



Communication

Seven-step total synthesis of α -cyclopiazonic acid

Shibin Shi, Kuo Yuan, Yanxing Jia*

State Key Laboratory of Natural and Biomimetic Drugs, School of Pharmaceutical Sciences, Peking University, Beijing 100191, China



ARTICLE INFO

Article history:

Received 12 May 2019
 Received in revised form 18 June 2019
 Accepted 26 June 2019
 Available online 27 June 2019

Keywords:

Indole alkaloids
 Total synthesis
 Cascade cyclization
 Natural products

ABSTRACT

A seven-step total synthesis of α -cyclopiazonic acid is reported from a commercially available 4-bromoindole. Salient feature of the work is the rapid formation of tetracyclic skeleton *via* a bioinspired [3 + 2] annulation to form the C/D rings.

© 2019 Chinese Chemical Society and Institute of Materia Medica, Chinese Academy of Medical Sciences. Published by Elsevier B.V. All rights reserved.

α -Cyclopiazonic acid (α -CPA, **1**), a mycotoxin, was firstly isolated from the fungus *Penicillium cyclopium* Westling, which is often isolated from stored grain and cereal products (Fig. 1A) [1]. In addition, α -CPA is also a nanomolar inhibitor of sarco/endoplasmic reticulum Ca^{2+} -ATPase (SERCA), which are essential for calcium reuptake in muscle contraction and relaxation cycles [2–4]. Importantly, SERCA is a promising target for the development of new drugs against various diseases and insect pests. Thus, α -CPA is one of the few potent, selective and reversible SERCA inhibitors which could act as a lead in drug development.

Structurally, α -CPA is a distinguished 3,4-fused indole alkaloid containing a 6/5/6/5/5 pentacyclic ring system and a highly substituted tetramic acid moiety (Fig. 1A) [1]. Since 2003, a number of highly oxygenated α -CPA-derived alkaloids, such as speradines A–C and aspergillines A–E, were isolated and identified [5–10].

Biosynthetic studies have revealed that β -cyclopiazonic acid (β -CPA, **4**) is the direct biosynthetic precursor of α -CPA (Fig. 1B) [11,12]. Oxidation of **4** by the flavin-dependent monoamine oxidase Mao A (CpaO) to generate a benzylic cation **5** that subsequently undergoes cascade cyclization to result in formation of the C/D rings of **1**.

To date, five total syntheses of α -CPA have been reported (Scheme 1A) [13–19]. The Kozikowski and Natsume groups constructed the C and D rings in a stepwise manner in which the C ring was formed by an intramolecular Michael addition [13,14]. In 2005, the Knight group assembled the C/D rings with high stereocontrol by an elegant carbocationic cascade [15,16]. In

2011, the Scherkenbeck group accomplished the first asymmetric synthesis of **1** by a modification of the Knight synthesis and found that cyclization of the same substrate **9** produced a 1:1 mixture of diastereomers across the C/D ring junction [17,18]. In 2018, the Aggarwal group reported the enantioselective synthesis of **1** by a formal [3 + 2] cycloaddition to form the C/D rings [19].

As part of our ongoing studies towards the concise and efficient synthesis of 3,4-indole alkaloids [20–25], we are attracted by the structure and biological activity of α -CPA and the related natural products. Inspired by the biosynthesis of α -CPA (Fig. 1B), we have discovered an unprecedented acid-catalyzed [3 + 2] annulation of *N*-Me dehydrotryptophan derivative **11** to produce tetracycle **12** by formation of the C/D rings, which enabled a ten-step total synthesis of speradine C (**2**) (Scheme 1B) [26]. Encouraged by this result, we envisioned that the cascade cyclization of the analogous *N*-Ts dehydrotryptophan derivative could be applied to the total synthesis of α -CPA. Herein, we report a concise total synthesis of α -CPA by using this novel acid-catalyzed [3 + 2] annulation.

Retrosynthetic analysis of α -CPA (**1**) was shown in Scheme 2. We envisioned that the tetramic acid residue of **1** could be generated from tetracycle **13** and diketene **14** by Dieckmann cyclization (Scheme 2) [14]. The C/D rings of the key tetracycle **13** could be constructed by acid-catalyzed [3 + 2] annulation of the indole *N*-Ts dehydrotryptophan derivative **15**. In turn, the cyclization precursor **15** could be readily prepared from compounds **16–18**.

Our synthesis commenced with 4-bromoindole **16**, which underwent a Suzuki cross-coupling with *tert*-prenylboronate **17** to give indole **19** as the sole regioisomer (Scheme 3) [27]. Vilsmeier-Haack formylation of **19** followed by *N*-tosylation provided the desired aldehyde **21**. Aldol reactions of *N*-Ts amino acid esters **18** with aldehyde **21** yielded the corresponding α -amino β -hydroxy

* Corresponding author.

E-mail address: yxjia@bjmu.edu.cn (Y. Jia).

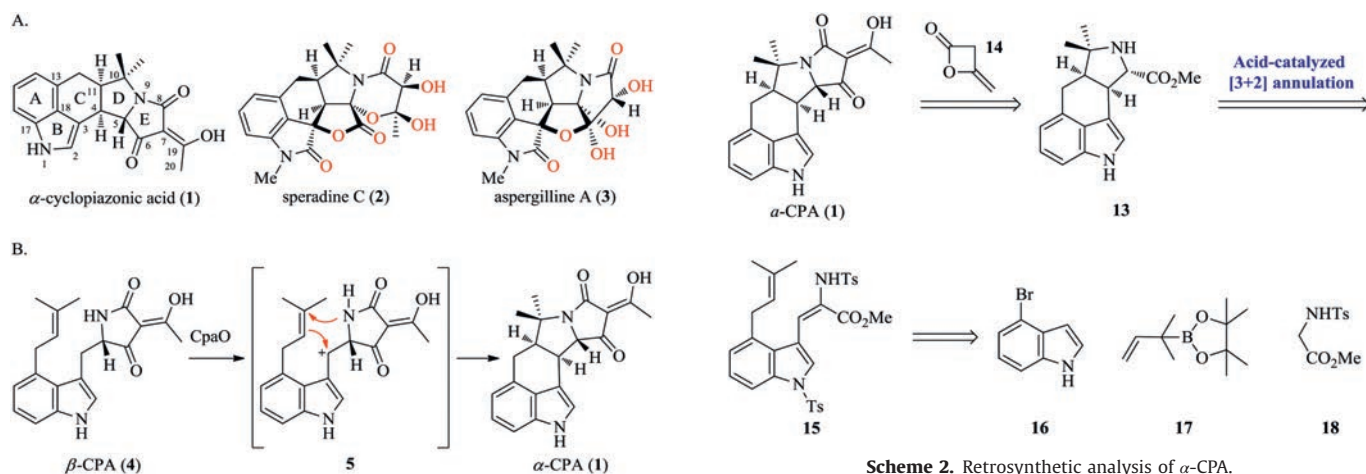
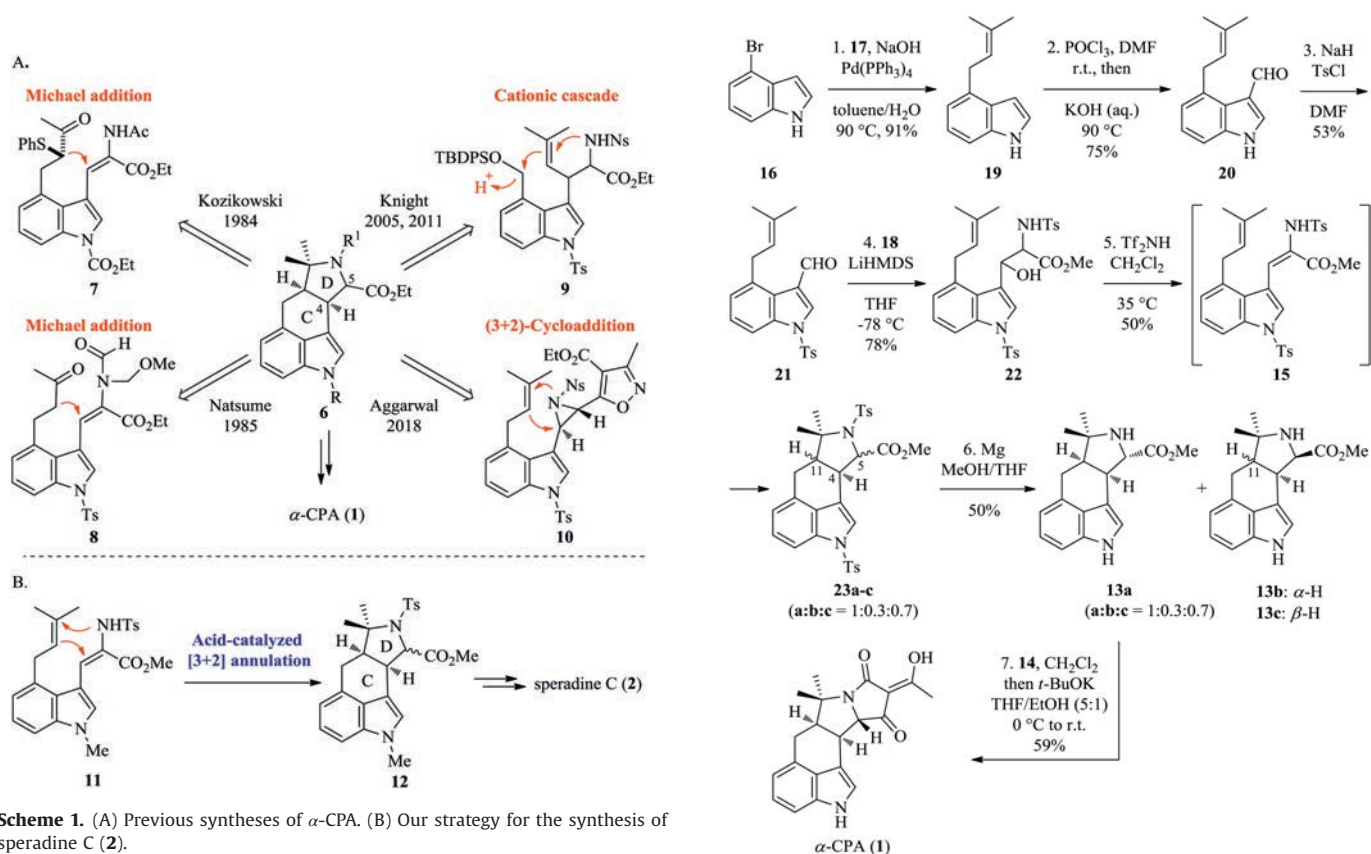


Fig. 1. (A) Structures of α -cyclopiazonic acid, speradines C, and aspergilline A. (B) Biosynthesis of α -CPA.



ester **22** as a mixture of *syn/anti* diastereoisomers ($dr = 1:1$), which could be separated and identified by careful column chromatography. Considering that compound **22** would then be converted to dehydrotryptophan derivative **15** that has no chiral center, compound **22** could be directly used for next step without separation.

With the α -amino β -hydroxy ester **22** in hand, we then investigated the critical [3+2] annulation. To our delight, compound **22** could be readily converted to the desired tetracycle **23a–c** ($dr = 1:0.3:0.7$ at C-5 and C-11) through dehydrotryptophan derivative **15** under the optimal reaction conditions (0.2 equiv. of TiF_2NH in CH_2Cl_2 at 35°C) [26]. Tetracycles **23a–c** were very hard to be separated by column chromatography, and only small amount of pure **23a** was obtained.

Having successfully constructed the C/D ring system, we turned our attention to the synthesis of α -CPA (**1**). At this stage, we needed to remove the two Ts groups in **23**, however, it proved exceptionally difficult, especially for the pyrrolidine N-Ts group. After several trials, removal of the two Ts groups in **23a–c** with a large excess of amount of Mg in MeOH/THF gave amines **13a–c**, and **13a** could be readily separated from **13b** and **13c** by chromatography. The relative configurations of **13a–c** were determined by extensive NMR spectra analysis (the Supporting information) and comparison with the known N-Me-**13a–c** ($dr = 1:0.3:0.7$ at C-5 and C-11) [26]. Finally, treatment of tetracycle **13a** with diketene **14** followed by stirring with *t*-BuOK in THF/EtOH (5:1) led smoothly to give α -CPA

(1). The physical data of our synthesized α -CPA (1) were identical to those reported in the literature.

In summary, we have achieved the total synthesis of α -CPA in only seven steps from commercially available 4-bromoindole (16). The synthesis features an acid-catalyzed [3 + 2] annulation to form the tetracyclic skeleton. This synthesis represents the shortest pathway for the total synthesis of α -CPA to date.

Acknowledgments

This research was supported by the National Natural Science Foundation of China (No. 21871013) and the Drug Innovation Major Project (No. 2018ZX09711-001-005-005).

Appendix A. Supplementary data

Supplementary material related to this article can be found, in the online version, at doi:<https://doi.org/10.1016/j.ccl.2019.06.048>.

References

- [1] C.W. Holzapfel, *Tetrahedron* 24 (1968) 2101–2119.
- [2] R.T. Riley, D.E. Goeger, H. Yoo, J.L. Showker, *Toxicol. Appl. Pharmacol.* 114 (1992) 261–267.
- [3] F. Martínez-Azorín, *FEBS Lett.* 576 (2004) 73–76.
- [4] K. Moncoq, C.A. Trieber, H.S. Young, *J. Biol. Chem.* 282 (2007) 9748–9757.
- [5] X.H. Ma, J.X. Peng, G.W. Wu, et al., *Tetrahedron* 71 (2015) 3522–3527.
- [6] M. Tsuda, T. Mugishima, K. Komatsu, et al., *Tetrahedron* 59 (2003) 3227–3230.
- [7] M. Zhou, M.M. Miao, G. Du, et al., *Org. Lett.* 16 (2014) 5016–5019.
- [8] X. Hu, Q.W. Xia, Y.Y. Zhao, et al., *Chem. Pharm. Bull.* 62 (2014) 942–946.
- [9] H. Zhu, C. Chen, J. Wang, et al., *Chem. Biodivers.* 12 (2015) 1547–1553.
- [10] V. Uka, G.G. Moore, N. Arroyo-Manzanares, et al., *Toxins* 9 (2017) 35–55.
- [11] P.K. Chang, K.C. Ehrlich, I. Fujii, *Toxins* 1 (2009) 74–99.
- [12] X. Liu, C.T. Walsh, *Biochemistry* 48 (2009) 8746–8757.
- [13] A.P. Kozikowski, M.N. Greco, J.P. Springer, *J. Am. Chem. Soc.* 106 (1984) 6873–6874.
- [14] H. Muratake, M. Natsume, *Heterocycles* 23 (1985) 1111–1117.
- [15] C.M. Haskins, D.W. Knight, *Chem. Commun.* (2005) 3162–3164.
- [16] C.M. Griffiths-Jones, D.W. Knight, *Tetrahedron* 67 (2011) 8515–8528.
- [17] C. Beyer, J. Scherkenbeck, F. Sondermann, A. Figge, *Tetrahedron* 66 (2010) 7119–7123.
- [18] W.R. Christian Beyer, K. Woihte, B. Lüke, et al., *Tetrahedron* 67 (2011) 3062–3070.
- [19] O. Zhurakovskiy, Y.E. Turkmen, L.E. Loffler, et al., *Angew. Chem. Int. Ed.* 57 (2018) 1346–1350.
- [20] H. Qin, Z.R. Xu, Y.X. Cui, Y.X. Jia, *Angew. Chem. Int. Ed.* 50 (2011) 4447–4449.
- [21] D. Shan, Y. Gao, Y.X. Jia, *Angew. Chem. Int. Ed.* 52 (2013) 4902–4905.
- [22] L. Li, Q. Yang, Y. Wang, Y.X. Jia, *Angew. Chem. Int. Ed.* 54 (2015) 6255.
- [23] H.C. Liu, X.W. Zhang, D. Shan, et al., *Org. Lett.* 19 (2017) 3323–3326.
- [24] J.B. Lv, B. Wang, K. Yuan, Y. Wang, Y.X. Jia, *Org. Lett.* 19 (2017) 3664–3667.
- [25] H.C. Liu, Y.X. Jia, *Nat. Prod. Rep.* 34 (2017) 411–432.
- [26] H.C. Liu, L.J. Chen, K. Yuan, Y.X. Jia, *Angew. Chem. Int. Ed.* 58 (2019) 6362–6365.
- [27] J.W. Clary, T.J. Rettenmaier, R. Snelling, et al., *J. Org. Chem.* 76 (2011) 9602–9610.