



Communication

Aconapelsulfonines A and B, seco C₂₀-diterpenoid alkaloids deriving via Criegee rearrangements of napelline skeleton from *Aconitum carmichaelii*

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ABSTRACT

Two sulfonated seco C₂₀-diterpenoid alkaloids, aconapelsulfonines A (**1**) and B (**2**), were isolated from an aqueous extract of the raw material of “Fu Zi” (the *Aconitum carmichaelii* lateral roots), of which the structures were elucidated by various spectroscopic data, combined with X-ray crystallographic analysis. The unprecedented skeletons are biogenetically proposed to be derived via Criegee rearrangements of the napelline-type architecture. The two compounds exhibited dose-dependent analgesic activities on an acetic acid-induced mice writhing test.

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Because of structural complexities and significant activities, diverse diterpenoid alkaloids have elicited continuous interests from chemical and pharmacological communities over a century [1]. Around 1300 diterpenoid alkaloid natural products have been reported mainly from ranunculaceous plants of the *Aconitum* genus [2]. The lateral root of *Aconitum carmichaelii* Debx., generally designated as “Fu Zi” (Chinese), is a popular Chinese herbal medicine for the treatment of various pains including rheumatism, neuralgia, and angina [3]. As fatal toxicity, this plant medicine must be utilized after long-time decocting or properly processing to remove toxicity [3]. Previous studies showed that poisonous aconitane-alkaloidal diesters could be decomposed into less or no poisonous alkaloidal alcohols or monoesters [3]. However, the aconitane-alkaloidal alcohols and monoesters had relatively low analgesic activity when compared with that of the diesters as well as the decocting extract [3]. This indicates that the extract contains unknown active constituents. Although over 120 constituents have been reported from *A. carmichaelii*, organic solvents (EtOH or MeOH) as well as acid (HCl) and base (NH₄OH) were applied to preparation and

fractionation of the crude extracts [3c,4]. Hence, to be relatively close to the clinical application protocol, we extracted the drug materials with water in our program to reinvestigate the chemical constituents of traditional Chinese medicines, mainly focusing on minor compounds [5]. From an aqueous extract of “Fu Zi”, we have reported 52 diterpenoid alkaloids including six unprecedented sulfonated C₂₀-diterpenoid alkaloids with two novel skeletons deriving from semipinacol rearrangements of napelline- and atisane-types of structural architectures, together with other structural diverse compounds [6]. A continued investigation of remaining active fractions from the extract has led to characterization of further two C₂₀-diterpenoid alkaloids with different novel carbon skeletons originating from the Criegee rearrangements, designated as aconapelsulfonines A (**1**) and B (**2**), respectively (Fig. 1). Herein, we describe their isolation (Supporting information), structural elucidation, possible biogenetic routes, and pharmacological activities.

Compound **1**, colorless prisms (MeOH-H₂O, 3:1; m.p. > 300 °C), [α]_D²⁰ -11.9 (c 0.21, H₂O), showed a strong broad IR absorption (3200–3500 cm⁻¹) for hydroxy functionalities. The nitrogen- and sulfur-containing molecular formula of **1** was determined as C₂₂H₃₃NO₈S by positive and negative mode HR-ESIMS at *m/z* 472.1992 [M+H]⁺ (calcd. for C₂₂H₃₄NO₈S, 472.2000) and 470.1848 [M-H]⁻ (calcd. for C₂₂H₃₂NO₈S, 470.1854), together with NMR spectroscopic data (Table 1). The ¹H NMR spectroscopic data of **1** in

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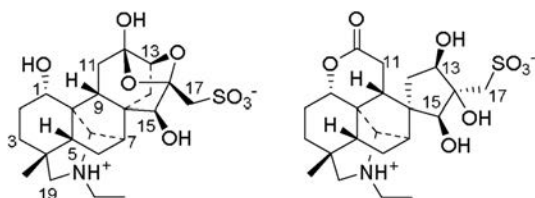
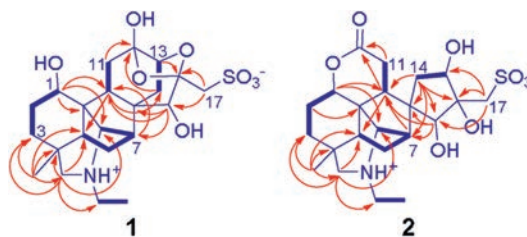
Fig. 1. Structures of **1** and **2**.Fig. 2. Main ^1H - ^1H COSY (blue bold lines) and HMBC correlations (red arrows, from proton to carbon) of **1** and **2**.

Table 1

NMR spectroscopic data for **1** and **2** in D_2O .^a

Position	1		2	
	δ_{H} (J in Hz)	δ_{C}	δ_{H} (J in Hz)	δ_{C}
1	4.12 dd (10.2, 7.2)	68.0	4.76 m	76.6
2a	2.09 m	30.8	2.23 m	25.3
2b	1.49 m		1.84 m	
3a	1.64 m	35.9	1.84 m	35.6
3b	1.46 m		1.58 m	
4		35.4		34.7
5	1.69 d (8.4)	47.8	1.96 d (7.8)	46.3
6a	3.10 dd (15.0, 8.4)	23.1	2.15 dd (15.0, 7.8)	22.6
6b	1.48 m		1.72 dd (15.0, 4.8)	
7	2.59 d (3.6)	45.7	2.54 d (4.8)	50.2
8		43.2		50.1
9	2.38 dd (10.8, 8.4)	40.7	2.76 d (12.0)	39.2
10		51.7		49.7
11a	2.50 dd (13.2, 11.4)	30.4	2.90 dd (16.8, 12.0)	31.2
11b	2.06 m		2.68 d (16.8)	
12		103.8		178.0
13	4.32 brs	80.6	4.24 d (5.4)	74.1
14a	2.07 m	32.3	2.38 dd (15.0, 5.4)	40.3
14b	1.62 m		1.39 d (15.0)	
15	4.22 brs	73.8	4.03 s	78.6
16		108.9		80.7
17a	3.71 d (14.4)	53.3	3.42 d (15.0)	53.2
17b	3.22 d (14.4)		3.20 d (15.0)	
18	0.88 s	24.7	0.92 s	23.9
19a	3.32 d (13.8)	57.9	3.44 d (13.8)	56.6
19b	2.92 d (13.8)		2.95 d (13.8)	
20	3.95 brs	65.1	3.35 brs	65.0
21a	3.24 m	54.5	3.28 m	55.1
21b	3.24 m		3.25 m	
22	1.35 t (7.2)	10.1	1.35 t (7.2)	9.9

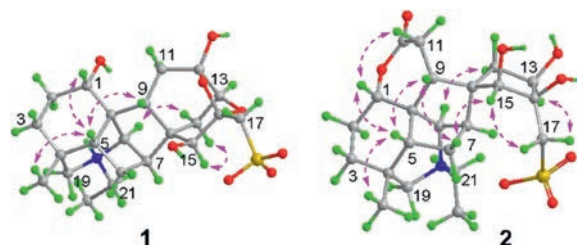
^a Chemical shift data (δ) were acquired at 600 MHz for ^1H NMR and at 150 MHz for ^{13}C NMR. The assignments were based on DEPT, ^1H - ^1H COSY, HSQC, and HMBC experiments.

D_2O revealed the presence of following structures units: (a) a quaternary methyl [δ_{H} 0.88 (s, H_3 -18)]; (b) an *N*-ethyl [δ_{H} 3.24 (2H, m, H_2 -21) and 1.35 (3H, t, J =7.2 Hz, H_3 -22)]; (c) two sulfur- and nitrogen-bearing methylenes attached to quaternary carbons [δ_{H} 3.71 and 3.22 (each d, J =14.4 Hz, H-17a and H-17b) and 3.32 and 2.92 (each d, J =13.8 Hz, H-19a and H-19b)]; (d) four oxygen- and nitrogen-bearing methines [δ_{H} 4.12 (dd, J =10.2 and 7.2, H-1), 4.32 (brs, H-13), 4.22 (brs, H-15), and 3.95 (brs, H-20)]; (e) three aliphatic methines [δ_{H} 1.69 (d, J =8.4 Hz, H-5), 2.59 (d, J =3.6 Hz, H-7), and 2.38 (dd, J =10.8 and 8.4 Hz, H-9)]; and (f) five aliphatic methylenes [δ_{H} 1.46–3.10 (10 H, partially overlapped multiplets, H_2 -2, H_2 -3, H_2 -6, H_2 -11, and H_2 -14)]. The presence of the above structural units was confirmed by further analysis of the ^{13}C NMR, DEPT, and HSQC experimental data, which led to unambiguous assignments of resonances in the NMR spectra of **1**, including five quaternary carbons resonated at δ_{C} 35.4 (C-4), 43.2 (C-8), 51.7 (C-10), 103.2 (C-12) and 108.9 (C-16). The aforementioned spectroscopic data suggested that **1** was a sulfonated unusual C_{20} -diterpenoid alkaloid [6g,6i,6j]. Especially the chemical shift values of the quaternary carbons C-12 and C-16 indicated that there were semi-acetal and acetal functionalities in the structure of

1, which were highly unusual in the C_{20} -diterpenoid alkaloids. Subsequent comprehensive explanation of correlation signals in the ^1H - ^1H COSY and HMBC spectra of **1** constructed an unique skeletal structure.

Four structural fragments in **1** (Fig. 2, bold lines) were elucidated from the homonuclear vicinal coupling cross-peaks of H-1/ H_2 -2/ H_2 -3, H-5/ H_2 -6/H-7/H-20, H-9/ H_2 -11, H-13/ H_2 -14 and H_2 -21/ H_3 -22 in the ^1H - ^1H COSY spectrum. A linkage of the quaternary C-4 with C-3, C-5, C-18 and C-19 was constructed by two- and three-bond heteronuclear correlation signals from H_3 -18 and H_2 -19 to C-3, C-4 and C-5 and from H_3 -18 to C-19 (Fig. 2) in the HMBC spectrum of **1**. A connection of the nitrogen with C-19, C-20, and C-21 was deduced from the HMBC signals of C-20 and C-21 with H_2 -19, together with their chemical shifts. An attachment of C-1, C-5, C-9 and C-20 to the same quaternary C-10 was determined by the HMBC signals from H-1 to C-9, C-10 and C-20 and from H-9 to C-5. The HMBC signals from H-7 to C-9, from H-9 to C-14, and from H-15 to C-7, C-8 and C-9 revealed a linkage of the quaternary C-8 with C-7, C-9, C-14 and C-15. According to the HMBC signals from H-11b and H-14b to C-12, the quaternary carbon C-12 was inserted between C-11 and C-13, while the HMBC signals from H-17b with C-16 and C-15 revealed that the quaternary C-16 connected by C-15 and C-17. Furthermore, the HMBC signal of C-16 with H-13 and their chemical shifts indicated the presence of an oxygen-bridge between C-13 and C-16. To satisfy requirements of the substitution, chemical shift, and molecular composition, three hydroxy groups were positioned at C-1, C-12 and C-15, respectively, a sulfonic acid group was located at C-17, and one more oxygen-bridge was assigned to have the planar structure as given in Fig. 1, which possesses unique novel 13,16-*seco*-napelline carbon skeleton and hydroxydioxolane motif formed *via* the oxygen-bridges between C-16 with C-12 and C-13. The zwitterionic formula was proposed on the basis of the presence of both the acidic and alkali functionalities in the molecule of **1**.

For the relative stereochemistry of **1**, a cofacial orientation of H-1, H-5, H-9, and H_3 -18 on one side of the fused ring system was determined by NOE signals of H-5 with H-1, H-9, H_3 -18 (Fig. 3) in the NOESY spectrum. The orientation of H_2 -14, H-15 and H-20 on

Fig. 3. Main 2D NOESY correlations (between the pink double arrowed hydrogens) of **1** and **2**.

another side of the fused ring system was deduced from the 2D NOE signals between H-14a and H-20 and between H-14b and H-15. The above spectroscopic deduction, together with restriction of the highly fused ring system, the relative stereochemical structure of **1** was assigned as shown in Fig. 2. Crystallization of **1** in a mixture of MeOH and H₂O (3:1) afforded a crystal suitable for crystallographic analysis. Follow-up single crystal X-ray diffraction analysis (anomalous Cu K α radiation scattering) assigned the absolute configuration of **1** [Flack parameter, 0.006 (12)], an ORTEP diagram shown in Fig. 4. Thus, the structure of compound **1** was unambiguously determined and named aconapelsulfonine A.

Compound **2**, a white amorphous powder, [α]_D²⁰ -14.6 (c 0.11, H₂O), is an isomer of **1** as indicated by HR-ESIMS and NMR spectroscopic data. When the NMR spectroscopic data of the two compounds were compared (Table 1), replacement of the semi-acetal and acetal carbons in **1** by a lactone carbonyl carbon [δ _C 178.0 (C-12)] and a hydroxy-bearing quaternary carbon [δ _C 80.7 (C-16)] in **2** was observed. This observation suggested that **2** had another seco-napelline carbon skeleton to generate the lactone carbonyl carbon, which was verified by 2D NMR spectroscopic data analysis. Besides the signals (Fig. 2) to prove the structural moiety shared by **1** and **2**, the ¹H-¹H COSY correlation signals H-9/H₂-11 and the HMBC signals from H-9 and H₂-11 to the lactone carbonyl C-12 demonstrated a 12,13-cleavage of the napelline carbon skeleton for **2**. Additionally the ¹H-¹H COSY correlation signals H-13/H₂-14 and the HMBC signals from H₂-14 to C-7, C-8, C-9, C-15 and C-16; from H-15 to C-7, C-8, and C-9; and from H₂-17 to C-13, C-15 and C-16 as well as their chemical shifts proved the presence of a 13,15,16-trihydroxy-17-sulfonate five-membered spiro-ring moiety consisting of C-8 and C-13–C-17 in **2**. Although the expected three-bond correlation signal from H-1 to C-12 was unobservable in the HMBC spectrum of **2**, the H-1 and C-1 resonances in **2** had larger chemical shift values than that in **1** and related compounds [6j], demonstrating that the lactone ring must be form between the oxygen-bearing C-1 and the carboxylic carbonyl C-12 in **2**. The lactone ring formation was supported by the molecular formula as well as a 2D NOE signal between H-1 and H-11a (Fig. 3) in the NOESY spectrum of **2**. Moreover, the NOE signals between H-5 with H-1, H-9 and H₃-18 and between H-6a and H-9 indicated that these hydrogens spatially oriented on the identical side of the fused rings in **2**. The NOE signals between H₂-17 with H-13 and H-15 demonstrated that the three hydroxy groups on the spiro-cyclopentane ring had the same orientation. The NOE signals between H-6a and H-15 and between H-14b and H-20 unraveled

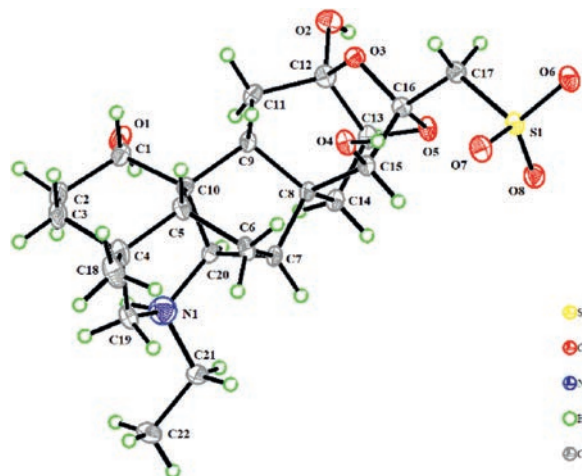
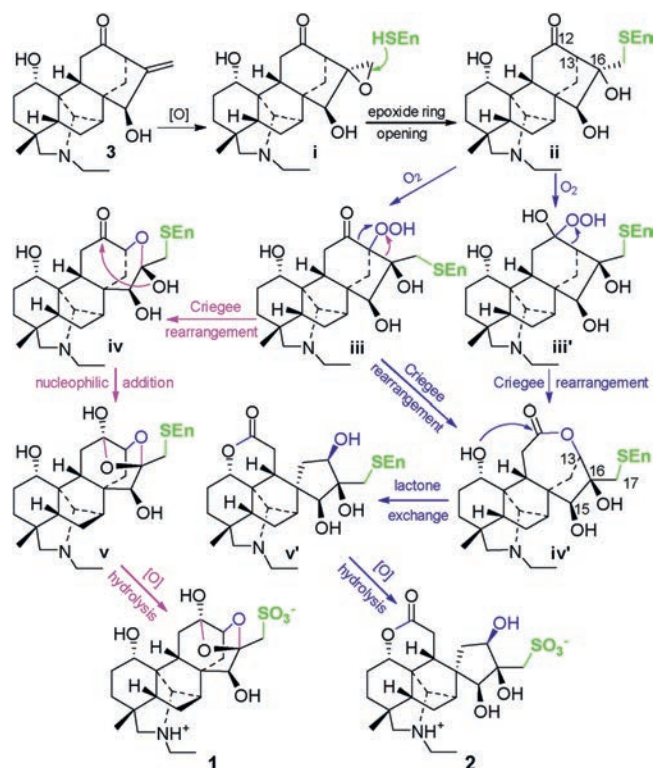


Fig. 4. ORTEP diagram of **1**.

the relative configuration of the spiro-atom C-8 as shown in Fig. 3. Based on the structural similarity and plausible biogenetic origin (see below) of **2** and **1**, the absolute configuration of **2** was assigned to be identical to that of **1**, except that the 13*S* configuration in **1** was reversed to a 13*R* configuration in **2** due to oxidative cleavage of the C-12–C-13 bond and hydroxylation of C-13. Comparison of the experimentally measured CD and theoretically calculated ECD spectra (Fig. S2 in Supporting information) was supportive to the assignment. Accordingly, the structure of compound **2** was elucidated to possess a 12,13-seco-napelline carbon skeleton with functionalities of 12,1-lactone and 13 β ,15 β ,16 β -trihydroxy-17-sulfonate, and designated as aconapelsulfonine B.

Compounds **1** and **2** are zwitterionic sulfonated C₂₀-diterpenoid alkaloids with different seco-napelline carbon skeletons. From their structural features, songorine (**3**, Scheme 1), belonging to the napelline-type C₂₀-diterpenoid alkaloid and co-occurring in this plant [6f], is traced to be the most possible biosynthetic precursor. A double bond oxidation of the precursor affords an epoxide **i**, then the epoxide ring is enzymatically opened to give an enzyme-adding product **ii**. Oxidation of **ii** at C-13 gives the Criegee intermediate **iii**, which undergoes either 13,16- and 12,13-bond migrating rearrangements to generate semi-acetal **iv** and lactone **iv'**, respectively. An intramolecular nucleophilic addition of **iv** would afford **v**, while a lactone exchange reaction of **iv'** would yield **v'**. Subsequently the enzyme-adducts **v** and **v'** would be broken by oxidative hydrolysis to liberate **1** and **2**, respectively. Alternatively, compound **2** may be formed from **ii** via the Criegee intermediate **iii'** of Baeyer-Villiger oxidation and successive rearrangement, lactone exchange, and oxidative hydrolysis. In addition, the enzyme-adducts might be broken to liberate the corresponding diterpenoid alkaloid intermediates at any step of the proposed pathways.

Inspired by clinic application of “Fu Zi” as an analgesic agent [3] and positive responses of our previous work [6g–6j], **1** and **2** were assayed on an acetic acid-induced mice writhing model for the



Scheme 1. Proposed biogenetic pathways of **1** and **2**.

analgesic effect [7] (Supporting information). At dosages of 0.1, 0.3 and 1.0 mg/kg (i.p.), the mice writhing was significantly inhibited in a dose-dependent manner by 58.1%, 63.6% and 70.5% for **1** and 13.8%, 19.3% and 45.5% for **2** [the positive control morphine, 0.3 mg/kg (i.p.), 84.6%].

In conclusion, two analgesic diterpenoid alkaloids with sulfonated zwitterionic properties and undescribed different carbon skeletons were characterized from the water extract of “Fu Zi”, supporting the clinic effects and application protocol of the herbal medicine. Biogenetically the skeletal architectures are proposed to be originated from metabolic oxidation accompanied by the Criegee rearrangements of the different carbon bonds in the napelline-type skeleton. The distinctive structures, along with the analgesic effects, are of interesting for future investigations. The proposed biogenetic routes are informative for biomimetic synthesis and structural modification as well as the involving biosynthetic enzymes (Baeyer-Villiger monooxygenases [8]) of the diterpenoid alkaloids.

Declaration of competing interest

The authors report no declarations of interest.

Acknowledgments

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Appendix A. Supplementary data

Supplementary material related to this article can be found, in the online version, at doi:<https://doi.org/10.1016/j.ccl.2020.09.062>.

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