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A series of simple curcumin-derived colorimetric and fluorescent probes for ratiometric-pH sensing and cell imaging



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ARTICLE INFO

Article history:

Received 18 March 2021

Revised 23 June 2021

Accepted 28 June 2021

Available online 20 July 2021

Keywords:

Natural product

Fluorescent probe

Ratiometric pH sensing

Colorimetric response

pH Bioimaging

ABSTRACT

Colorimetric and fluorescent probes have emerged as a potent tool for pH sensing due to easy operation and high sensitivity. However, most of the existing bimodal probes require complicated synthesis, which greatly limits their wide applications. Herein, a simple fluorescent dye (called BFCUR) featuring a D- π -A- π -D conjugated system was developed from the natural polyphenol curcumin (CUR). BFCUR exhibited significant red-shift in UV absorption and fluorescence emission as pH increased because of the deprotonation of the phenolic hydroxyl groups, which resulted in the enhanced intramolecular charge transfer (ICT). The ratiometric pH detection of BFCUR was achieved with remarkable accuracy by monitoring both the absorbance ratio A_{500}/A_{650} and the fluorescence intensity ratio I_{622}/I_{743} under various pH values. In addition, the clear color changes of BFCUR under different pH conditions were visible, which enabled BFCUR to be used in test strips for rapid, visual pH detection. Moreover, BFCUR exhibited low cytotoxicity, and was successfully applied for intracellular pH detection, where the fluorescence intensity was linearly related to pH value. This study highlighted the great potential of CUR-derived BFCUR as colorimetric and fluorescent probes for ratiometric-pH sensing and cell imaging.

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pH Value is one of the important physical and chemical parameters in the fields of industry, agriculture, medicine, environmental protection and scientific research [1]. pH is also associated with many health complaints, including cancer and neurodegenerative diseases, because slightly disrupted pH balance leads to cell dysfunctions [2–4]. Thus, high-precision pH sensing and monitoring is essential for understanding its effect on related cellular functions and pathological processes. Fluorescence probes have recently shown great potential for monitoring pH fluctuations because of its high spatiotemporal resolution, high sensitivity, non-invasiveness and easy operation [5]. Various fluorescent probes have been developed for pH sensing [6–13]. Among them, ratiometric fluorescent probes that report solution pH by exhibiting changes of the measured intensity ratio at two excitation or emission wavelengths are superior in quantitative pH analysis compared to probes with single detection window [14–17]. The in-

trinsic self-calibration capability of ratiometric fluorescence probes avoided the interference caused by external factors such as fluctuation of probe concentration, environmental conditions and instrument sensitivity, accordingly enhancing the measurement accuracy [18–20]. Additionally, red/near-infrared fluorescent sensors are highly desirable for *in vivo* physiological pH measurement since longer wavelengths photons can more easily penetrate into deep tissues and were less interfered by autofluorescence [21–27].

Most of the reported fluorescent pH chemosensors were obtained by complex, environmentally-unfriendly and costly chemical synthesis, which limited their widespread use [28,29]. Fluorescent molecules and dyes from natural sources are often easily obtained and environmentally friendly [30], and their photophysical properties have therefore attracted increasing attention from the research community. Accordingly, several fluorescence probes based on natural products have been recently reported [31–33], showing great promise as biocompatible and biodegradable alternatives in the development of new pH sensors. Curcumin (CUR), a fluorescent polyphenol extracted from *Curcuma longa* roots, has been widely investigated for its pharmacological activity as, for example, an antioxidant or anticancer drug [34,35]. The excellent op-

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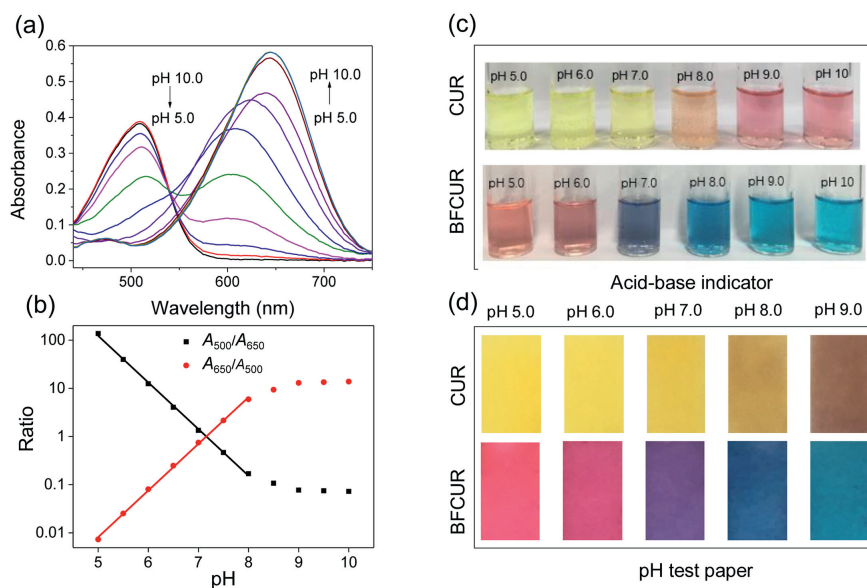


Fig. 1. (a) UV-vis spectra of BFCUR (10 μmol/L) at different pH values (5.0–10.0) in the B-R buffer/EtOH (1/1, v/v) solution. (b) The plot of the absorbance ratios of BFCUR at A_{500}/A_{650} and A_{650}/A_{500} vs. pH values including the linear relationships of both plots ranging from pH 5.0 to 8.0. (c) The color changes of CUR and BFCUR solutions at different pH values (5.0–9.0). (d) The observed color changes of CUR- and BFCUR-coated test strips after soaking in aqueous solution at different pH values (5.0–9.0) by naked eyes.

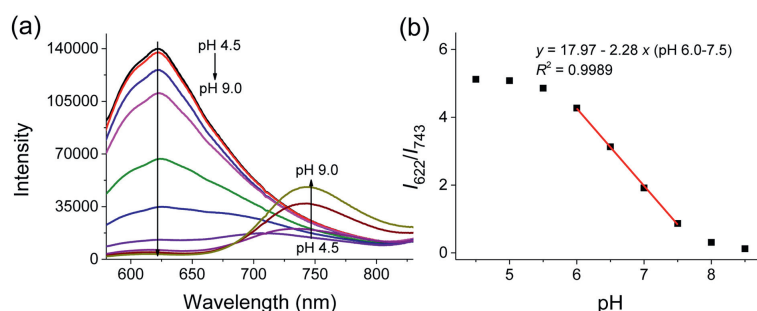


Fig. 2. (a) Fluorescence titration spectra of BFCUR (10 μmol/L) at different pH values (4.5–9.0) in the B-R buffer/EtOH (1/1, v/v) solution ($\lambda_{\text{ex}} = 550$ nm). (b) The plot of the fluorescence intensity ratios of BFCUR at I_{622}/I_{743} vs. pH values.

tical characteristics also allowed CUR to be used in sensor development [36,37]. Till now, several CUR analogues have been developed for $A\beta$ plaque and cyanide detection [38–40]. It has also been suggested that CUR could be used in qualitative pH sensing test strips due to the colorimetric behavior [36]. However, the short emission wavelength of CUR has prevented the use of CUR-based fluorescent probes in biological applications.

To obtain longer emission wavelengths, a CUR derivative (BFCUR, Scheme 1) was prepared by introducing boron trifluoride moiety in the middle of CUR molecule. This pH-sensitive fluorescent BFCUR exhibited absorption and emission maxima at longer wavelengths than those of CUR. The underlying mechanism for the photophysical behavior was further explored by singly or doubly methylating the phenolic hydroxyl groups of BFCUR to obtain MBFCUR and MBFCURM, respectively. The acid-base response of BFCUR, MDFCUR and MBFCURM demonstrated that the deprotonation process of phenol hydroxyl groups was responsible for the fluorescence intensity changes caused by a change in solution pH. BFCUR demonstrated the strongest response to pH changes and was evaluated as a ratiometric colorimetric and fluorescent pH sensing probe by monitoring the absorbance ratio A_{500}/A_{650} and the fluorescence intensity ratio I_{622}/I_{743} , realizing further test strip-based detection and cell imaging.

The phenol groups in the CUR structure are easily protonated and deprotonated, which cause clear color changes in response to

the changing pH conditions. Therefore, CUR has been considered as an optical acid-base sensor [41]. However, the short UV-vis absorption and fluorescence emission wavelength of CUR has so far limited its widespread use. CUR features a D- π -A- π -D conjugated system with two intramolecular charge transfer (ICT) interactions [42]. The electron acceptors and donors are respectively in the middle and at both ends of the molecule. Enhancing the electron-withdrawing ability is an effective strategy to shift the absorption and emission peaks to longer wavelengths. Therefore, boron trifluoride with high electronegativity was introduced into CUR to obtain the derivative BFCUR via a one-pot synthesis with high yield and longer excitation/emission wavelengths (Scheme 1, Figs. S1–S3 in Supporting information). Subsequently, singly or doubly methylated MBFCUR and MBFCURM were also synthesized to provide mechanism insight into the pH sensitivity and to compare the sensing performance (Scheme 1, Figs. S4–S9 in Supporting information).

UV-vis spectra of BFCUR at diverse pH values were studied. Compared with CUR, the absorption maxima wavelength of BFCUR shifted from 430 nm to 500 nm under acidic and neutral conditions and from 530 nm to 650 nm under alkaline conditions (Fig. S10). BFCUR displayed a strong absorbance at 500 nm and no obvious absorption peak at 650 nm at pH 5.0. As the pH increased from 5.0 to 8.0, the absorbance maximum at 500 nm disappeared gradually, which coincided with a remarkable absorbance turn-on re-

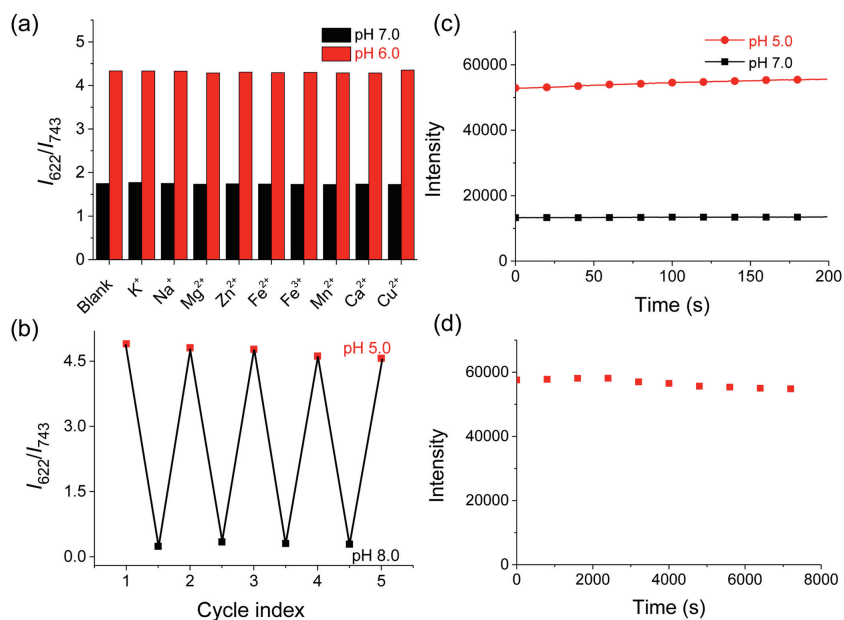


Fig. 3. (a) The fluorescence intensity ratio of BFCUR (10 $\mu\text{mol/L}$) at I_{622}/I_{743} containing different metal cations in the solution (B-R buffer/EtOH: 1/1, v/v) at pH 6.0 and 7.0 ($\lambda_{\text{ex}} = 550 \text{ nm}$). The ions are KCl (150 mmol/L), NaCl (15 mmol/L), MgCl_2 (2 mmol/L), ZnCl_2 (2 mmol/L), FeCl_2 (50 $\mu\text{mol/L}$), FeCl_3 (50 $\mu\text{mol/L}$), MnCl_2 (50 $\mu\text{mol/L}$), CaCl_2 (2 mmol/L) and CuCl_2 (50 $\mu\text{mol/L}$) at intracellular physiological concentrations. (b) Fluorescence reversibility of BFCUR (10 $\mu\text{mol/L}$) in the solution (B-R buffer/EtOH: 1/1, v/v) between pH 5.0 and 8.0. (c) Time courses of the fluorescence intensity at 622 nm ($\lambda_{\text{ex}} = 550 \text{ nm}$) of BFCUR (10 $\mu\text{mol/L}$) in the solution (B-R buffer/EtOH: 1/1, v/v) at pH 5.0 and 7.0. (d) The fluorescence intensity at 622 nm ($\lambda_{\text{ex}} = 550 \text{ nm}$) of BFCUR in the continuous excitation light irradiation for 120 min.

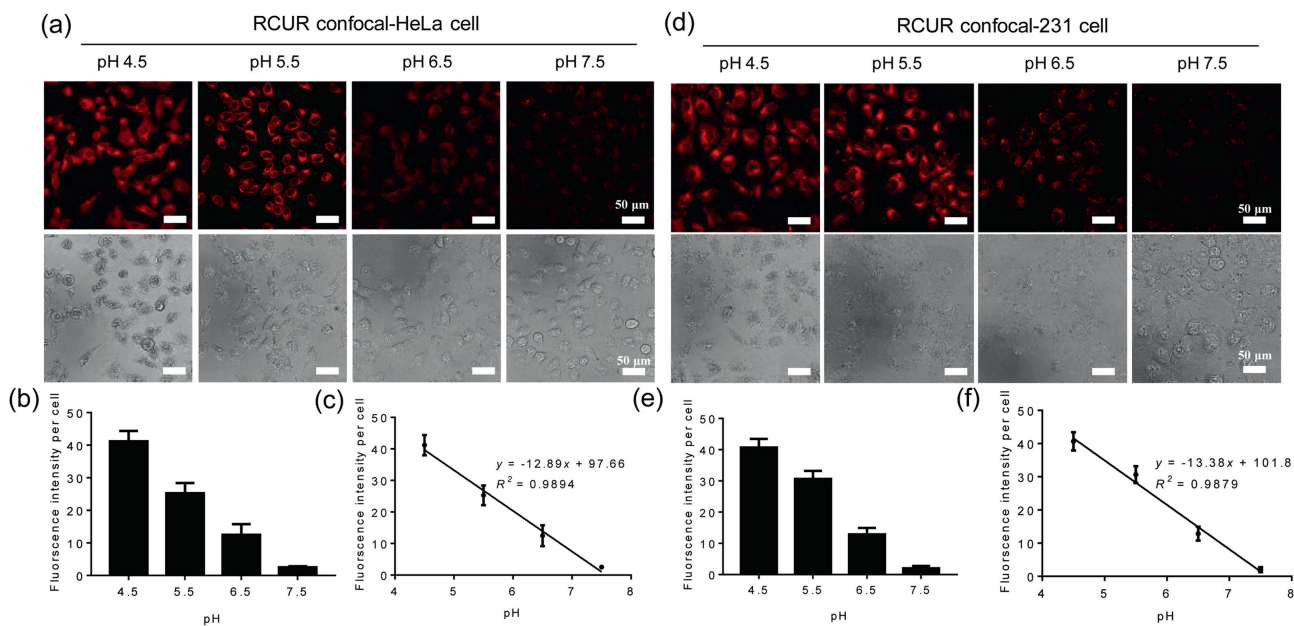
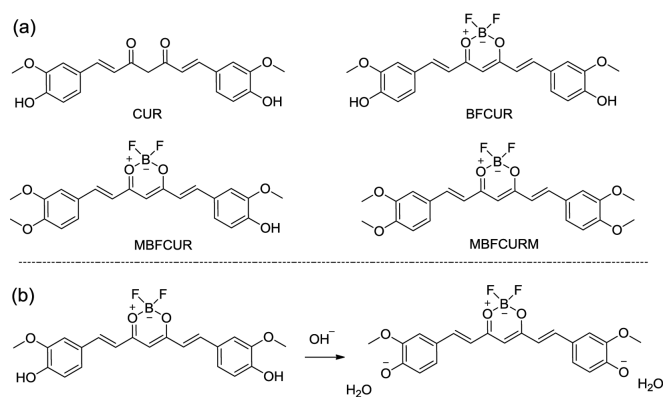


Fig. 4. Images of (a) HeLa cells and (d) MDA-MB-231 cells with BFCUR incubated at pH 4.5, 5.5, 6.5, 7.5. (b, c, e, f) The fluorescence intensity quantitation was analyzed by the Image J.

sponse at 650 nm (Fig. 1a). The ratio of absorbance (A_{500}/A_{650}) was linearly correlated with the pH between 5.0 and 8.0 ($R^2 = 0.99$, Fig. 1b). Moreover, clear color changes from red to purple to blue were observed as the pH of BFCUR solutions in B-R buffer changed from 5.0 to 8.0 (Fig. 1c). In order to exploit this, BFCUR-based test strips for the qualitative pH determination were prepared. As the pH gradually increased from 5.0 to 8.0, the color of the BFCUR-based test papers expectedly changed from red (pH < 7) to purple (pH 7) to blue (pH > 7) (Fig. 1d). For comparison, the correspond-

ing colorimetric response of the CUR-based test strips was much less clear, where the color changed from yellow to sandy brown.

The fluorescence emission spectra of BFCUR varied with different excitation wavelengths (Fig. S11 in Supporting information). When $\lambda_{\text{ex}} = 500 \text{ nm}$, the fluorescence intensity at 622 nm increased markedly as pH value decreased from 8 to 6. Meanwhile, when $\lambda_{\text{ex}} = 650 \text{ nm}$, the fluorescence intensity at 743 nm increased significantly as pH increased from 7 to 9. Notably, when $\lambda_{\text{ex}} = 550 \text{ nm}$, the fluorescence intensity at 622 nm also decreased as the pH increased, while a new emission peak at 743 nm grad-



Scheme 1. (a) Structures of CUR, BFCUR, MBFCUR and MBFCURM. (b) Proposed response mechanisms of BFCUR to pH.

ually emerged (Fig. 2a). The pH values in the range of 6.0–7.5 and the ratios of fluorescence intensities (I_{622}/I_{743}) showed a strong linear relationship (Fig. 2b). This suggested that BFCUR could be applied as a ratiometric fluorescence sensor for detecting changes of pH. The large pK_a of 6.75 indicated its capability of BFCUR to test weak acid.

Since both CUR and BFCUR could respond to pH in real time, we hypothesized that the underlying response mechanism was related to the increased electron donating ability of BFCUR after deprotonation under basic condition. Deprotonation of the phenolic hydroxyl groups enhanced the intramolecular charge transfer (ICT), leading to a red shift of UV-vis and fluorescence spectra. In order to confirm this, another two derivatives MBFCUR and MBFCURM were further synthesized by singly or doubly methylating the phenolic hydroxy groups on BFCUR, respectively. By diminishing the extent of deprotonation, a change in response to pH was expected. UV-vis and fluorescence spectra obtained from MBFCUR showed less red-shift in response to an increasing pH compared to BFCUR, while MBFCURM showed only negligible response to an increased pH (Fig. S13 in Supporting information). These data proved the rationality of the hypothesized response mechanism.

The selectivity of BFCUR was evaluated *via* measuring the fluorescence spectra of BFCUR in the presence of relevant metal ions, such as K^+ , Na^+ , Mg^{2+} , Zn^{2+} , at physiological concentrations. None of the tested metal ions had any effect on the fluorescent properties of BFCUR at pH 6.0 or 7.0 (Fig. 3a). The reversibility of BFCUR was examined by cycling the pH between 5.0 and 8.0 in B-R buffer, where only negligible changes in fluorescent properties were observed after four cycles (Fig. 3b). The time-dependent fluorescence of BFCUR at pH 5.0 and 7.0 was investigated by monitoring the fluorescence intensity at 622 nm. After BFCUR was added to B-R buffer solutions at pH 5.0 or 7.0, the fluorescence intensity at 622 nm rapidly reached a stable state, which indicated a real-time response to pH change (Fig. 3c). After continuous light irradiation ($\lambda_{ex} = 550$ nm) for 2 h, the fluorescence intensity of BFCUR at 622 nm showed only a slight decrease, indicating excellent photostability (Fig. 3d).

Inspired by the excellent pH-dependent fluorescence properties of BFCUR, we wondered if it can be applied *in vivo* as a real-time pH probe. The cytotoxicity of BFCUR was first assessed against HeLa cells and MDA-MB-231 cells, where cell viabilities exceeded 75% in groups treated with up to 5 $\mu\text{mol/L}$ BFCUR for 24 h (Fig. S14 in Supporting information). Confocal micrographs showed that the red fluorescence intensity increased as the BFCUR concentration increased, suggesting excellent cell permeability (Fig. S15 in Supporting information). Evaluation of the intracellular pH reporting capabilities was performed using an intracellular pH calibration

buffer kit [27]. A clear decrease in fluorescence intensity was observed as the pH increased from 4.5 to 7.5, where the average fluorescence intensity per cell was linearly related to intracellular pH values (Fig. 4). Moreover, the same response was observed in both cell types, indicating the good stability and universality of BFCUR in reporting intracellular pH.

In this work, three CUR derivatives were designed with different numbers of phenolic hydroxyl groups, and the role of phenolic hydroxyl in pH sensing was explored. BFCUR displayed pH reporting capabilities through ratiometric analysis of absorption spectra between pH 5.0 and 8.0 and fluorescence emission spectra between pH 6.0 and 7.5 with a large pK_a of 6.75; and through colorimetric analysis between pH 5.0 and 8.0. Qualitative pH test papers prepared by BFCUR displayed significantly different colors depending on the solution pH. Moreover, BFCUR was also used to report the intracellular pH of different cell lines with a high accuracy. Owing to the high sensitivity, excellent stability and reversibility, and crystal-clear color changes observed by eye in response to subtle changes in pH, BFCUR shows great promise as a powerful and universal tool for the rapid and accurate reporting of pH.

Declaration of competing interest

The authors declare no conflict of interest.

Acknowledgments

This study was supported by the National Natural Science Foundation of China (Nos. 51922111, 21774054, 21574061), the Science and Technology Development Fund, Macau SAR (No. 083/2017/A2), Guangdong-Hong Kong-Macao Joint Laboratory of Optoelectronic and Magnetic Functional Materials (No. 2019B121205002), and funded by Shenzhen Fundamental Research Programs (No. JCYJ20170412152922553).

Supplementary materials

Supplementary material associated with this article can be found, in the online version, at doi:10.1016/j.ccl.2021.06.076.

References

- [1] S. Dong, M. Luo, G. Peng, W. Cheng, *Sens. Actuator. B* 129 (2008) 94–98.
- [2] T.A. Davies, R.E. Fine, R.J. Johnson, et al., *Biochem. Biophys. Res. Commun.* 194 (1993) 537–543.
- [3] M. Schindler, S. Grabski, E. Hoff, S.M. Simon, *Biochemistry* 35 (1996) 2811–2817.
- [4] S. Sinharay, M.D. Pagel, *Annu. Rev. Anal. Chem.* 9 (2016) 95–115.
- [5] J. Han, K. Burgess, *Chem. Rev.* 110 (2010) 2709–2728.
- [6] S. Takahashi, Y. Kagami, K. Hanaoka, et al., *J. Am. Chem. Soc.* 140 (2018) 5925–5933.
- [7] X. Liu, Y. Su, H. Tian, et al., *Anal. Chem.* 89 (2017) 7038–7045.
- [8] F. Su, S. Agarwal, T. Pan, et al., *ACS Appl. Mater. Interfaces* 10 (2018) 1556–1565.
- [9] D. Iacopini, A. Moscardini, F. Lessi, et al., *Bioorg. Chem.* 105 (2020) 104372.
- [10] M.J. Marin, F. Galindo, P. Thomas, D.A. Russell, *Angew. Chem. Int. Ed.* 51 (2012) 9657–9661.
- [11] C.B. He, K.D. Lu, W.B. Lin, *J. Am. Chem. Soc.* 136 (2014) 12253–12256.
- [12] J. Joniak, H. Stankovicova, J. Filo, et al., *Sens. Actuator. B* 307 (2020) 127646.
- [13] X. Zhao, C. Wang, G. Yuan, et al., *Sens. Actuator. B* 290 (2019) 79–86.
- [14] A.P. de Silva, H.Q. Gunaratne, T. Gunnlaugsson, et al., *Chem. Rev.* 97 (1997) 1515–1566.
- [15] F. Galindo, M.I. Burguete, L. Vigara, et al., *Angew. Chem. Int. Ed.* 44 (2005) 6504–6508.
- [16] M. Lee, N.G. Gubernator, D. Sulzer, D. Sames, *J. Am. Chem. Soc.* 132 (2010) 8828–8830.
- [17] Y. Yan, X. Zhang, X. Zhang, et al., *Chin. Chem. Lett.* 31 (2020) 1091–1094.
- [18] X. Liu, Q. Yang, W. Chen, et al., *Org. Biomol. Chem.* 13 (2015) 8663–8668.
- [19] M. Wang, G. Meng, Q. Huang, et al., *Chem. Commun. (Camb.)* 47 (2011) 3808–3810.
- [20] X. Feng, T. Zhang, J.T. Liu, J.Y. Miao, B.X. Zhao, *Chem. Commun. (Camb.)* 52 (2016) 3131–3134.
- [21] L. Yuan, W. Lin, K. Zheng, L. He, W. Huang, *Chem. Soc. Rev.* 42 (2013) 622–661.
- [22] D.S. Koktysh, *Mater. Res. Bull.* 123 (2020) 110686.

- [23] Z.P. She, Y. Tian, Y.S. Xia, et al., *Dyes Pigm.* 179 (2020) 108402.
- [24] W. Shen, L. Wang, S. Zhu, et al., *Anal. Biochem.* 596 (2020) 113609.
- [25] H.X. Li, H. Dong, M.M. Yu, et al., *Anal. Chem.* 89 (2017) 8863–8869.
- [26] J.F. Yang, M. Li, W.H. Zhu, *Res. Chem. Intermed.* 44 (2018) 3959–3969.
- [27] J. Li, X.K. Li, J.B. Jia, et al., *Dyes Pigm.* 166 (2019) 433–442.
- [28] F.Y. Su, S. Agarwal, T.T. Pan, et al., *ACS Appl. Mater. Interfaces* 10 (2018) 1556–1565.
- [29] X.J. Liu, Y.A. Su, H.H. Tian, et al., *Anal. Chem.* 89 (2017) 7038–7045.
- [30] Y. Gu, Z. Zhao, H. Su, et al., *Chem. Sci.* 9 (2018) 6497–6502.
- [31] A. Mars, M. Hamami, L. Bechnak, D. Patra, N. Raouafi, *Anal. Chim. Acta* 1036 (2018) 141–146.
- [32] W. Qin, P.F. Zhang, H. Li, et al., *Chem. Sci.* 9 (2018) 2705–2710.
- [33] F. Zsila, Z. Bikadi, M. Simonyi, *Org. Biomol. Chem.* 2 (2004) 2902–2910.
- [34] M.T. Huang, Y.R. Lou, W. Ma, et al., *Cancer Res.* 54 (1994) 5841–5847.
- [35] H. Hatcher, R. Planalp, J. Cho, F.M. Tortia, S.V. Torti, *Cell. Mol. Life Sci.* 65 (2008) 1631–1652.
- [36] M. Pavai, J. Mihaly, A. Paszternak, *Food Anal. Meth.* 8 (2015) 2243–2249.
- [37] B. Kuswandi, T.S. Larasati Jayus, A. Abdullah, L.Y. Heng, *Food Anal. Meth.* 5 (2012) 881–889.
- [38] C.Z. Ran, X.Y. Xu, S.B. Raymond, et al., *J. Am. Chem. Soc.* 131 (2009) 15257–15261.
- [39] X.L. Zhang, Y.L. Tian, Z. Li, et al., *J. Am. Chem. Soc.* 135 (2013) 16397–16409.
- [40] A. Chaicham, S. Kulchat, G. Tumcharern, T. Tuntulani, B. Tomapatanaget, *Tetrahedron* 66 (2010) 6217–6223.
- [41] D. Xiang, Q. Meng, H. Liu, M. Lan, G. Wei, *Talanta* 146 (2016) 851–856.
- [42] L. Zhou, L. Xie, C. Liu, Y. Xiao, *Chin. Chem. Lett.* 30 (2019) 1799–1808.