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# Quinoline-based anti-MRSA agents: Current development, structure-activity relationships, and mechanisms

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## ABSTRACT

Methicillin-resistant *Staphylococcus aureus* (MRSA), the most common pathogen in hospital and community environments, can cause serious and even fatal infections. The antibiotics currently used for clinical treatment of MRSA have developed resistance, and there is an urgent need to develop new antimicrobials to treat infections caused by MRSA strains. Quinoline analogues play an important role in the development of antimicrobials. Herein, we discussed the current development of antibacterial activities of quinoline analogues, mainly for anti-MRSA activity, and their structure-activity relationships (SARs) from the perspective of using the quinoline nucleus to search for novel potential anti-MRSA candidates. Additionally, the mechanisms of some representative quinoline analogues against MRSA were clarified. Altogether, this review could provide further insights for the rational development of quinoline-based antibacterial drugs, especially against MRSA.

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## 1. Introduction

Antibiotics are primarily used to treat a variety of bacterial infections, but due to overuse and misuse of antibiotics, bacteria have become resistant to currently approved marketed drugs [1,2]. The spread of antimicrobial resistance (AMR) has made once treatable diseases deadly again, and bacterial infections have become a widespread global health risk [3–6]. *Staphylococcus aureus* (*S. aureus*) is a round, Gram-positive bacteria that causes a variety of diseases ranging from skin and soft tissue infections to severe pneumonia, sepsis, and meningitis. Currently, more than 60% of *S. aureus* isolates are methicillin-resistant *S. aureus* (MRSA) [7–12]. Methicillin resistance in *S. aureus* strains is caused by the *in vivo* gene encoding PBP2a, which has an ultra-low  $\beta$ -lactam affinity and is capable of cell wall synthesis at normally lethal  $\beta$ -lactam concentrations to complete the bacterial reproductive

growth, hence MRSA is also known as a superbug [13,14]. And as the last line of defense, vancomycin, MRSA has also developed resistance to it. Therefore, there is an urgent need to develop new antimicrobials with anti-MRSA potency [15,16].

Among the heterocyclic derivatives containing nitrogen atoms, quinolines are used in drug design and discovery due to their wide range of biological activities [17,18]. Quinolines have been reported to have anticancer [19], antimalarial [20], antihypertensive [21], anti-inflammatory and analgesic [22,23], antibacterial [24], anti-human immunodeficiency virus (HIV) [25], antitubercular [26], anti-cardiovascular activities [27], and inhibition of tyrosine kinase [28] properties. At present, the drugs such as ofloxacin, norfloxacin, ciprofloxacin, and sparfloxacin, which have been approved for marketing to treat various bacterial infections, all contain a quinoline backbone in their structures [29,30]. Although there are some reviews on the biological activity of quinolines, a review on the antibacterial activity of quinolines against MRSA has not been reported. Herein, we reviewed the structure-activity relationships (SARs) and different mechanisms of potential antibacterial effects of quinoline analogues (Fig. 1) against MRSA strains in recent two decades to provide insights into the development of new potent quinoline-based antimicrobial agents.

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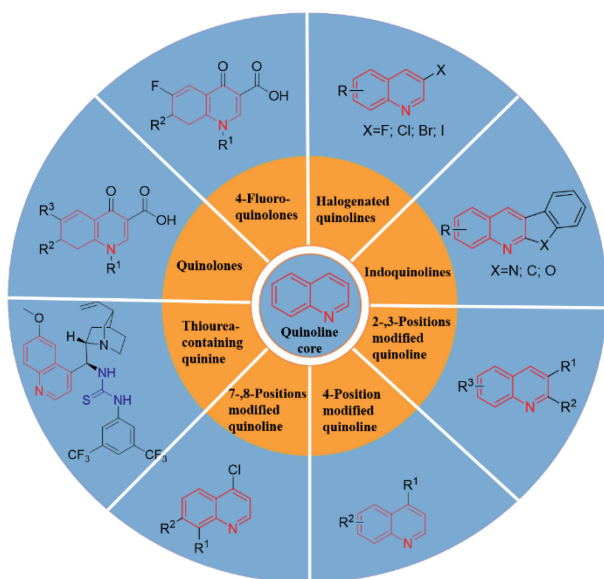


Fig. 1. Structures of quinoline analogues related to anti-MRSA activities.

## 2. Quinolones

4-Quinolones such as nemonoxacin [31,32] and delafloxacin [33] play a key role in the clinical anti-MRSA infection treatment. However, MRSA has developed resistance to 4-quinolones [34], so it is becoming increasingly urgent to improve the efficacy of 4-quinolones. Modifications of the C-7 position of 4-quinolones are thought to be beneficial for improving antibacterial potency, solubility, and safety [35,36].

HT-61 (**1**, Fig. 2), is a quinolone derivative with a molecular weight of about 400 Daltons that is particularly active against *S. aureus*, including MRSA [37,38]. Compound **1** not only acted on reproductive bacteria with minimum inhibitory concentrations (MICs) higher than those of marketed antibiotics but also displayed significant efficacy against non-reproductive bacteria. The MICs of **1** against 45 MRSA isolates were 4–8  $\mu\text{g/mL}$  and its minimum stationary phase-cidal concentration 50 (MSC<sub>50</sub>) values against 103 MRSA isolates were 1.5–7.5  $\mu\text{g/mL}$ . Compound **1** was significantly more potent than daptomycin, which required 50  $\mu\text{g/mL}$  to completely kill fixed-stage MRSA. Additionally, compound **1** can target bacterial cell membranes and act on the plasma membrane of bacterial cells by disrupting the membrane potential of *S. aureus*, resulting in the release of cellular contents. Moreover, compound **1** in combination with several antibiotics against clinical MRSA strains was also investigated, and the results showed that compound **1** had significant synergistic activity when it combined with gentamicin, neomycin, or chlorhexidine.

A few tricyclic fluoroquinolones, [1,2,4]triazolo[3,4-h][1,8]naphthyridine-8-one-7-carboxylic acid derivatives **2a–h** (Fig. 3), whose C-8 positions contained a functional Mannich base part, were evaluated for their *in vitro* antibacterial activities against MRSA [39]. The results showed that the Mannich base derived

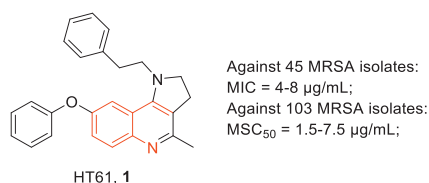


Fig. 2. Structure of compound **1** and its anti-MRSA activity.

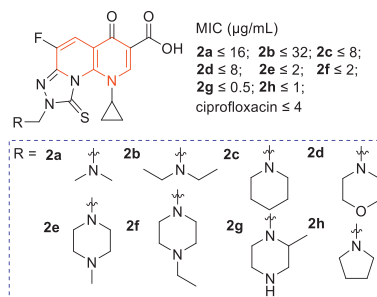


Fig. 3. Structures of compounds **2a–h** and their anti-MRSA activities.

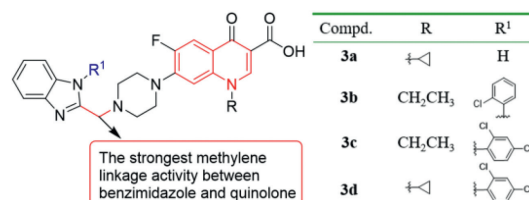


Fig. 4. Structures of compounds **3a–d**.

from the cycloaliphatic amine donors (**2d–h**) exhibited strong antibacterial activity against MRSA, with MIC values of 0.5–2  $\mu\text{g/mL}$ , and its potency was 2–8 times stronger than that of ciprofloxacin. Among them, especially compound **2g** had the best antibacterial activity with an MIC value of 0.5  $\mu\text{g/mL}$ , which was 8 times higher than that of ciprofloxacin (Fig. 3).

The new benzimidazole quinolones displayed promising antibacterial action [40]. Antibacterial evaluation *in vitro* showed that most of the title benzimidazole quinolones exhibited good antibacterial activity against the tested strains, especially against MRSA, even better than the reference drugs [40]. For example, methylene-bridge linkers, halogenated benzyl derivatives,  $\text{NH}_2\text{CH}_2$  linkers and benzimidazole complexes directly linked to quinolone nucleus displayed good anti-MRSA activity, with MIC values of 0.125–0.5  $\mu\text{g/mL}$ . They were more active than chloramphenicol (MIC = 16  $\mu\text{g/mL}$ ), norfloxacin (MIC = 8  $\mu\text{g/mL}$ ), ciprofloxacin (MIC = 2  $\mu\text{g/mL}$ ), and clinafloxacin (MIC = 1  $\mu\text{g/mL}$ ). The anti-MRSA activity of methylene-linked compounds **3a**, **3b**, **3c**, and **3d** (Fig. 4, MIC = 0.125  $\mu\text{g/mL}$ ) was 8–64 times higher than that of norfloxacin, ciprofloxacin, and clinafloxacin. Compound **3d** showed rapid bactericidal activity, and the number of viable bacteria decreased by more than 3 logs (CFU/mL) within an hour at a concentration of 6  $\times$  MIC. Moreover, **3d** not only can inhibit the formation of biofilm but also can disperse the formed bacterial biofilm, while exhibiting low toxicity to mammalian cells and not inducing bacterial resistance. The results of SARs indicated that when the benzimidazole and quinolone nucleus were linked by a methylene linkage, there was a significant effect on the enhancement of antibacterial activity (Fig. 4).

In 2015, some novel quinolone derivatives based on levofloxacin as the core were synthesized by Huang *et al.* [41]. The antibacterial activity of **4a–d** (Fig. 5, MIC = 1  $\mu\text{g/mL}$ ) was 4 times greater than

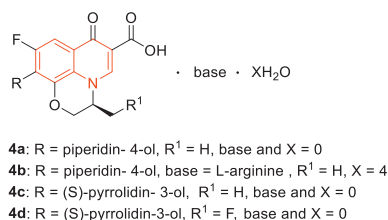
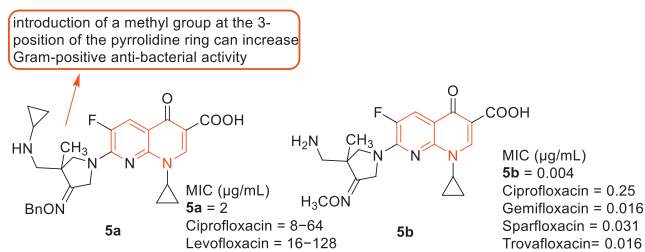
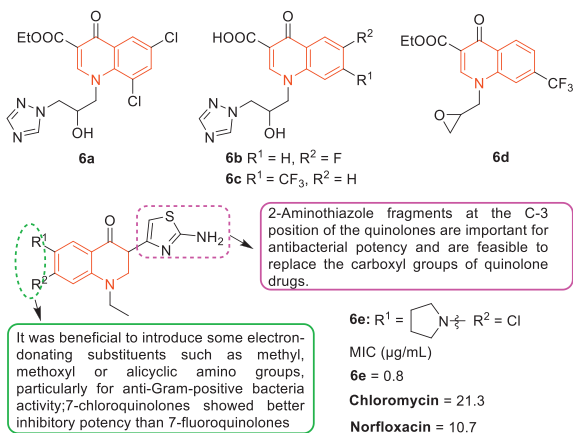


Fig. 5. Structures of compounds **4a–d**.



**Fig. 6.** Structures of compounds **5a**, **5b**, and their anti-MRSA activities.



**Fig. 7.** Structures of compounds **6a–e** and their SARs.

that of moxifloxacin (MIC = 4  $\mu\text{g/mL}$ ), 32 times stronger than that of levofloxacin (MIC = 32  $\mu\text{g/mL}$ ) and as isoenergetic as vancomycin (MIC = 1  $\mu\text{g/mL}$ ). Compound **4b** had better solubility (5.33 mg/mL) and lower toxicity than levofloxacin. In addition, the lethal dose 50 (LD<sub>50</sub>) of **4b** was 1402 mg/kg, while the LD<sub>50</sub> of levofloxacin was 358 mg/kg.

Zhang *et al.* [42] reported the development of fluoroquinolone derivatives containing 3-alkoxyimino-4-(cyclopropylamino)methylpyrrolidine moiety and evaluated their biological activities. Among these fluoroquinolone derivatives, compound **5a** (Fig. 6) exhibited the strongest antibacterial activity against MRSA with an MIC value of 2  $\mu\text{g/mL}$ , 4–64 times higher than ciprofloxacin and levofloxacin (MIC: 8–128  $\mu\text{g/mL}$ ) against MRSA. The SARs indicated that the introduction of methyl at 3-position of the pyrrolidine ring increased antibacterial activity against Gram-positive bacteria, which was consistent with the results reported by Yun *et al.* [43]. The compounds from the two research groups had the same parent nuclear structure, and compound **5b** synthesized by Yun *et al.* [43] displayed good *in vivo* efficacy against MRSA in a mouse infection model (Fig. 6).

Cui *et al.* [44] inserted the triazole ethanol moiety into the N-1 position of quinolones to change the different substituents on the benzene ring of quinolones, and prepared a new class of quinolones triazoles. Most of these new quinolone triazoles can effectively inhibit MRSA growth (MIC = 0.5–16  $\mu\text{g/mL}$ ), which were more effective than clinical drug chloramphenicol (MIC = 16  $\mu\text{g/mL}$ ). Compounds **6a**, **6b**, and **6c** (Fig. 7) with MIC values of 1  $\mu\text{g/mL}$  were 8 and 16 times more active than norfloxacin (MIC = 8  $\mu\text{g/mL}$ ) and chloramphenicol (MIC = 16  $\mu\text{g/mL}$ ), respectively. In particular, 7-trifluoromethyl intermediate **6d** (Fig. 7, MIC = 0.5  $\mu\text{g/mL}$ ), as a new potential anti-MRSA candidate, exhibited 16 and 32 times higher antibacterial activity than the standard drugs norfloxacin and chloramphenicol, respectively. In addition, Cui's [45] group also synthesized a series of novel 3-aminothiazole quinolones analogues as antibacterial agents. Among these 3-aminothiazol-quinolones, 3-(2-aminothiazol-4-yl)-

**Table 1**

Structures of compounds **7a**, **7b**, and their anti-MRSA activities.

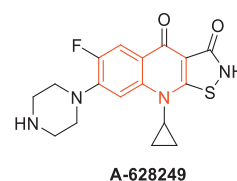
$\text{R}^1$  has methoxy substitution which is beneficial to increase antibacterial activity

Compd.	R <sup>1</sup>	R <sup>2</sup>	R <sup>3</sup>	MIC ( $\mu\text{g/mL}$ )
<b>7a</b>	-OMe	H		0.016
Ciprofloxacin	/	/	/	0.49
<b>7b</b>	-OMe	H		0.125
Sarafloxacin	/	/	/	0.25

7-chloro-6-(pyrrolidine-1-yl) quinolone **6e** (Fig. 7) showed significant antibacterial activity against MRSA at low concentrations (MIC = 0.8  $\mu\text{g/mL}$ ). Its antibacterial activity was 13 and 27 times higher than that of norfloxacin and chloramphenicol, respectively. Moreover, **6e** had low cytotoxicity to liver cells, strong inhibition of DNA cyclotron enzyme, and a wide antimicrobial spectrum, including MDR strains. Notably, the active molecule **6e** also induced bacterial resistance more slowly than norfloxacin. Analysis of the SARs (Fig. 7) revealed that the 2-aminothiazole fragment at C-3 position of quinolones increased antibacterial potency and could replace the carboxyl group of quinolones. The introduction of some electron-donating substituents, such as methyl, methoxy, or alicyclic amino groups, at 6- and 7-positions of quinolones was particularly beneficial for anti-Gram-positive bacteria. The antibacterial evaluation showed that 7-chloroquinolones had better inhibitory activity against MRSA than 7-fluoroquinolones. Additionally, the substituents on the benzene ring of quinolones also affected their antibacterial potency.

In 2020, Maryam *et al.* synthesized some new fluoroquinolones with ciprofloxacin and sarafloxacin as the core skeletons [46]. All the compounds synthesized from ciprofloxacin derivatives showed significant anti-MRSA activity, with MIC values ranging from 0.016  $\mu\text{g/mL}$  to 0.29  $\mu\text{g/mL}$ . The antibacterial activity of compound **7a** against MRSA (MIC = 0.016  $\mu\text{g/mL}$ ) was 60 times higher than that of the positive control ciprofloxacin (MIC = 0.49  $\mu\text{g/mL}$ ) (Table 1). This meant that the presence of methoxy at the R<sup>1</sup> position produced beneficial anti-MRSA activity. All the synthetic sarafloxacin derivatives exhibited approximately the same activity as the parent sarafloxacin against Gram-positive bacteria. Especially compound **7b** with a methoxy group at the R<sup>1</sup> position showed the most promising antibacterial activity against MRSA (MIC = 0.125  $\mu\text{g/mL}$ ), which was twice as high as the positive control sarafloxacin (MIC = 0.25  $\mu\text{g/mL}$ ) (Table 1).

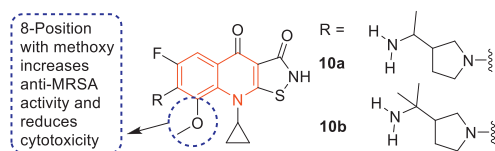
In 1987, isothiazoloquinolones (ITQs), originally reported by Chu and colleagues [47,48], were more potent than quinolones against *S. aureus*. The best represented of ITQs was the ciprofloxacin analogue **A-628249** (Fig. 8). Wiles and Wang *et al.* [49–52] synthesized ITQ-related compounds containing functionalized aryl and heteroaryl groups connected by a C–C bond at 7-position and



**Fig. 8.** Structure of compound **A-628249**.

**Table 2**  
Structures of compounds **8a**, **8b**, **9** and their anti-MRSA activities.

Compd.	Structure	MIC ( $\mu\text{g/mL}$ )
<b>8a</b>		0.125
<b>8b</b>		0.125
<b>9</b>		0.12

**Fig. 9.** Structures of compounds **10a** and **10b**.**Table 3**  
*In vitro* activities of compounds **10a** (*R,S*) and **10b** (*R*).

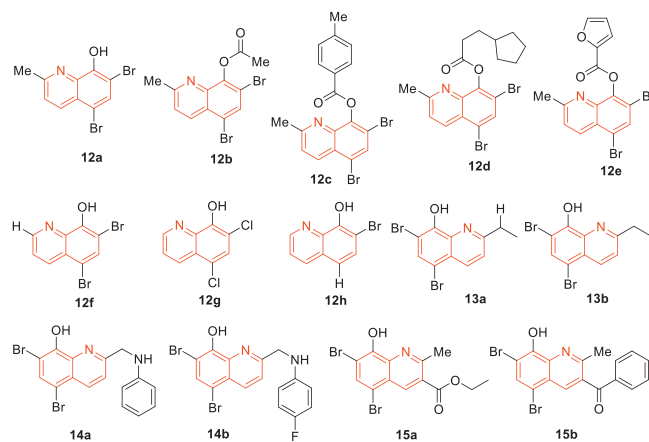
Compd.	Inhibition of <i>S. aureus</i> enzyme				MRSA, MIC ( $\mu\text{g/mL}$ )
	TopoIV WT	TopoIV Ser80Phe	DNA gyrase WT	DNA gyrase Ser84Leu	
CPFX <sup>a</sup>	3	57	62	>200	32
<b>10a</b> ( <i>R,S</i> )	0.04	0.47	0.68	3.2	0.06
<b>10b</b> ( <i>R</i> )	0.12	0.6	0.68	4.2	0.06

<sup>a</sup> Ciprofloxacin (CPFX).

structurally modified at 6-, 8-, and 9-positions, such as compounds **8a**, **8b**, and **9** (Table 2). These compounds exhibited excellent *in vitro* antibacterial activity against MRSA as well as strong inhibitory activity against DNA rotamases and topoisomerase IV. In 2011, a series of 8-methoxy ITQs with a 7-aminocyclic ring was synthesized by Kim *et al.* [53]. The structures of **10a** (*R,S*) and **10b** (*R*) are shown in Fig. 9. The MICs of **10a** and **10b** against MRSA and their inhibitory activities against target enzymes from *S. aureus*, wild-type topoisomerase IV and DNA rotamase, and the corresponding enzymes from *Staphylococcal* mutants expressing fluoroquinolone resistance are outlined in Table 3. Compounds **10a** (*R,S*) and **10b** (*R*) exhibited potent anti-MRSA activity with an MIC value of 0.06  $\mu\text{g/mL}$ . Notably, compared to ciprofloxacin, **10b** (*R*) increased inhibition of wild-type DNA gyrase activity by 91-fold, while **10a** (*R,S*) increased inhibition of wild-type topoisomerase IV activity by 75-fold. Similar improvements were observed for **10b** (*R*) and **10a** (*R,S*) inhibition of mutant DNA gyrase, with >48-fold and >62-fold increases, respectively. Compounds **10b** (*R*) and **10a** (*R,S*) increased the inhibition of mutant topoisomerase IV by 95-fold and 121-fold, respectively. The above data suggested that 8-position with methoxy could increase anti-MRSA activity and reduce cytotoxicity.

### 3. Halogenated quinolines

In 2010, O'Donnell *et al.* [54] performed an antimicrobial evaluation of haloquinoline derivatives, in which 4-hydroxy-3-iodoquinolin-2-one (**11**) showed strong to excellent antibacterial activity against eight clinical MRSA isolates (Table S1 in Supporting information). For clinical isolates EMRSA-16, CWMRSA, and

**Fig. 10.** Structures of the halogenated quinolines **12–15**.

HMRSA, the MIC value of **11** was equal to or lower than that of vancomycin, an antibiotic used in the clinical treatment of MRSA infection. While the antibacterial activity of **11** against other clinical MRSA isolates was better than vancomycin (MIC = 0.78–1.56  $\mu\text{g/mL}$ ).

In 2014, Abouelhasan's group [55] synthesized a library of 21 quinoline analogues and evaluated their antimicrobial activities (Fig. 10). Among these 21 quinoline analogues, some quinoline analogues showed effective activity against MRSA-2. The quinoline derivative **12a–e** (Fig. 10) exhibited very effective biofilm dispersion against MRSA-2 ( $\text{EC}_{50}$  = 2.80  $\mu\text{mol/L}$ ). In particular, quinoline derivative **12c** ( $\text{EC}_{50}$  = 2.06  $\mu\text{mol/L}$ ) exhibited the most effective biofilm dispersion activity. The 2-position of the halogenated quinoline (HQ) scaffold played an important role in the corresponding antibacterial activity of the HQ analogues. Compared with quinoline derivative **12a**, bromoxyquinoline **12f** (Fig. 10) had a hydrogen atom at 2-position, and **12a** had a methyl group at 2-position of the HQ scaffold (Fig. 10). This monomethyl difference resulted in a 16-fold increase in the antibacterial activity of **12a** against *Staphylococcal* pathogens and a 128-fold elimination of antibacterial activity against the Gram-negative pathogens *Acinetobacter baumannii*. In addition, **12a** can also eradicate MRSA biofilms. Based on the potent anti-MRSA activity, they subsequently synthesized a series of HQ analogues, which had different alkylation groups at the 2-position of the HQ scaffold [56]. In addition to 2-alkylated HQ analogues, they also designed an alternative pathway to synthesize reductive amination HQ analogues by reducing amination at 2-position of the HQ scaffold with various amines and anilines [56]. Some HQ analogues showed effective antibacterial activity, such as **13a** and **13b** (Fig. 10). It is worth mentioning that the MIC values of **14a** (Fig. 10) and **12a** (MIC  $\leq$  0.78  $\mu\text{mol/L}$ ) against MRSA-2 were identical. Among these HQ analogues, **14b** (Fig. 10) exhibited the highest anti-MRSA activity with MIC of 0.39  $\mu\text{g/mL}$ , and it did not show hemolysis toward red blood cells (RBC) at 200  $\mu\text{mol/L}$ . In general, HQ analogues containing aniline halide groups displayed good to highly potent antibacterial activity, while HQ analogues containing alkyl and methoxyaniline groups usually showed low antibacterial activity. For example, **13b** [minimum biofilm eliminating concentration (MBEC) = 93.8  $\mu\text{mol/L}$ ] and **14b** (MBEC = 93.8  $\mu\text{mol/L}$ ) displayed improved biofilm eradication activity against MRSA-2 compared with **12a** (MBEC = 188  $\mu\text{mol/L}$ ). These synthesized HQ analogues were 20 times more potent as MRSA-2 biofilm eradicators than current anti-MRSA therapeutic agents including vancomycin, daptomycin, and linezolid. Moreover, they displayed low hemolysis and low cytotoxicity to HeLa cells [56]. Eighty percent of bacterial infections are related to biofilm, and halogenated 8-

hydroxyquinolines had the characteristics of biofilm degradation. Accordingly, they used Friedlander reactions to synthesize a series of halogenated quinolines, capable of eradicating bacterial biofilms while exhibiting minimal mammalian cytotoxicity and hemolytic activity [56]. In preliminary MIC assays, some HQ compounds exhibited higher antimicrobial activity compared to **12a**. HQ compounds **15a** and **15b** (Fig. 10) showed improved activity against MRSA-2, with MIC of 0.39 and 0.59  $\mu\text{mol/L}$ , respectively, and **15b** proved to be one of the most effective biofilm eradicators ever reported against MRSA (MBEC = 3.9–23.5  $\mu\text{mol/L}$ ) [57–59]. Antimicrobial enhancement is defined as the ability of a non-growth inhibitory enhancer [the plant-derived chemical gallic acid (GA)] to reduce the MIC value of HQ by more than 4-fold. It is proven that combination therapy can improve antibacterial activity. Abouelhasan *et al.* [60] also evaluated enhancement assays in combination with GA against four clinical MRSA (MRSA-ATCC, MRSA-1, MRSA-2, and SA-156). Compounds **12f–h** (Fig. 10) had a 2–1000-fold enhancement effect when combined with GA. When **12f** was combined with GA, its activity was increased by 1000-fold potentiation against MRSA SA-156. They also studied the combination of GA with conventional antibacterial agents, such as ciprofloxacin and vancomycin. However, none of these antibacterial agents were enhanced by GA. These results indicated that the unique antibacterial mechanism of the HQ compounds was different from that of traditional antibiotics. HQ compounds operated through a metal (II) dependent mechanism and unlike conventional antibiotics, exhibited good cytotoxicity when tested in HeLa cells in lactate dehydrogenase (LDH) release assay. In summary, Abouelhasan *et al.* [60] found that the selective combination of GA and HQ small molecules had strong antibacterial activity against a variety of pathogens, including multidrug-resistant clinical isolates. The combination therapy has also been effective in eradicating MRSA biofilms. The phytochemical-HQ combination provided a promising new platform for the development of clinically useful antibacterial combination therapy [60].

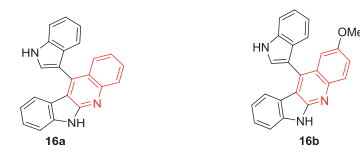
#### 4. Indoquinolines

The indole part is an important basic subunit for a large number of natural products and drugs [61], and indole-containing molecules have been reported to possess some antibacterial activity [62,63]. Moreover, indoquinoline analogues also play an important role in antimicrobial activity.

Under 2,3-dichloro-5,6-dicyano-1,4-benzoquinone (DDQ)-mediated oxidation conditions, Challa *et al.* [64] prepared a library of indolo[2,3-*b*]quinolines, chromeno[2,3-*b*]indoles, and 3-enyloctadiols using readily available 3,3'-diindolylmethane (DIM) (the synthetic route of the representative compounds **16a** and **16b** was shown in Scheme S1 in Supporting information). The MIC and minimum bactericidal concentration (MBC) of indoloquinolines against four different methicillin-resistant *S. aureus* strains (three of which were clinical isolates) were determined. The results showed that **16a** and **16b** possessed excellent activity against methicillin-sensitive *S. aureus* and MRSA (Table 4). Methicillin was active against susceptible strains, and the MIC value was 1  $\mu\text{g/mL}$ , but it had MIC values of 32  $\mu\text{g/mL}$  to >64  $\mu\text{g/mL}$  for all MRSA strains. This result indicated that the MRSA strain was highly resistant to methicillin. In contrast, compounds **16a** and **16b** showed similar activity against both drug-sensitive and drug-resistant strains, while the MIC values of these two compounds against methicillin-sensitive strains were 1  $\mu\text{g/mL}$ . For MRSA strains, the MIC values of **16a** and **16b** ranged from 1  $\mu\text{g/mL}$  to 4  $\mu\text{g/mL}$ , and the MBC values of **16a** and **16b** ranged from 2  $\mu\text{g/mL}$  to 8  $\mu\text{g/mL}$ , while methicillin had an MIC value of >64  $\mu\text{g/mL}$  in all tested MRSA strains (Table 4). The results manifested that compounds **16a** and **16b** were selective anti-MRSA agents, and their activities

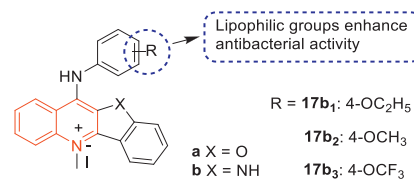
**Table 4**

Structures of compounds **16a**, **16b**, and their anti-MRSA activities.



Bacteria	MIC ( $\mu\text{g/mL}$ )			MBC ( $\mu\text{g/mL}$ )		
	<b>16a</b>	<b>16b</b>	MET <sup>a</sup>	<b>16a</b>	<b>16b</b>	MET
MRSA-ATCC33591	2	2	64	4	4	>64
MRSA-R3545	2	1	32	4	2	>64
MRSA-R3889	2	1	32	8	4	>64
MRSA-R3890	2	1	64	8	8	>64

<sup>a</sup> Methicillin (MET).



**Fig. 11.** Structures of compounds **a**, **b**, **17b<sub>1</sub>**, **17b<sub>2</sub>**, and **17b<sub>3</sub>**.

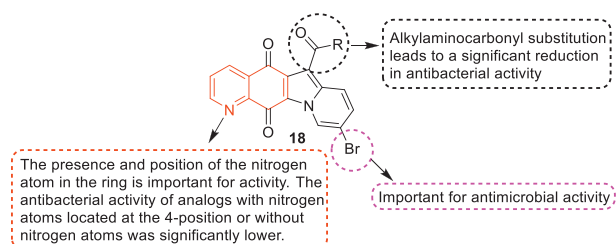
against MRSA were better than that against other bacteria. At the concentration of 16  $\mu\text{g/mL}$ , both compounds **16a** and **16b** completely killed MRSA within 360 min.

In 2017, the antibacterial activity and mode of action of some *N*-methylbenzofurano[3,2-*b*]quinolones and *N*-methylbenzoindolo[3,2-*b*]quinoline derivatives containing quaternary pyridine centers were studied by Sun *et al.* [65]. The results showed that all compounds had good antibacterial activity compared to the 11 clinical antibiotics evaluated in this study. In general, the antibacterial activity of 11-aniline-substituted 5-*N*-methyl-10-*H*-indole[3,2-*b*]quinolinium derivatives (b series) was slightly higher than that of 11-aniline-substituted 5-*n*-methylbenzofuran[3,2-*b*]quinoline derivatives (a series) (Fig. 11). This result may be due to the interaction between the imine in the quaternary pyridine center and the amino acids in the filamentous temperature-sensitive protein Z (FtsZ). In addition, the addition of lipophilic groups at the 4-position of 11-aniline slightly enhanced the antibacterial activity of these *N*-methyl quaternary ammonium derivatives. Among these derivatives, **17b<sub>1</sub>**, **17b<sub>2</sub>**, and **17b<sub>3</sub>** (Fig. 11) were the most effective against MRSA, with MIC values of 2  $\mu\text{g/mL}$ , and their antibacterial activities were about 100 times higher than that of berberine and methicillin. In the guanosine triphosphatase (GTPase) assay, **17b<sub>2</sub>** inhibited the GTPase activity of FtsZ in a dose-dependent manner. The binding of the quinoline derivatives to the C-terminal domain gap interfered with the GTPase activity of FtsZ, and then disrupted the formation of FtsZ polymer, leading to abnormal cell division and cell death.

The ester analogues in 9-bromo-substituted indolylquinoline-5,12-dione derivatives showed strong antibacterial activity against Gram-positive strains, especially against MRSA [66]. The MIC values of compounds **18a**, **18b**, and **18c** against MRSA were as high as 0.063  $\mu\text{g/mL}$ , which was 16 times higher than that of vancomycin. Compound **18d** exhibited the best activity with an MIC value of 0.031  $\mu\text{g/mL}$ , which was 32 times higher than vancomycin (Table 5). Compound **18a** had a strong inhibitory effect on the superhelix activity of DNA rotation of *E. coli* and the relaxation activity of topoisomerase IV of *S. aureus*, exhibiting a mechanism similar to ciprofloxacin [67,68]. Although compound **18a** displayed good anti-MRSA activity *in vitro*, its bioavailability in mouse models was very low, probably because of its low water solubility.

**Table 5**  
Structure of compounds **18a–d** and their anti-MRSA activities.

Compd.	R	MIC ( $\mu\text{g/mL}$ )
<b>18a</b>		0.063
<b>18b</b>		0.031
<b>18c</b>		0.063
<b>18d</b>		0.063
Vancomycin		1

**Fig. 12.** Summary of SARs study of compound **18**.

SARs (Fig. 12) studies have revealed that: (1) The substitution of 9-position with bromine can enhance the antibacterial activity, but the substitution of 9-position with chlorine shows a lower activity [69]. (2) Although 7-fluorinated analogues exhibited strong antibacterial activity with the same MIC value as compound **18a**, it had high cytotoxicity, indicating that it may not be a good choice for further development of good antibacterial agents [70]. (3) The substitution of alkyl, amino, and carbonyl groups at the 6-position resulted in a significant decrease in antibacterial activity, which may be attributed to their reduced solubility. (4) The presence and position of nitrogen atoms in the ring were important for activity. The antibacterial activity of analogues with nitrogen atoms at 4-position or without nitrogen atoms decreased significantly. On the other hand, substitution of the 6-position by the alkoxy-carbonyl group was important for enhancing the antibacterial activity. The modification of compound **18a** was guided by SARs and 28 indolylquinoline-5,12-diketone derivatives were synthesized to obtain the derivatives with higher anti-MRSA activity and good water solubility. Compound **18a<sub>1</sub>** exhibited strong activity against clinical MRSA strains, with MIC<sub>50</sub> and MIC<sub>90</sub> values lower than 0.0078  $\mu\text{g/mL}$ . Compound **18a<sub>2</sub>** was soluble in water with a solubility of 1.98 mg/mL and also displayed strong activity against clinical MRSA strains with an MIC<sub>50</sub> value of 0.063  $\mu\text{g/mL}$  and an MIC<sub>90</sub> value of 0.125  $\mu\text{g/mL}$ , which was 16 times higher than vancomycin (Table S2, Supporting information), suggesting that it could be further developed as an anti-MRSA lead compound.

In 1990, compound **19** was first synthesized by Yamato *et al.* [71] for the study of anti-tumor activity (Table 6). Zhao *et al.* [72–75] believed that compound **19**, containing the new skeleton of indoquinoline and its analogues, might have antibacterial activity and then measured the anti-MRSA activity of compound **19** against MRSA strains OM481 and OM584. The results showed that compound **19** had no activity against MRSA. Subsequently,

**Table 6**  
Structures of indoquinoline analogues and their anti-MRSA activities.<sup>a</sup>

Compd.	X	R <sup>1</sup>	R <sup>2</sup>	MIC ( $\mu\text{g/mL}$ )	
				OM481	OM584
<b>20</b>	/	/	/	8	16
<b>20a<sub>1</sub></b>	N	H	H	4	2
<b>20a<sub>2</sub></b>	N	MeO	H	2	2
<b>20a<sub>3</sub></b>	CH	H	Me	16	4
<b>20a<sub>4</sub></b>	CH <sub>2</sub>	H	/	>250	>250
<b>20a<sub>5</sub></b>	O	H	/	2	2
<b>21</b>	/	/	/	>250	>250
Vancomycin	/	/	/	2	1

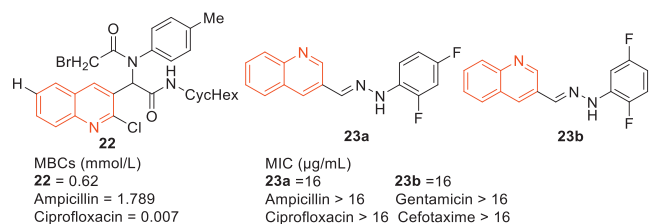
<sup>a</sup> Reproduced with permission [76]. Copyright 2015, Elsevier publications.

they determined the anti-MRSA activity of analogues of compound **19** and found that compound **20** had an anti-MRSA activity with MIC of 8  $\mu\text{g/mL}$  (against MRSA OM481) and 16  $\mu\text{g/mL}$  (against MRSA OM584). Afterwards, Zhao *et al.* [76] synthesized indolo[3,2-*b*]quinoline analogues, 4-(acridine-9-amino)phenol hydrochloride (**21**), benzofurano[3,2-*b*]quinoline, and indolo[1,2-*b*]quinoline based on the indolo[3,2-*b*]quinoline backbone of lead compound **20** and investigated their anti-MRSA activities against *s*-type OM481 and OM584 strains. The results showed that anti-MRSA activity of indoquinoline tetracyclic compounds against MRSA OM481 and OM584 strains was superior to that of tricyclic compound 4-(acridine 9-amino)phenol hydrochloride. The positions and types of substituents in indoquinoline analogues were important for the anti-MRSA activity against both strains. The indoquinoline analogues **20a<sub>1</sub>**, which introduced a hydroxyl group at the 4'-position showed obvious anti-MRSA activity against OM481 (MIC=4  $\mu\text{g/mL}$ ) and OM584 (MIC=2  $\mu\text{g/mL}$ ), revealing that 4'-hydroxyl group was important for the anti-MRSA activity towards both MRSA OM481 and OM584 (Table 6). The introduction of substituents at 7-position of indoquinoline analogues also exhibited good anti-MRSA activity, especially methoxy (**20a<sub>2</sub>**) exhibited obvious anti-MRSA activity against both OM481 and OM584 with MIC of 2  $\mu\text{g/mL}$  (Table 6). The anti-MRSA activity of indoquinoline (**20a<sub>3</sub>**) and methylindoquinoline (**20a<sub>4</sub>**) was weak, but benzofuran-quinoline (**20a<sub>5</sub>**) displayed significant anti-MRSA activity against both strains, with an MIC of 2  $\mu\text{g/mL}$  (Table 6). The SARs indicated that the indoquinoline ring, benzofuranquinoline ring, and 4-aminophenol group were the basic structures for the anti-MRSA activity.

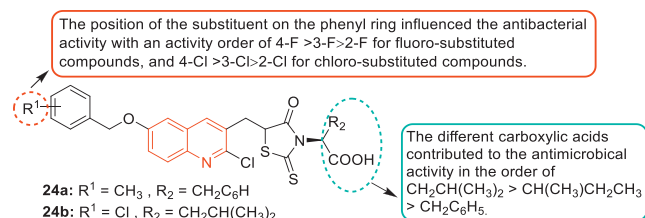
## 5. 2-,3-Positions modified quinoline derivatives

In 2016, Shiri *et al.* [77] reported a series of diamide derivatives containing 2-chloroquinoline scaffolds by Ugi condensation reaction of 2-chloroquinoline-3-carboxaldehyde amines, carboxylic acids, and isonitriles. Among them, compound **22** exhibited the strongest antibacterial activity against MRSA with an MBC value of 0.62 mmol/L, which was stronger than ampicillin but lower than ciprofloxacin (Fig. 13).

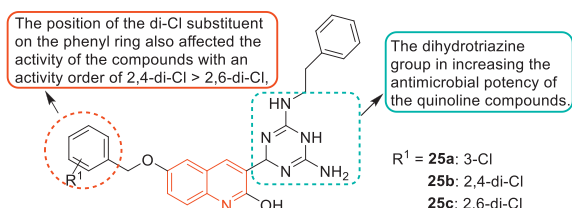
Compounds **23a** and **23b** are new compounds synthesized based on quinoline-3-formaldehyde hydrazone compounds (MIC: 16  $\mu\text{g/mL}$ ), which have a significant antibacterial effect against MRSA compared with ampicillin, gentamicin, ciprofloxacin, and cefotaxime (MIC: >16  $\mu\text{g/mL}$ ) (Fig. 13). According to the results of MTT test, compounds **23a** and **23b** had no significant effect on McF-7 cells at 100  $\mu\text{mol/L}$  concentration [78].



**Fig. 13.** Structure of compounds **22**, **23a**, **23b** and their anti-MRSA activities.



**Fig. 14.** Structures of compounds **24a** and **24b** and their SARs.



**Fig. 15.** Structures of compounds **25a-c** and SARs.

In 2013, Guo *et al.* prepared a series of rhodanine derivatives containing the quinoline fraction and evaluated their *in vitro* antibacterial activities [79]. The *in vitro* antibacterial assay showed that most of the rhodanine derivatives exhibited excellent antibacterial activity against different MDR Gram-positive bacteria (MIC value was 1–2  $\mu\text{g/mL}$ ). In particular, compounds **24a** and **24b** (Fig. 14) displayed the strongest level of inhibitory activity (MIC = 1  $\mu\text{g/mL}$ ), with a 4–8-fold increase in antibacterial activity compared with the standard drug norfloxacin (MIC = 4–8  $\mu\text{g/mL}$ ). A comparison of carboxylic acid derivatives at the *N*-position of the rhodanine ring indicated that the contribution order of different carboxylic acids to antibacterial activity was  $-\text{CH}_2\text{CH}(\text{CH}_3)_2 > -\text{CH}(\text{CH}_3)\text{CH}_2\text{CH}_3 > -\text{CH}_2\text{C}_6\text{H}_5$ . In addition, the position of substituents on the benzene ring affected the antibacterial activity, and the sequence of antibacterial activity of fluorine-substituted compounds was 4-F > 3-F > 2-F, and the sequence of antibacterial activity of chlorine-substituted compounds was 4-Cl > 3-Cl > 2-Cl. Compound **24a** did not affect the cell viability of human cervical (HeLa) cells when the concentration was equal to the MIC value but showed cytotoxicity at higher concentrations.

In 2019, Bai *et al.* [80] designed and synthesized some dihydrotriazine derivatives containing the quinoline fraction as novel antimicrobial agents. Dihydrotriazine derivatives **25a-c** (Fig. 15) were the most active compounds against MRSA3506 with an MIC of 1  $\mu\text{g/mL}$ . Based on the activity analysis of these dihydrotriazine derivatives, the following SARs were obtained. The dihydrotriazine group can improve the antibacterial activity of the quinoline compounds. The introduction of phenylethyl at the *N*-position of the dihydrotriazine ring resulted in significant differences in anti-MRSA activity, suggesting that the substituents of aromatic nuclei were critical for the activity of these dihydrotriazine compounds. In addition, the position of the 2-Cl substituent on the phenyl ring also affected the anti-MRSA activity. The 2,4-di-Cl sub-

**Table 7**  
 Structure of compound **26** and its anti-MRSA activity.

Compd.	MRSA, MIC/IC <sub>90</sub> ( $\mu\text{mol/L}$ )	
	24 h	48 h
<b>26</b>	3.9	7.81
Bacitracin	15.62	31.62
Penicillin V	125	125
Ciprofloxacin	500	500

stituted phenyl ring played a key role in the activity of these dihydrotriazine compounds, and its activity was greater than that of 2,6-Cl substituted compounds. These results were consistent with the previously reported results of a series of rhodanine and dihydrotriazine derivatives [81,11]. *In vitro* enzyme studies showed that compound **25a** exhibited inhibition of dihydrofolate reductase (DHFR), and compound **25a** did not show significant cytotoxic activity (HCT116 and LO2 cells IC<sub>50</sub> > 100  $\mu\text{mol/L}$ ). These findings suggested that these dihydrotriazine derivatives containing the quinoline fraction could be potential lead compounds for rational development of novel quinoline-based antibacterial agents.

The antimicrobial screening of novel ring-substituted styrylquinolines and two oxorhenium complexes compounds was carried out by Cieslik *et al.* [82]. Compound **26** (Table 7) was the most effective anti-MRSA candidate, with MIC/IC<sub>90</sub> values of 3.9 and 7.81  $\mu\text{mol/L}$  at 24 and 48 h, respectively, which were more potent than the standard drugs bacitracin, penicillin V, and ciprofloxacin.

## 6. 4-Position modified quinoline derivatives

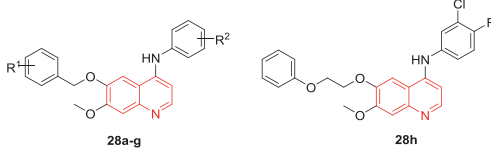
In 2018, Fang *et al.* [83] reported that 1-methylquinolinium iodide derivative (**27**) had good antibacterial activity against bacterial strains, and it had a synergistic effect in combination with  $\beta$ -lactam antibiotics against antibiotic-resistant strains of *S. aureus*. The antibacterial activity of **27** (Table 8, MIC = 1.5  $\mu\text{g/mL}$ ) against MRSA strains was 100-fold higher than that of methicillin and berberine. Compound **27** at 0.375  $\mu\text{g/mL}$  significantly increased the antibacterial activity of methicillin against MRSA and decreased the MIC value from 1024  $\mu\text{g/mL}$  to 32  $\mu\text{g/mL}$ . The fractional inhibitory

**Table 8**  
 MICs of **27** combined with  $\beta$ -lactam antibiotics against drug-resistant *S. aureus*.<sup>a</sup>

Compd.	MIC (single compound, $\mu\text{g/mL}$ )	MIC (checkerboard assay, $\mu\text{g/mL}$ )	FICI
<b>27</b>	1.5	0.375	0.5
Ampicillin	48	12	
<b>27</b>	1.5	0.375	0.281
Methicillin	1024	32	
<b>27</b>	1.5	0.375	0.375
Oxacillin	256	32	
<b>27</b>	1.5	0.75	0.75
Imipenem	24	6	
<b>27</b>	1.5	0.75	0.75
Ceftazidime	96	24	

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**Table 9**  
Structures of compounds **28a–h** and their anti-MRSA activities.

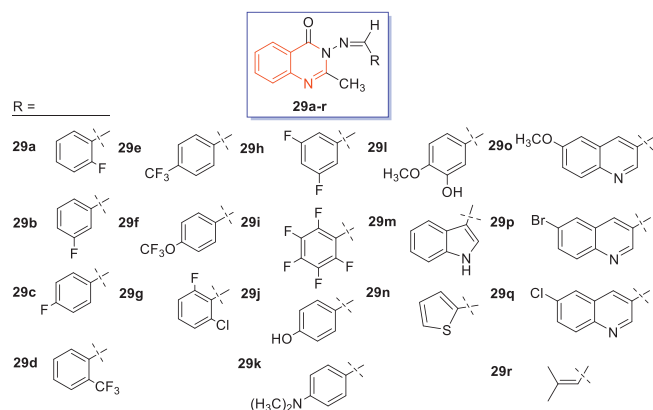


Compd.	R <sup>1</sup>	R <sup>2</sup>	MIC (μg/mL)
<b>28a</b>	4-CH <sub>3</sub>	4-CH <sub>3</sub>	12
<b>28b</b>	4-CF <sub>3</sub>	4-CH <sub>3</sub>	3
<b>28c</b>	4-CF <sub>3</sub>	2-CF <sub>3</sub>	3
<b>28d</b>	4-CF <sub>3</sub>	4-CF <sub>3</sub>	0.75
<b>28e</b>	4-CF <sub>3</sub>	3-Cl,4-F	1.5
<b>28f</b>	4-C(CH <sub>3</sub> )	3-Cl,4-F	1.5
<b>28g</b>	2-CH <sub>3</sub>	3-Cl,4-F	1.5
<b>28h</b>	/	/	3
Daptomycin	/	/	0.5

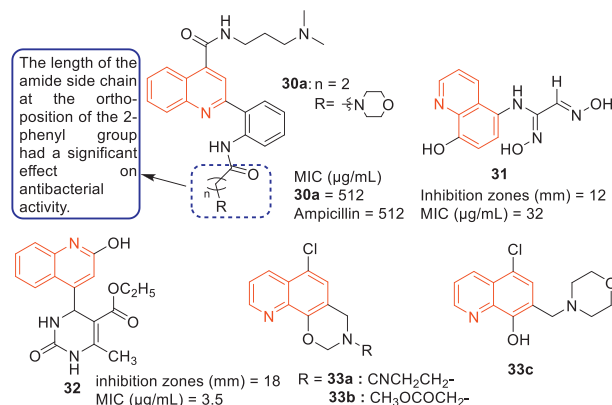
concentration index (FICI) of **27** with methicillin was 0.281. The combination of **27** with ampicillin or oxacillin also exhibited synergistic effects on MRSA with FICIs of 0.5 and 0.375, respectively. Compound **27** increased the antibacterial activity of ampicillin by 4 times (MIC decreased from 48 μg/mL to 12 μg/mL) and increased the antibacterial activity of oxacillin against MRSA by 8 times (MIC decreased from 256 μg/mL to 32 μg/mL) at a concentration of 0.375 μg/mL. In addition, when combined with compound **27**, imipenem and ceftazidime exhibited enhanced antibacterial activity with an FICI of 0.75 (Table 8). Mechanistic studies revealed that compound **27** inhibited bacterial growth by inhibiting the GT-Pase activity of FtsZ.

Teng *et al.* [84] evaluated the antibacterial activity of eight quinoline derivatives with daptomycin as a reference drug. As shown in Table 9, when both R<sup>1</sup> and R<sup>2</sup> groups were kept as *p*-methyl, the only weak activity of compound **28a** was detected, and the MIC of **28a** against MRSA was 12 μg/mL. When the R<sup>1</sup> group was replaced by trifluoromethyl, the resulting compound **28b** had better activity with an MIC of 3.0 μg/mL against MRSA. Subsequently, Teng's group [84] retained the R<sup>1</sup> as the CF<sub>3</sub> group and modified R<sup>2</sup> as *m*-trifluoromethyl, *p*-trifluoromethyl, and 3-chloro-4-fluoro groups, respectively, yielding compounds **28c**, **28d**, and **28e**. Compared with **28b**, quinoline derivative **28c** showed no change in antibacterial activity against MRSA, while compound **28d** exhibited enhanced activity against MRSA with a MIC of 0.75 μg/mL, indicating the *p*-CF<sub>3</sub> was more beneficial than the *p*-CH<sub>3</sub> group in enhancing the anti-MRSA activity of these quinoline derivatives. Compared with compound **28b**, the substitution of the 3-chloro-4-fluoro group on compound **28e** also showed better activity against MRSA. Based on this result, the R<sub>2</sub> group of compounds **28f**, **28g** and **28h** were changed to 3-chloro-4-fluoro groups, but with various R<sub>1</sub> groups. The quinoline compound **28f** with a *p*-isopropyl phenyl ring also exhibited strong antibacterial activity against MRSA with an MIC value of 1.5 μg/mL. Additionally, compound **28g** was synthesized by substituting the phenyl ring in the *ortho* position, and it also displayed strong efficacy against MRSA with an MIC value of 1.5 μg/mL. The introduction of an ethoxy spacer between the benzene ring and oxygen at 4-position resulted in a compound **28h**, whose antibacterial activity was not completely destroyed, with an MIC value of 3.0 μg/mL.

In 2021, compounds **29a–r** (Fig. 16) were synthesized by Manhas *et al.* [85] using materials reported in the relevant literature to react with various aldehyde condensations. All synthesized quinazolin-4-one Schiff bases **29a–r** were subjected to *in vitro* antibacterial screening, and the zone of inhibition was determined at a concentration of 1.0 mg/mL, and the zone of inhibition between 7 and 9 mm and was considered effective [86].



**Fig. 16.** Structures of compounds **29a–r**.



**Fig. 17.** Structure of compounds **30–33** and their anti-MRSA activities.

Monofluoroquinazolin-4-ketone was not effective against MRSA, but the *p*-trifluoromethyl derivative **29e**, the 3-oxygenated trifluoromethyl derivative **29f**, the 2-chloro-6-fluoro derivative **29g**, and the 3,5-difluoro derivative **29h** exhibited good antibacterial effects against MRSA. Notably, dimethylaminophenyl and 3-hydroxy-4-methoxyphenyl derivatives **29k** and **29l** were also active against MRSA. Therefore, these compounds could be used as lead compounds for modifying their structures and improving their anti-MRSA activities.

During 2000–2016, Dinakaran *et al.*, Strigacova *et al.*, and Wang *et al.* [87–89] synthesized three series of quinoline-4-carboxylic acid derivatives on the basis of quinoline-4-carboxylic acid and evaluated their antibacterial activities. Among them, compound **30a** (Fig. 17) showed good antibacterial activity against MRSA with an MIC value of 512 μg/mL, which was comparable to the antibacterial activity of ampicillin (Fig. 17). Additionally, MTT assay showed that compound **30a** had low cytotoxicity. SARs revealed that 2-phenyl ortho amide side chain length had a significant effect on the antibacterial activity, for the same end of the alkaline, with a long side chain ( $n = 2$ , three bonds between the basic N terminus and carbonyl group) compounds exhibited stronger antibacterial activity than those derivatives with shorter side chain and a carbonyl group ( $n = 1$ , two bonds between the basic N terminus and carbonyl group), respectively. The cyclic amino group at 2-phenyl group can increase the antibacterial activity of quinoline-4-carboxylic acid derivatives against MRSA.

In 2012, Sevgi *et al.* [90] synthesized several quinoline-based ethylenedione derivatives and *N*-(8-hydroxyquinolin-5-yl)-aminoethylenedione and evaluated their anti-MRSA activities. It was worth mentioning that compound **31** (Fig. 17) exhibited the

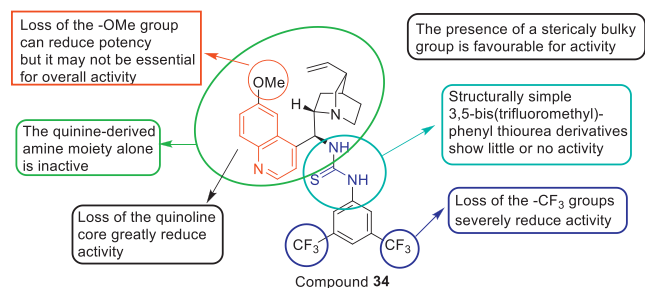
most effective anti-MRSA activity with an inhibition zone of 12 mm and an MIC value of 32  $\mu\text{g}/\text{mL}$ . Among the dihydropyrimidinone derivatives synthesized by Paul *et al.* [91], compound **32** (Fig. 17) displayed good antibacterial activity against MRSA with an MIC value of 3.5  $\mu\text{g}/\text{mL}$ , superior to the positive control streptomycin (MIC = 3.9  $\mu\text{g}/\text{mL}$ ).

## 7. 7-,8-Positions modified quinoline derivatives

The compound 5-chloro-13-phenethyl-13,14-dihydro-2H-[1,3]oxazino[5,6-H]quinoline (**33**, Fig. 17) was first reported by Enquist's research group [92], which had a unique scaffold and was effective against Gram-negative bacteria, including *E. coli* and *Pseudomonas aeruginosa*. In 2019, Fu's group firstly elucidated the new mechanism of action of compound **33** against *E. coli*, mainly through blocking lipopolysaccharide (LPS) transport (Lpt) A-LptC interaction by targeting LptA [93]. Using compound **33** as a precursor, a series of oxazole-quinoline derivatives were designed and another series of quinoline derivatives with 5-chloro atoms were constructed, while the hybrid compound **33c** was prepared by linking a quinolone fragment as a special substituent to the quinoline core (Fig. 17) [94]. The antibacterial experiments showed that compounds **33a**, **33b**, and **33c** had potent effects on drug-resistant strain MRSA, with MIC values of 8  $\mu\text{g}/\text{mL}$ . The results of bacterial molecular docking studies revealed that compound **33c** had the advantage of dual-targeting mechanism of LptA and topoisomerase IV.

## 8. Thiourea-containing quinone derivatives

Among the thiourea-containing quinone derivatives (Fig. 18) synthesized by Dolan *et al.* [95], compound **34** was the most active quinone derivative with an MIC<sub>90</sub> value of 17.74  $\mu\text{g}/\text{mL}$ , which was higher than the reported MIC<sub>90</sub> value of 1.35  $\mu\text{g}/\text{mL}$  or 2 mg/L for vancomycin against MRSA. The SARs in Fig. 18 indicated that: (1) loss of the -OMe group can reduce antibacterial potency but it may not be essential for overall activity; (2) The presence of a sterically bulky group was favorable for anti-MRSA activity and the quinone-derived amine moiety alone was inactive; (3) Structurally simple 3,5-bis(trifluoromethyl)-phenyl thiourea derivatives exhibit little or no anti-MRSA activity. Additionally, toxicity evaluation showed that **34** was not only non-toxic to *Galleria mellonella* larvae but also did not appear to impede larvae development. Furthermore, compound **34** was also non-toxic to *G. mellonella* larvae at concentrations up to 1000  $\mu\text{g}/\text{mL}$ . Encouraged by these results, thiourea-containing quinone derivative **34** had great potential as a new antibacterial agent against MRSA.



**Fig. 18.** SARs of quinone-based thiourea **34**. Reproduced with permission [95], Copyright 2016, Elsevier publications.

## 9. Antibacterial mechanism of quinoline analogues against MRSA

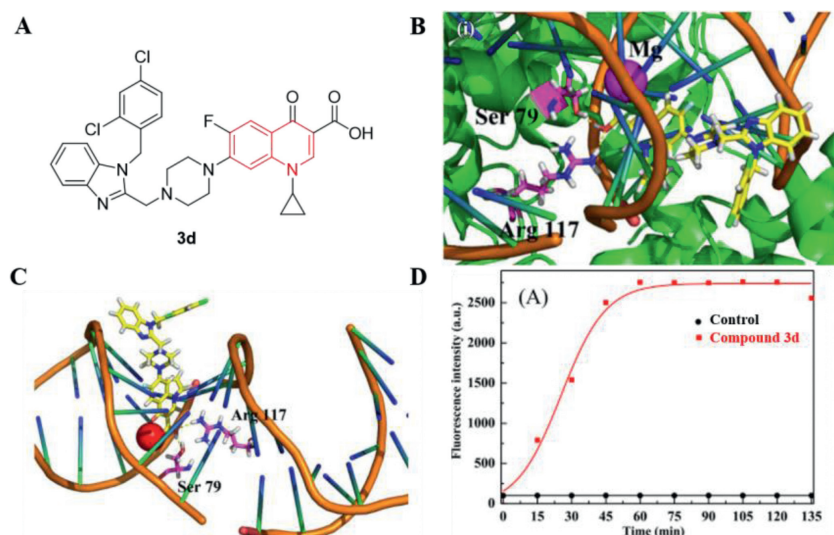
Here, we briefly summarized the anti-MRSA mechanisms of some representative quinoline analogues based on the types of targets and the process of action between quinoline analogues and bacteria, mainly including the interaction of DNA topoisomerase, disruption of cell membranes and inhibition of the FtsZ associated with cell division.

### 9.1. DNA topoisomerase

It is well known that DNA topoisomerases are the main targets of quinolone antibiotics. Herein, compound **3d** was able to inhibit the relaxation activity of EcTopo IV at a concentration of 10  $\mu\text{mol}/\text{L}$  [40]. Interactions of compound **3d** (Fig. 19A) with DNA topoisomerase IV receptor were shown in Figs. 19B and C, the carboxyl group of this molecule was very close to residue Arg117 of the DNA topoisomerase IV complex. Compound **3d** can also form hydrogen bonds with Ser79 of the DNA topoisomerase IV complex via the hydrogen atom of the carboxyl group. In addition, compound **3d** can insert into the superhelical DNA of the enzyme-DNA complex. This synergistic binding may facilitate the formation of a stable quinolone-DNA enzyme complex, which may account for the strong inhibitory effect of **3d** on MRSA. Moreover, Zhang *et al.* [40] used propidium iodide (PI) to study the ability of **3d** to penetrate bacterial cell membranes and found that **3d** can cross the damaged bacterial cell membranes and bind to DNA to emit stronger fluorescence (Fig. 19D).

### 9.2. Cell membranes

The bacterial cell membrane is one of the important structures on which bacteria depend for survival and is rich in enzyme systems that perform many important metabolic functions. In 2010, Hu *et al.* reported that a quinoline derivative HT61 (**1**) can act on the plasma membrane of bacterial cells by disrupting the bacterial cell membrane potential of MSSA, resulting in the release of cellular contents [38]. The polarization effect of the quinoline derivative HT61 on MSSA bacterial membranes was detected by the fluorescent probe 3,3'-dipropylthiadicarbocyanine iodide (DiSC3(5)). As shown in Fig. S1A (Supporting information), 1 min after the addition of HT61, the fluorescence intensity was enhanced in a concentration-dependent manner for the stationary phase MSSA. While the fluorescence intensity did not increase significantly after treatment for 1 min. This result suggested that the maximum depolarization of the cytoplasmic membrane of stationary phase MSSA occurred within 1 min to 4 min after being treated with HT61. However, for log phase MSSA (Fig. S1B, Supporting information), the release of fluorescence after HT61 treatment was slower than that of the stationary phase and achieved a peak after 45 min. It appears that the increased fluorescence intensity was concentration-dependent only at lower concentrations of HT61. When HT61 reached higher concentrations, the fluorescence release did not increase when the concentration of HT61 was higher than 20 mg/mL. Furthermore, transmission electron micrographs showed that after treatment with HT61 for 10 min, the cell membrane of *S. aureus* was disrupted and the cytoplasm exuded into the extracellular space (Fig. S2B, Supporting information). Additionally, after treatment with HT61 at higher concentrations, the cell wall cracked and the cell contents were expelled (Figs. S2C and D, Supporting information).



**Fig. 19.** (A) Structure of **3d**; (B, C) Three-dimensional conformation of **3d** docked in topoisomerase IV-DNA complex; (D) Mechanism of antibacterial action of **3d** at concentrations of  $12\times$  MIC. Reproduced with permission [40], Copyright 2016, Elsevier publications.

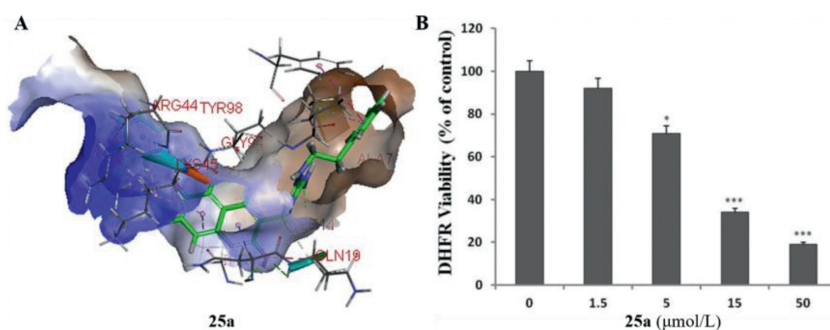
### 9.3. FtsZ

FtsZ is highly conserved in bacteria and is essential for cell division [96]. Recent studies have reported that quinoline derivatives induce cell death in antibacterial assays possibly due to their inhibitory effect on the GTPase activity of FtsZ, making FtsZ a novel target for the development of broad-spectrum antibiotics [97]. As shown in Fig. S3 (Supporting information), compound **17b<sub>2</sub>** strongly inhibited Sa-FtsZ polymerization in a dose-dependent manner, exhibiting a significant inhibition of FtsZ polymerization at a concentration of  $16\ \mu\text{g}/\text{mL}$  [65]. The effect of the *N*-methylbenzindolo[3,2-*b*]quinoline derivative **17b<sub>2</sub>** on FtsZ polymerization was also analyzed by transmission electron microscopy by Sun *et al.* [65]. As displayed in Fig. S4A (Supporting information), a dense network of FtsZ protofilaments with an average width of  $104 \pm 18\ \text{nm}$  was observed in the absence of **17b<sub>2</sub>**. While a concentration of  $8\ \mu\text{g}/\text{mL}$  of **17b<sub>2</sub>** greatly reduced the size and thickness of FtsZ polymer and the bundling of FtsZ protofilaments (Fig. S4B, Supporting information). Since the GTPase activity of FtsZ correlates with the dynamic polymerization of FtsZ [98], the effect of compound **17b<sub>2</sub>** on the dynamic polymerization of FtsZ was investigated. In the GTPase assay, **17b<sub>2</sub>** showed about 20% inhibition at a concentration of  $4\ \mu\text{g}/\text{mL}$  (Fig. S4C, Supporting information). However, approximately 50% and 60% inhibition were reached at concentrations of 8 and  $16\ \mu\text{g}/\text{mL}$ , respectively. These results suggested that **17b<sub>2</sub>** significantly inhibited the GTPase activity of FtsZ

in a dose-dependent manner. In 2018, Fang *et al.* [83] found that compound **27** inhibited bacterial growth by inhibiting the GTPase activity of FtsZ. They used transmission electron microscopy to find a significant increase in the size of the FtsZ polymer after incubation of FtsZ with **27** at a concentration of  $1.5\ \text{mg}/\text{mL}$  (Figs. S4D and E, Supporting information). Moreover, compound **27** at  $0.75\ \text{mg}/\text{mL}$  showed about 20% inhibitory activity, and **27** at concentrations of 1.5, 3, and  $6\ \text{mg}/\text{mL}$  can achieve 50%, 70%, and 80% inhibition effects (Fig. S4F, Supporting information). These results suggested that the anti-MRSA activity of compounds **17b<sub>2</sub>** and **27** may be due to their inhibitory effects on GTPase activity and FtsZ polymerization.

### 9.4. DHFR

DHFR, is involved in the biosynthetic pathway of folate, a nitrogenous base and precursor of some amino acids. DHFR is one of the key components of this pathway and inhibition of DHFR can be lethal to bacteria. Thus, DHFR can be used as a target against pathogenic bacteria. Bai *et al.* [80] reported that compound **25a** was well inserted into the active pocket of *S. aureus* DHFR (Fig. 20A) and it could reduce the activity of DHFR by 81% at a concentration of  $50\ \mu\text{mol}/\text{L}$  compared to the negative control (Fig. 20B). It was confirmed that compound **25a** showed an inhibitory effect on DHFR, indicating that DHFR may be a potential antibacterial target [80].



**Fig. 20.** Inhibition of DHFR activities of compound **25a**. Reproduced with permission [80], Copyright 2019, John Wiley publications. \* $P < 0.05$ , \*\*\* $P < 0.001$ , significant with respect to the control ( $0\ \mu\text{mol}/\text{L}$ ) group.

## 10. Conclusions and outlook

In this review, we addressed what is known about the antimicrobial activity of quinoline analogues over the past two decades, particularly against MRSA, their SARs, as well as mechanisms. Quinoline analogues have a wide range of biological and pharmaceutical activities, and a number of quinoline analogues have been designed, developed, and screened against Gram-positive, Gram-negative, and MDR bacteria in recent two decades. Many quinoline analogues exhibited more potent antibacterial activity against MRSA than the reference antibiotics and were less toxic. The SARs suggested that the anti-MRSA activity of quinoline analogues is related to the position, species, and spatial relationships of the various substituents. Their abundant SARs may provide better insights for further rational development of quinoline-based antimicrobials to combat MRSA infections. Additionally, the elucidation of the anti-MRSA mechanism of some representative quinoline analogues will provide a reference for the design of novel quinoline-based antibacterial drugs in the future.

## Declaration of competing interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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## Supplementary materials

Supplementary material associated with this article can be found, in the online version, at doi:10.1016/j.ccl.2023.108511.

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