2018 through 2022 [EB/OL]. Silver Spring, MD: U. S. Food & Drug Administration, 2022 [2024-07-26]. <https://www.fda.gov/media/99140/download>.
- [16] Model-informed drug development pilot program [EB/OL]. Silver Spring, MD: U. S. Food & Drug Administration, 2020 [2024-07-26]. <https://www.fda.gov/newsevents/pressreleases/ucm294657>.
- [17] Physiologically based pharmacokinetic analyses-format and content guidance for industry [EB/OL]. Silver Spring, MD: U. S. Food & Drug Administration, 2018 [2024-07-26]. <https://www.fda.gov/media/101469/download>.
- [18] Guideline on the reporting of physiologically based pharmacokinetic (PBPK) modelling and simulation [EB/OL]. Amsterdam: European Medicines Agency, 2018 [2024-07-26]. https://www.ema.europa.eu/en/documents/scientific-guideline/guideline-reporting-physiologically-based-pharmacokinetic-pbpbk-modelling-and-simulation_en.pdf.
- [19] Guideline of model-informed drug development [EB/OL]. Beijing: Center for Drug Evaluation, NMPA, 2020 [2024-07-26]. <https://www.cde.org.cn/main/news/viewInfoCommon/23b634adf79ecd4616bb91bcd66815f0>.
- [20] Guidelines for the exploration and optimization of innovative drug dosages guided by models (draft for comments) [EB/OL]. Beijing: Center for Drug Evaluation, NMPA, 2024 [2024-07-26]. <https://www.cde.org.cn/main/news/viewInfoCommon/efd1f4faac2a5cbecc147c2e98a76361>.
- [21] Dumontet C, Reichert JM, Senter PD, et al. Antibody-drug conjugates come of age in oncology [J]. *Nat Rev Drug Discov*, 2023, 22: 641-661.
- [22] Gao HY, Li XJ, Zhong SL, et al. Advances in pharmacokinetic research of monoclonal antibody drugs and antibody-drug conjugates [J]. *Pharm Clin Res (药学与临床研究)*, 2019, 27: 212-215.
- [23] Guo JJ, Gao R, Quan TF, et al. Advances in pharmacokinetic research of antibody-drug conjugates [J]. *Acta Pharm Sin (药学报)*, 2015, 50: 1203-1209.
- [24] Ji SM, Wang YZ, Yang JB. Molecular characteristics and pharmacokinetic research considerations of antibody-drug conjugates [J]. *Chin J Clin Pharmacol (中国临床药理学杂志)*, 2021, 37: 777-782.
- [25] Tao ZP, Xu BH, Zhou L, et al. Research progress on antibody-drug conjugates and their metabolism *in vivo* and *in vitro* [J]. *Drug Eval Res (药物评价研究)*, 2022, 45: 2574-2582.
- [26] Shah DK, Haddish BN, Betts A. Bench to bedside translation of antibody drug conjugates using a multiscale mechanistic PK/PD model: a case study with brentuximab-vedotin [J]. *J Pharmacokinet Pharmacodyn*, 2012, 39: 643-659.
- [27] Haddish BN, Shah DK, Ma D, et al. On translation of antibody drug conjugates efficacy from mouse experimental tumors to the clinic: a PK/PD approach [J]. *J Pharmacokinet Pharmacodyn*, 2013, 40: 557-571.
- [28] Shah DK, King LE, Han X, et al. A priori prediction of tumor payload concentrations: preclinical case study with an auristatin-based anti-5T4 antibody-drug conjugate [J]. *AAPS J*, 2014, 16: 452-463.
- [29] Bender B, Leipold DD, Xu K, et al. A mechanistic pharmacokinetic model elucidating the disposition of trastuzumab emtansine (T-DM1), an antibody-drug conjugate (ADC) for treatment of metastatic breast cancer [J]. *AAPS J*, 2014, 16: 994-1008.
- [30] Wada R, Erickson HK, Lewis Phillips GD, et al. Mechanistic pharmacokinetic/pharmacodynamic modeling of *in vivo* tumor uptake, catabolism, and tumor response of trastuzumab maytansinoid conjugates [J]. *Cancer Chemother Pharmacol*, 2014, 74: 969-980.
- [31] Vasalou C, Helmlinger G, Gomes B. A mechanistic tumor penetration model to guide antibody drug conjugate design [J]. *PLoS One*, 2015, 10: e0118977.
- [32] Chen Y, Samineni D, Mukadam S, et al. Physiologically based pharmacokinetic modeling as a tool to predict drug interactions for antibody-drug conjugates [J]. *Clin Pharmacokinet*, 2015, 54: 81-93.
- [33] Maass KF, Kulkarni C, Betts AM, et al. Determination of cellular processing rates for a trastuzumab-maytansinoid antibody-drug conjugate (ADC) highlights key parameters for ADC design [J]. *AAPS J*, 2016, 18: 635-646.

- [34] Singh AP, Maass KF, Betts AM, et al. Evolution of antibody-drug conjugate tumor disposition model to predict preclinical tumor pharmacokinetics of trastuzumab-emtansine (T-DM1) [J]. AAPS J, 2016, 18: 861-875.
- [35] Betts AM, Haddish BN, Tolsma J, et al. Preclinical to clinical translation of antibody-drug conjugates using PK/PD modeling: a retrospective analysis of inotuzumab ozogamicin [J]. AAPS J, 2016, 18: 1101-1116.
- [36] Cilliers C, Guo H, Liao J, et al. Multiscale modeling of antibody-drug conjugates: connecting tissue and cellular distribution to whole animal pharmacokinetics and potential implications for efficacy [J]. AAPS J, 2016, 18: 1117-1130.
- [37] Singh AP, Sharma S, Shah DK. Quantitative characterization of *in vitro* bystander effect of antibody-drug conjugates [J]. J Pharmacokinet Pharmacodyn, 2016, 43: 567-582.
- [38] Sukumaran S, Zhang C, Leipold DD, et al. Development and translational application of an integrated, mechanistic model of antibody-drug conjugate pharmacokinetics [J]. AAPS J, 2017, 19: 130-140.
- [39] Singh AP, Shah DK. Application of a PK-PD modeling and simulation-based strategy for clinical translation of antibody-drug conjugates: a case study with trastuzumab emtansine (T-DM1) [J]. AAPS J, 2017, 19: 1054-1070.
- [40] Ait-Oudhia S, Zhang W, Mager DE. A mechanism-based PK/PD model for hematological toxicities induced by antibody-drug conjugates [J]. AAPS J, 2017, 19: 1436-1448.
- [41] Li H, Han TH, Hunder NN, et al. Population pharmacokinetics of brentuximab vedotin in patients with CD30-expressing hematologic malignancies [J]. J Clin Pharmacol, 2017, 57: 1148-1158.
- [42] Singh AP, Shah DK. Measurement and mathematical characterization of cell-level pharmacokinetics of antibody-drug conjugates: a case study with trastuzumab-vc-MMAE [J]. Drug Metab Dispos, 2017, 45: 1120-1132.
- [43] Kågedal M, Gibiansky L, Xu J, et al. Platform model describing pharmacokinetic properties of vc-MMAE antibody-drug conjugates [J]. J Pharmacokinet Pharmacodyn, 2017, 44: 537-548.
- [44] Khot A, Tibbitts J, Rock D, et al. Development of a translational physiologically based pharmacokinetic model for antibody-drug conjugates: a case study with T-DM1 [J]. AAPS J, 2017, 19: 1715-1734.
- [45] Khera E, Cilliers C, Bhatnagar S, et al. Computational transport analysis of antibody-drug conjugate bystander effects and payload tumoral distribution: implications for therapy [J]. Mol Syst Des Eng, 2018, 3: 73-88.
- [46] Shah DK, Loganzo F, Haddish BN, et al. Establishing *in vitro-in vivo* correlation for antibody drug conjugate efficacy: a PK/PD modeling approach [J]. J Pharmacokinet Pharmacodyn, 2018, 45: 339-349.
- [47] Singh AP, Shah DK. A "dual" cell-level systems PK-PD model to characterize the bystander effect of ADC [J]. J Pharm Sci, 2019, 108: 2465-2475.
- [48] Singh AP, Guo L, Verma A, et al. A cell-level systems PK-PD model to characterize *in vivo* efficacy of ADCs [J]. Pharmaceutics, 2019, 11: 98-116.
- [49] Fostvedt LK, Hibma JE, Masters JC, et al. Pharmacokinetic/pharmacodynamic modeling to support the re-approval of gemtuzumab ozogamicin [J]. Clin Pharmacol Ther, 2019, 106: 1006-1017.
- [50] Singh AP, Guo L, Verma A, et al. Antibody coadministration as a strategy to overcome binding-site barrier for ADCs: a quantitative investigation [J]. AAPS J, 2020, 22: 28-41.
- [51] Menezes B, Cilliers C, Wessler T, et al. An agent-based systems pharmacology model of the antibody-drug conjugate kadcyla to predict efficacy of different dosing regimens [J]. AAPS J, 2020, 22: 29-42.
- [52] Sharma S, Li Z, Bussing D, et al. Evaluation of quantitative relationship between target expression and antibody-drug conjugate exposure inside cancer cells [J]. Drug Metab Dispos, 2020, 48: 368-377.
- [53] Samineni D, Ding H, Ma F, et al. Physiologically based pharmacokinetic model-informed drug development for polatuzumab vedotin: label for drug-drug interactions without dedicated clinical trials [J]. J Clin Pharmacol, 2020, 60: 120-131.
- [54] Suri A, Mould DR, Song G, et al. Population pharmacokinetics of brentuximab vedotin in adult and pediatric patients with relapsed/refractory hematologic malignancies: model-informed hypothesis generation for pediatric dosing regimens [J]. J Clin Pharmacol, 2020, 60: 1585-1597.
- [55] Singh AP, Seigel GM, Guo L, et al. Evolution of the systems PK-PD model for antibody-drug conjugates (ADC) to characterize tumor heterogeneity and *in vivo* bystander effect [J]. J Pharmacol Exp Ther, 2020, 374: 184-199.
- [56] Weddell J, Chiney MS, Bhatnagar S, et al. Mechanistic modeling of intra-tumor spatial distribution of antibody-drug conjugates: insights into dosing strategies in oncology [J]. Clin Transl Sci, 2021, 14: 395-404.
- [57] Yin O, Xiong Y, Endo S, et al. Population pharmacokinetics of trastuzumab deruxtecan in patients with HER2-positive breast cancer and other solid tumors [J]. Clin Pharmacol Ther, 2021, 109: 1314-1325.
- [58] Yin O, Iwata H, Lin CC, et al. Exposure-response relationships in patients with HER2-positive metastatic breast cancer and other solid tumors treated with trastuzumab deruxtecan [J]. Clin Pharmacol Ther, 2021, 110: 986-996.
- [59] Rathi C, Collins J, Struemper H, et al. Population pharmacokinetics of belantamab mafodotin, a BCMA-targeting agent in patients with relapsed/refractory multiple myeloma [J]. CPT Pharmacometrics Syst Pharmacol, 2021, 10: 851-863.
- [60] Menezes B, Linderman JJ, Thurber GM. Simulating the selection

- of resistant cells with bystander killing and antibody coadministration in heterogeneous human epidermal growth factor receptor 2-positive tumors [J]. *Drug Metab Dispos*, 2022, 50: 8-16.
- [61] Scheuher B, Ghusinga KR, McGirr K, et al. Towards a platform quantitative systems pharmacology (QSP) model for preclinical to clinical translation of antibody drug conjugates (ADCs) [J]. *J Pharmacokinet Pharmacodyn*, 2023, 10: 1-19.
- [62] Chang HP, Le HK, Shah DK. Pharmacokinetics and pharmacodynamics of antibody-drug conjugates administered *via* subcutaneous and intertumoral routes [J]. *Pharmaceutics*, 2023, 15: 1132-1162.
- [63] Chang HP, Cheung YK, Liu S, et al. Development of a generalized pharmacokinetic model to characterize clinical pharmacokinetics of monomethyl auristatin E-based antibody-drug conjugates [J]. *Br J Clin Pharmacol*, 2024, 90: 1637-1655.
- [64] Lu Y, Shimizu S, Sawamura R, et al. Population pharmacokinetics of patritumab deruxtecan in patients with solid tumors [J]. *J Clin Pharmacol*, 2023, 63: 77-90.
- [65] Vasalou C, Proia TA, Kazlauskas L, et al. Quantitative evaluation of trastuzumab deruxtecan pharmacokinetics and pharmacodynamics in mouse models of varying degrees of HER2 expression [J]. *CPT Pharmacometrics Syst Pharmacol*, 2024, 13: 994-1005.
- [66] Calopiz MC, Linderman JJ, Thurber GM. Optimizing solid tumor treatment with antibody-drug conjugates using agent-based modeling: considering the role of a carrier dose and payload class [J]. *Pharm Res*, 2024, 41: 1109-1120.
- [67] Lam I, Pilla Reddy V, Ball K, et al. Development of and insights from systems pharmacology models of antibody-drug conjugates [J]. *CPT Pharmacometrics Syst Pharmacol*, 2022, 11: 967-990.
- [68] Liao MZ, Lu D, Kågedal M, et al. Model-informed therapeutic dose optimization strategies for antibody-drug conjugates in oncology: what can we learn from US food and drug administration-approved antibody-drug conjugates? [J]. *Clin Pharmacol Ther*, 2021, 110: 1216-1230.
- [69] Li J, Wang J. Model-informed oncology drugs dose selection [J]. *Chin J Clin Pharmacol (中国临床药理学杂志)*, 2024, 40: 1693-1696.
- [70] Gerber HP, Sapra P, Loganzo F, et al. Combining antibody-drug conjugates and immune-mediated cancer therapy: what to expect? [J]. *Biochem Pharmacol*, 2016, 102: 1-6.
- [71] Gao LL, Wang YZ, Wang Y, et al. General considerations for clinical pharmacology of antitumor antibody-conjugated drugs: implications from FDA review cases [J]. *Chin J Clin Pharmacol Ther (中国临床药理学与治疗学)*, 2023, 28: 75-85.
- [72] Gong C, Ruiz-Martinez A, Kimko H, et al. A spatial quantitative systems pharmacology platform spQSP-IO for simulations of tumor-immune interactions and effects of checkpoint inhibitor immunotherapy [J]. *Cancers (Basel)*, 2021, 13: 3751-3784.
- [73] Zhao C, Li GL, Wang YN. Advances and applications of quantitative systems pharmacology modeling and virtual clinical trials in modern drug development [J]. *Acta Pharm Sin (药学报)*, 2023, 58: 3296-3310.
- [74] Lemaire V, Bassen D, Reed M, et al. From cold to hot: changing perceptions and future opportunities for quantitative systems pharmacology modeling in cancer immunotherapy [J]. *Clin Pharmacol Ther*, 2023, 113: 963-972.