



Communication

Thiazolyldhydrazone derivatives as inhibitors for insect *N*-acetyl- β -D-hexosaminidase and chitinaseHuibin Yang^{a,b,1}, Huitang Qi^{c,1}, Zesheng Hao^b, Xusheng Shao^a, Tian Liu^c, Qing Yang^{c,*}, Xuhong Qian^{a,*}^a Shanghai Key Laboratory of Chemical Biology, Institute of Pesticides and Pharmaceuticals, School of Pharmacy, East China University of Science and Technology, Shanghai 200237, China^b State Key Laboratory of the Discovery and Development of Novel Pesticide, Shenyang Sinochem Agrochemicals Research and Development Co., Ltd., Shenyang 110021, China^c School of Bioengineering, Dalian University of Technology, Dalian 116024, China

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ABSTRACT

Insect chitinase and *N*-acetyl- β -D-hexosaminidases (Hex) are potential targets for developing new pesticides. Here, a series of thiazolyldhydrazones **I** (with substituted group R₁ at N3) and **II** (with substituted group R₁ at N2) were designed, synthesised and evaluated as competitive inhibitors of *Of*Hex1 and *Of*Chi-h, from the agricultural pest *Ostrinia furnacalis*. Derivatives **I-3d** and **II-3d**, with phenoxyethyl group at R₁, demonstrated the best inhibitory activities against *Of*Hex1 and *Of*Chi-h. Molecular docking analysis indicated that the branched conformation compound **II-3d** ($K_i = 1.5 \mu\text{mol/L}$) formed more hydrogen bonds with *Of*Hex1 than the stretched conformation compound **I-3d** ($K_i = 5.9 \mu\text{mol/L}$). The differences in compounds' binding conformations with *Of*Chi-h explained differences in inhibitory activity of compounds **I-3d** ($K_i = 1.9 \mu\text{mol/L}$) and **II-3d** ($K_i = 4.1 \mu\text{mol/L}$). This work suggests a novel scaffold for developing specific Hex and Chi-h inhibitors.

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Heterocyclic compounds are an important role building block in medicinal chemistry. Thiazolidinones have been extensively investigated over recent decades for a varied range of biological activities, including antifungal [1], antiparasitic [2], antimicrobial [3], antioxidant [4], anticonvulsant [5], anti-HIV [6], anti-inflammatory [7], anti-tuberculosis [8], and anti-tumour activities [9]. Thiazolyldhydrazone derivatives, with a R₁R₂C=N-N= substituent in the 2-position of thiazolidin-4-one, have been studied for their biological properties, such as antiparasitic [2,10], antifungal [11], antiurease [12] and antiviral activities [13].

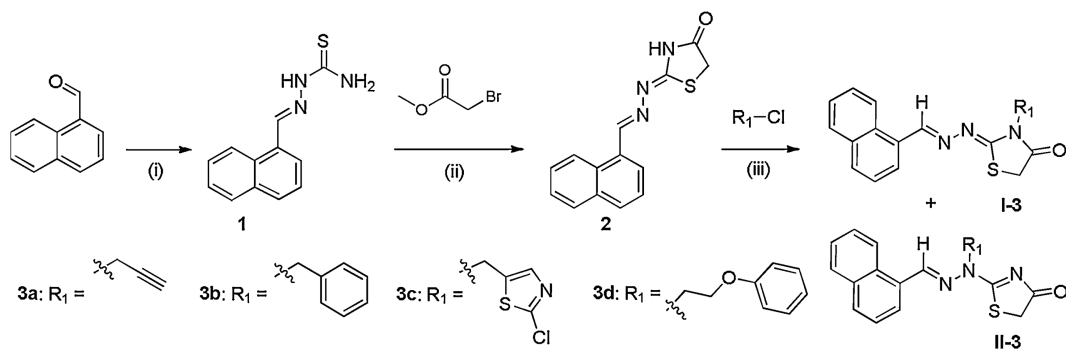
Chitin, a linear homopolymer of *N*-acetyl- β -D-glucosamines and a major structural component of insect cuticles, plays an important role in the molting [14–16]. In the degradation system of insect chitin, two glycoside hydrolase family members, the glycosyl hydrolase family 18 (GH18) chitinase (EC 3.2.1.14) [17] and the glycosyl hydrolase family 20 (GH20) β -*N*-acetylhexosaminidase

(Hex; EC 3.2.1.52) [18], cooperate with each other to facilitate the degradation of chitin. The chitinase can act on random positions of the chitin polymer chain to produce chitooligosaccharides [19,20], while Hex is responsible for hydrolyzing chitooligosaccharides to *N*-acetyl-D-glucosamine during chitin degradation [18,21]. Because of the absence of chitin from vertebrates and higher plants, small molecules inhibitors against chitinase and Hex are promising potential targets for pesticide development.

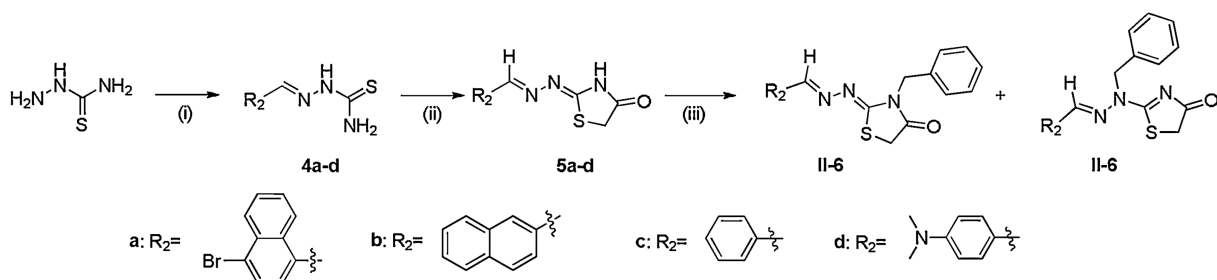
Reported GH20 inhibitors have varied chemical structures. Some are carbohydrate-based inhibitors, such as DNJNAC and its derivatives [22–25], GDL and its derivative PUGNAC [26–29], NTA-glucal [30], 6-Ac-CAS [31,28], NAG-thiazoline and its derivative NMAGT [32,33], LNB-thiazoline [34] and TMG-chitotriomycin [35]. Non-carbohydrate-based GH20 inhibitors include naphthalimides [36], pyrimethamine and its derivatives [37]. Small molecules against GH18 chitinases have been reported. The pseudotrisaccharide allosamidin isolated from the *Streptomyces* sp. [38], cyclopentapeptides argifin isolated from *Gliocladium* sp. FTD-0668 [39], argadin isolated from *Clonostachys* sp. FO-7314 [40], cyclo(L-Arg-D-Pro) from *Pseudomonas* sp. IZ208 [41], chitobiose and chitotriose thiazolines [42], xanthine derivatives [43], and CI-4 [44,45], all exhibit effective inhibitory activities. However, it is

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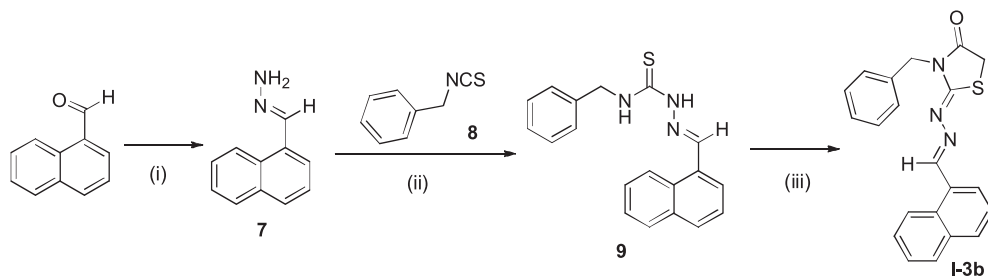
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Scheme 1. Overview of synthetic methods for **3a–d**. Reagents and conditions: (i) thiosemicarbazide, EtOH, acetic acid (cat.), r.t., 15 h; (ii) MeOH, CH₃COONa, reflux, 5 h; (iii) CH₃CN, K₂CO₃, reflux, 5 h.



Scheme 2. Overview of synthetic methods for **6a–d**. Reagents and conditions: (i) R₂CHO, EtOH, acetic acid (cat.), r.t., 15 h; (ii) BrCH₂COOEt, MeOH, CH₃COONa, reflux, 5 h; (iii) chloromethylbenzene, CH₃CN, K₂CO₃, reflux, 5 h.



Scheme 3. Overview of synthetic methods for **I-3b**. Reagents and conditions: (i) H₂NNH₂·H₂O, EtOH, reflux, 6 h; (ii) isothiocyanatomethylbenzene, EtOH, reflux, 3 h; (iii) BrCH₂COOEt, MeOH, CH₃COONa, reflux, 5 h.

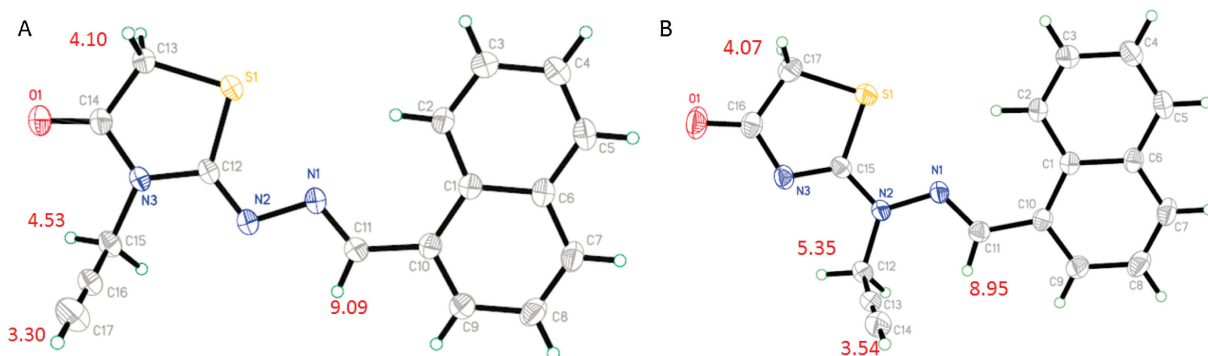


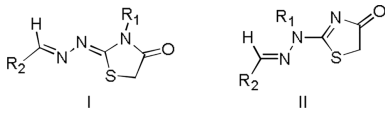
Fig. 1. The crystal structures and chemical shifts of ¹H NMR of compounds **I-3a** (A) and **II-3a** (B).















more challenging to drive these leads into large-scale application because of poor physicochemical properties or complex synthetic chemistry.

OfHex1 [46] and *OfChi-h* [47], from the agricultural pest *Ostrinia furnacalis*, have been demonstrated to function exclusively in chitin

degradation. Previously, our group reported N3 substituted thiazolyhydrazone compounds as *OfHex1* inhibitors [48], and we were surprised to observe the by-product N2 substituted thiazolyhydrazone compounds during the synthesis of N3 substituted thiazolyhydrazone. Only relatively few researches

Table 1
Inhibitory activities of compounds **I** and **II** against *OfHex1* and *OfChi-h*.



R ₁	R ₂	Compd.	Inhibitory rate (% , 10 μmol/L)		Inhibitory rate (% , 1 μmol/L)	
			<i>OfHex1</i>	<i>OfChi-h</i>	<i>OfHex1</i>	<i>OfChi-h</i>
		I-3b	62.6 ± 0.3	62.0 ± 3.2	25.7 ± 3.1	26.0 ± 1.8
		II-3b	72.8 ± 1.7	60.1 ± 3.0	31.5 ± 7.1	27.9 ± 2.3
		I-3c	20.6 ± 6.3	18.0 ± 1.0	6.8 ± 2.9	1.9 ± 1.5
		II-3c	67.7 ± 0.4	23.1 ± 0.7	30.3 ± 0.2	5.3 ± 0.2
		I-3d	65.8 ± 3.4	79.6 ± 0.1	29.0 ± 4.5	38.7 ± 1.2
		II-3d	81.3 ± 0.5	59.2 ± 0.5	42.9 ± 1.6	28.6 ± 2.4
		I-6a	38.3 ± 0.2	62.3 ± 1.8	10.3 ± 3.3	30.1 ± 1.9
		II-6a	67.9 ± 0.2	33.8 ± 3.8	29.9 ± 2.5	12.6 ± 2.0
		I-6b	50.2 ± 3.4	53.7 ± 0.7	18.1 ± 0.9	23.1 ± 1.3
		II-6b	55.8 ± 0.1	11.1 ± 1.9	25.9 ± 0.6	2.3 ± 2.1
		I-6c	37.4 ± 0.9	23.7 ± 1.4	11.2 ± 1.8	7.0 ± 0.6
		II-6c	47.7 ± 6.4	37.8 ± 5.0	20.8 ± 0.3	9.9 ± 1.3
		I-6d	45.5 ± 0.6	32.9 ± 0.0	16.3 ± 0.8	11.0 ± 2.1
		II-6d	44.4 ± 0.1	11.4 ± 0.3	15.5 ± 3.0	1.3 ± 0.8

on the exocyclic N substituted thiazolyldihydrazone compounds have been reported. In a continued effort in search for biologically active molecules, here we report the synthesis of thiazolyldihydrazone derivatives and their inhibitory activities against *OfHex1* and *OfChi-h*. We also investigated the inhibitory mechanisms of these compounds using structure-based molecular docking.

General experimental, synthesis, characterizations of new compounds can be found in Supporting information. The N2 substituted thiazolyldihydrazone **II** were isolated as by-product during the synthesis of thiazolyldihydrazone **I** according to previously reported method [48] (Schemes 1 and 2). Compounds **I-3a**, **I-3c** and **I-3d** had been reported in our previous work [48]. The *R_f* values of thiazolyldihydrazone **I** and **II** is about 0.8 and 0.2 in solvent of ethyl acetate-petroleum ether (1:1), respectively, and the ratio of thiazolyldihydrazone **I** to **II** afforded was approximately 9:1. To confirm the regiochemistry, the N3 substituted thiazolyldihydrazone were synthesised by the route illustrated in Scheme 3. The geometric configuration of double bond and the regiochemistry were further established by X-ray analysis of compounds **I-3a** (CCDC No. 1851329) and **II-3a** (CCDC No. 1912786). As shown in Fig. 1, for thiazolyldihydrazone **I**, the substituted group R₁ was at N3 position of thiazoline ring, and the configurations of the double bonds N(2)=C(12) and N(1)=C(11) were *Z* and *E*, respectively. For thiazolyldihydrazone **II**, the

substituted group R₁ was at the exocyclic N2 position, and the configuration of the N(1)=C(11) double bonds was *E*. When comparing the ¹H NMR data of compounds **II** with **I**, the chemical shift of CH₂ protons of substituted R₁ of **II** (such as **II-3a**, 5.35 ppm) appeared at a higher frequency due to the conjugative effect of the N(1)=C(11) double bonds with naphthalene ring than one of compound **I** (such as **I-3a**, 4.53 ppm).

The inhibitory activities of thiazolyldihydrazone derivatives **I** and **II** against *OfHex1* and *OfChi-h* are outlined in Table 1. Some of the compounds exhibited good inhibitory activities at 1 μmol/L and 10 μmol/L. Our previous work investigating the structure-activity relationships of the thiazolyldihydrazone derivatives **I** against *OfHex1* reported the stretched conformation. Interestingly, the branched conformation of thiazolyldihydrazone derivatives **II**, with aromatic groups at the N2 atom, such as benzyl (**3b**) and phenoxyethyl (**3d**), exhibited potent inhibitory activity. Further studies then focused on the naphthalene ring R₂. The inhibitory activities of the resultant derivatives were weakly influenced by either the substitutes or their position on the naphthalene ring (compare **3b** with **6a** and **6b**). However, replacement of the naphthalene ring with benzene, as in compounds **6c** and **6d**, resulted in a significant inhibitory activity decrease. Gratifyingly, it was found that the thiazolyldihydrazone derivatives **I** and **II** exhibited promising inhibitory activity against *OfChi-h*. In

particular, compounds **I-3d** and **II-3d** were found to display considerable activity against *OfHex1* and *OfChi-h* (Fig. 2).

To further explore possible binding modes of these compounds, molecular docking studies were carried out using the *OfHex1* (PDB code: 3NSN) and *OfChi-h* (PDB code: 5GQB) as templates. The predicted docking results reveal that the thiazolyhydrazone derivatives bind in the active sites of *OfHex1* and *OfChi-h* with different modes (Fig. 3). As shown in Fig. 3A, compound **I-3d** binds the entire active pocket of *OfHex1* via hydrogen bonds, π - π stacking and van der Waals interactions. The naphthalene and thiazolinone rings binds the -1 and $+1$ subsites and stack with Trp524 and Trp490, respectively. The oxygen atom of the thiazolinone ring forms a hydrogen bond with the Glu328 and Glu526 residue. The phenoxyethyl group binds a subpocket formed by Trp483 and Asn489 on Loop478–496, and Gln527 via van der Waals interactions (Fig. 3A). In the docked structure of *OfHex1* in complex with **II-3d**, the N1 and N2 atoms of the linker form hydrogen bonds with the Glu328, Glu526 and Trp490 residues, respectively. The N3 atom forms a hydrogen bond with Glu526, and the oxygen atom of thiazolinone ring forms a hydrogen bond with the Gln527 and Asn489 residues (Fig. 3B). Compound **II-3d** forms more hydrogen bonds with *OfHex1* than compound **I-3d**. The superimposition of compound **I-3d** and **II-3d** reveals that two compounds bind in the -1 and $+1$ subsites of *OfHex1* (Fig. 3C). The

predicted different binding modes of **I-3d** ($K_i = 5.9 \mu\text{mol/L}$, Fig. 2A) and **II-3d** ($K_i = 1.5 \mu\text{mol/L}$, Fig. 2B) could explain the difference in inhibitory activity.

Interestingly, experiments showed that compound **I-3d** ($K_i = 1.9 \mu\text{mol/L}$, Fig. 2C) has better inhibitory activity against *OfChi-h* than compound **II-3d** ($K_i = 4.1 \mu\text{mol/L}$, Fig. 2D). As shown in Fig. 3D, the compound **I-3d** binds the entire active pocket of *OfChi-h* in a stretched conformation. The naphthalene ring is sandwiched by Trp268 and Trp389, and the phenoxyethyl group occupies the hydrophobic patch comprised by Phe184, Thr269 and Leu270. The oxygen atom of the phenoxyethyl group forms a hydrogen bond with the Trp268 residue. The N1 and N2 atoms form hydrogen bonds with Glu308, stabilizing the binding conformations. While the mechanism of interaction of **II-3d** with *OfChi-h* is different from that of **I-3d**, the naphthalene ring occupied the hydrophobic patch comprised by Phe184, Thr269 and Leu270, the oxygen atom of the thiazolinone ring only forms a hydrogen bond with the Leu352 residue (Fig. 3E). The superimposition of compound **I-3d** and **II-3d** reveals that the differences in the compounds' binding conformations with *OfChi-h* explained the differences in their inhibitory activity (Fig. 3F).

In summary, we have designed, prepared and evaluated a series of thiazolyhydrazone derivatives **I** and **II** as potential inhibitors of *OfHex1* or *OfChi-h*. To further explore possible binding modes of

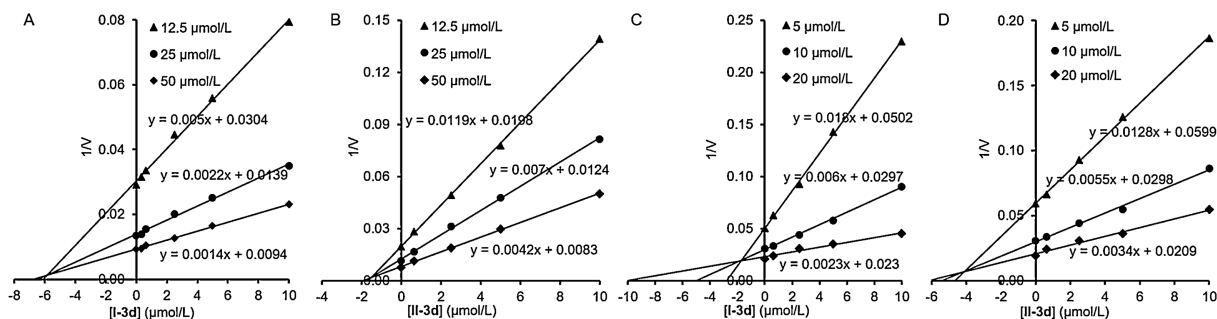


Fig. 2. Inhibitory kinetics of compounds **I-3d** (A), **II-3d** (B) against *OfHex1* and **I-3d** (C), **II-3d** (D) against *OfChi-h*.

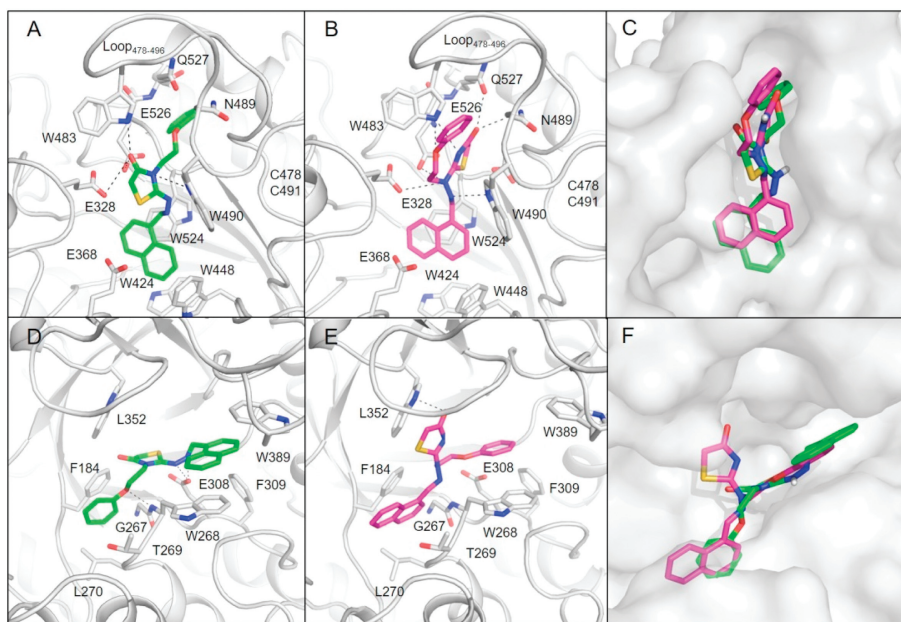


Fig. 3. Proposed binding modes of compounds **I-3d** and **II-3d** to the enzymes *OfHex1* and *OfChi-h* respectively: (A) Binding mode **I-3d** in the active pocket of *OfHex1*. The compound **I-3d** was shown in green. The hydrogen bonds were shown in black dashes. (B) Binding mode of **II-3d** in the active pocket of *OfHex1*. The compound **II-3d** was shown in magenta. (C) Superimposition of compound **I-3d** and **II-3d** in the active pocket of *OfHex1*. (D) Binding mode **I-3d** in the active pocket of *OfChi-h*. (E) Binding mode of **II-3d** in the active pocket of *OfChi-h*. (F) Superimposition of compounds **I-3d** and **II-3d** in the active pocket of *OfChi-h*.

these compounds, molecular docking studies were carried out using the *OfHex1* and *OfChi-h* as templates. Docking results reveal the mechanism of improved inhibitory activity against *OfHex1* and *OfChi-h* of compounds **I-3d** and **II-3d**. This work provides a novel scaffold for developing specific Hex and Chi-h inhibitors.

Declaration of competing interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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Appendix A. Supplementary data

Supplementary material related to this article can be found, in the online version, at doi:<https://doi.org/10.1016/j.ccllet.2019.11.035>.

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