



Chinese Pharmaceutical Association
Institute of Materia Medica, Chinese Academy of Medical Sciences

Acta Pharmaceutica Sinica B

www.elsevier.com/locate/apsb
www.sciencedirect.com



REVIEW

Novel liquid-based approaches for transmucosal drug delivery

Daniëlle van Staden*, Hendrik J.R. Lemmer, Wilna Liebenberg,
Josias H. Hamman

Centre of Excellence for Pharmaceutical Sciences, North-West University, Potchefstroom 2531, South Africa

Received 28 February 2025; received in revised form 30 June 2025; accepted 16 July 2025

KEY WORDS

Clinical translation;
Double-emulsions;
Eutectic systems;
Ionic liquid formulations;
Liquid crystal;
Nanoemulsion;
Self-emulsifying drug
delivery system;
Stimuli-responsive;
Transmucosal

Abstract Advancements in drug discovery, such as artificial intelligence, computational technology, and combinatorial chemistry, have led to numerous new promising drug candidates. However, most still fail to reach the market due to poor physicochemical properties, resulting in off-target or toxic effects. As a potential solution, non-invasive and targeted drug delivery at mucosal surfaces can deliver drugs directly to therapeutic sites to avoid off-target effects, decrease required drug doses, bypass hepatic first-pass metabolism, and circumvent uncontrolled drug release. Liquid-based drug delivery systems have advanced tremendously over recent years, with novel dosage forms such as ionic liquids, liquid crystals, stimuli-responsive phase transforming liquid systems, nanoemulsions, double-emulsions, self-emulsifying delivery systems, and eutectic systems being developed. These systems hold vast promise for transmucosal drug delivery as liquids generally provide superior spreadability compared to solid dosage forms, and some liquid systems provide prolonged mucosal retention times compared to conventional solutions. However, liquid dosage forms present regulatory challenges such as preservation, sterility, and stability requirements. This review discusses novel liquid-based formulation approaches to cross the ocular-, nasal-, oromucosal-, and vaginal mucosal barriers emphasizing advanced liquid drug delivery systems and the progress made toward clinical translation as a platform for future transmucosal liquid-based dosage form development.

© 2026 The Authors. Published by Elsevier B.V. on behalf of Chinese Pharmaceutical Association and Institute of Materia Medica, Chinese Academy of Medical Sciences. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

This article is part of special issue entitled: Hot Topic Revs in Drug Delivery (II) published in Acta Pharmaceutica Sinica B.

*Corresponding author.

E-mail address: dvanstaden711@gmail.com (Daniëlle van Staden).

Peer review under the responsibility of Chinese Pharmaceutical Association and Institute of Materia Medica, Chinese Academy of Medical Sciences.

<https://doi.org/10.1016/j.apsb.2025.12.042>

2211-3835 © 2026 The Authors. Published by Elsevier B.V. on behalf of Chinese Pharmaceutical Association and Institute of Materia Medica, Chinese Academy of Medical Sciences. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

1. Introduction

The drug discovery process places immense focus on finding new lead molecules to treat various diseases¹. Initially, drug discovery was a serendipity-based practice which evolved into a competitive modern science employing sophisticated technologies such as computer-aided design of drugs, combinatorial chemistry, establishing relationships between drug activity and molecular structure, high-throughput screening and artificial intelligence^{1–4}. Despite these advanced technologies, safety, unfavorable physicochemical properties, poor membrane permeation, and insufficient therapeutic action frequently cause the elimination of drugs from further investigation during the final stages of clinical investigations⁵. This manifests a bottleneck effect impeding the translation of potent drug entities from clinical trials to the market as these compounds exhibit toxicities due to uncontrolled dosing or unfavorable off-target effects and low bioavailability^{1,5,6}. This emphasizes that the therapeutic efficacy of highly promising drug entities is often hindered by insufficient and nonselective drug delivery⁷. Therefore, drug administration at specific locations for targeted delivery to reach therapeutic sites has gained increased scientific attention as a strategy to overcome uncontrolled systemic dosing, avoiding systemic toxic effects and reducing the dose needed for therapeutic efficacy while drastically decreasing side effect manifestation⁷. However, localized drug administration is a challenging field of drug delivery because vehicles or systems should be tailored to mediate the crossing of biological barriers^{8,9}.

Mucus is a distinct physical barrier to drug delivery across mucosal surfaces as it protectively covers the epithelia of drug administration sites such as the ophthalmic membranes, nasal cavity surfaces, oromucosal linings, and vaginal mucosal tissues¹⁰. Interestingly, the mechanical characteristics and chemical composition of mucus vary between different physiological sites as they perform different functions¹¹. Additionally, chemical fluctuations like hormonal changes can also influence the structural and chemical capacity of mucus, implying that mucus is a constant yet evolving barrier to drug delivery^{12,13}. Mucus can be described as a complex gel-resembling mixture harboring mucin proteins, lipids, cells, ions, pathogens, and immune components^{1,13}. Due to the gel-resembling nature of mucus, liquid-based formulations have gained significant scientific attention attributed to advantages such as the ability to cover large surface areas, swift drug absorption, mucoadhesive properties, and reduced tissue irritancy potential when compared to solid formulations¹⁴.

Conventionally, liquid dosage forms refer to a specific type of preparations used in the field of drug delivery where the drug is solubilized or dispersed in an appropriate liquid vehicle¹⁵. Examples include solutions, various types of emulsions, suspensions, elixirs, drops, tinctures, syrups, sprays, aromatic water, liniments, and aerosols^{15,16}. These liquid dosage forms were reserved for aqueous soluble drugs with acceptable organoleptic properties and predominantly intended for administration in pediatric patients^{15–17}. However, modern liquid-based drug delivery has evolved from emulsions to nanoemulsions, pickering emulsions stabilized by nanotechnology, multiple-emulsions, and innovative formulation techniques such as spontaneous or self-emulsification^{18–22}. While ionic liquids, liquid crystals, and eutectic mixtures form part of a rapidly expanding field aimed at improving the aqueous solubility of drugs^{23–25}. Therefore, liquid-based drug delivery can also provide a platform to optimize lipophilic drug delivery as the literature reports that 70%–90% of most newly created drug entities are inherently lipophilic^{26–28}.

Approximately 22% of marketed dosage forms are liquid-based with a significant expected increase within the near future due to the registration of numerous vaccines, drug–antibody conjugates, and monoclonal antibodies in liquid-type formulations^{29–32}. Liquid formulations require detail-orientated design approaches to compensate for biopharmaceutical factors and physicochemical properties that do not present challenges in solid dosage forms such as preservatives and stabilizers^{29,33}. This is of particular importance when considering localized trans-mucosal drug delivery from liquid-based drug delivery systems to facilitate the crossing of drug molecules across mucosal surfaces³⁴. Additionally, liquid formulations intended for ophthalmic, intra-muscular, inhalation, or intranasal purposes should comply with specific sterility and stability requirements²⁹. Liquid-based formulations have garnered significant scientific attention over the past decades with a clear interest in specific liquid-based drug delivery systems such as nanoemulsions, self-emulsifying drug delivery systems (SEDDSs), double-emulsions, stimuli-responsive liquid systems with sol-gel transition, ionic liquid-based systems, liquid crystals, and eutectic mixtures. Each of these liquid-based drug delivery systems holds unique advantages and limitations to crossing mucosal surfaces at different sites of administration as thoroughly considered by this review.

2. Mucosal barriers to drug delivery

2.1. The composition of mucus on surfaces at different physiological sites

The literature describes the mucus layer on biological surfaces as hosting the most immune cells of any bodily site and it functions as a barrier to the passage of xenobiotics such as drug molecules, pathogens, and foreign objects^{35,36}. Mucus can be described as a viscoelastic fluid network maintained by mucus secretion from specialized goblet cells, submucosal glands, or secretory cells found in the underlying epithelia^{37,38}. Mucus protectively coats organs and glands interacting with the external environment by trapping, degrading, and washing away pathogens or foreign objects while ensuring hydration and lubrication of mucosal surfaces^{37,39,40}. Regarding structure, mucus presents a heterogenous matrix harboring mobile subunits that are loosely arranged relative to one another⁴¹. This is also broadly referred to as the macrostructure of mucus as particles, drugs, therapeutic carriers and pathogens exceeding 200 nm generally cannot traverse through the macrostructure to reach the underlying epithelia⁴¹. The microstructure of mucus refers to the mesh-like subunits established by mucin networks to entrap foreign substances. Due to the fine filtering capacity of mucin networks, the literature reports that nanocarriers between the range of 10–200 nm should theoretically be able to travel *via* channels within the mucin network^{42,43}. Hence, larger particles utilizing the passages of the mucosal macrostructure to cross mucosal barriers can take longer to reach the underlying epithelia, exposing these particles to a higher risk of entrapment followed by expulsion during mucosal shedding⁴³. However, Lai's group⁴³ have observed that utilizing relatively lower molecular weight chains of polyethylene glycol (PEG) to modify nanoparticles rendered permeation enhancement compared to unmodified nanoparticles. Their permeation experiments involving the crossing of cervical mucosal networks revealed that larger PEGylated nanoparticles (approximate diameter of 200 and 500 nm) displayed more rapid permeation than

smaller PEGylated nanoparticles (approximate diameter of 100 nm). Importantly, this study signifies that optimized physical properties such as droplet/particle size and tailored chemical characteristics like surface chemistry can facilitate the crossing of larger particles despite the reported mucin mesh size of 10–200 nm for cervical mucus⁴³.

Mucosal membranes exposed to the external environment, such as the ocular surfaces, nasal cavities, oromucosal region, and female reproductive tract, are considered promising physiological drug delivery sites due to advantages like targeted drug delivery³⁹. Despite the filtering nature of mucus, which allows nutrient, gas, water, and hormonal exchange, the passage of most pathogenic organisms and drugs is restricted *via* adhesion and steric barrier formation³⁹. Mucus attains its biophysical and rheological properties from water as its main component, which accounts for up to 95% of its total weight⁴⁴. Regardless of the mucosal site pathogens, lipids, proteins, salts, discarded cells, and macromolecules are entangled within the crosslinked, filter structure provided by mucins³⁸. Importantly, the composition of mucus differs between mucosal surfaces located at divergent anatomical areas despite providing equivalent protection to the covered epithelium^{11,39}. For instance, clearance of ocular mucus occurs every 5 to 10 min to remove potential irritants compared to the vaginal mucus lining that thins and thickens as governed by hormonal changes throughout the menstrual cycle^{12,39}. The turnover rate for mucus production can be accelerated upon exposure to toxic substances or irritants to mediate successful removal³⁹. Fig. 1 illustrates the mucosal layers on anatomical sites considered by this review, highlighting the complexity required for developing drug delivery systems for transmucosal drug delivery.

Apart from the unique composition of mucosal sites observed in Fig. 1, each mucosal surface forms part of an organ that performs different bodily functions and necessitates different physiological environments to optimally fulfil its protective capacity. As presented in Fig. 2, the mucosal clearance rate differs between anatomical and physiological sites and can significantly impact drug delivery, as this intrinsic dynamic process can hinder the

crossing of drug particles to the underlying epithelia. As a rule of thumb, the mucosal turnover rate is linked to the thickness of mucus layers³⁸. Hence, thicker mucus layers will generally present slower mucosal turnover rates to maintain the equilibrium between mucus shedding and secretion in preserving the efficacy of the mucosal barrier⁴⁵. One should also bear in mind that the thickness of mucus can be influenced by diseases or as a response to irritant or toxin exposure to guard the underlying epithelia⁴⁶. Generally, mucus unaffected by diseases or the environment can be described as either loose or firm mucus³⁸. Loose mucus refers to mucosal surfaces of the respiratory tract, stomach, small intestine, and colon. Loose mucus is easily cleared by shear or suction mechanisms due to its fibrous composition, causing water retention and leading to thicker mucosal coatings³⁸. On the other hand, firm mucus is a synonym for membrane-bound mucus. Membrane-bound mucus is not easily cleared by shear or suction mechanisms, as this mucus is anchored to the underlying tissue *via* a transmembrane domain to line mucosal surfaces of the eyes and nose³⁸. Moreover, membrane-bound mucus harbors mucins comprising high molecular mass proteins of up to 40 MDa and mucin monomers of up to 0.5 MDa³⁵. Membrane-bound mucosal barriers are capable of regulating the immune response as a responsive and dynamic physiological component closely affiliated with adaptive and innate immune reactions⁴⁷. Due to its dynamic nature, mucus presents a formidable barrier to drug transport as influenced by physicochemical characteristics such as pH, ionic strength, charge, viscoelasticity, and pore size of different mucosal surfaces^{13,37,44,46,48,49}. The literature reports numerous formulation strategies to tailor drug delivery vehicles to optimize mucosal barrier crossing *via* ocular-, -nasal-, -oromucosal-, and vaginal administration.

2.2. Mucoadhesion as a means for drug molecules to cross mucosal barriers

Mucoadhesion is a strategy that can be used to anchor a drug delivery vehicle to mucosal surfaces to achieve site-specific drug

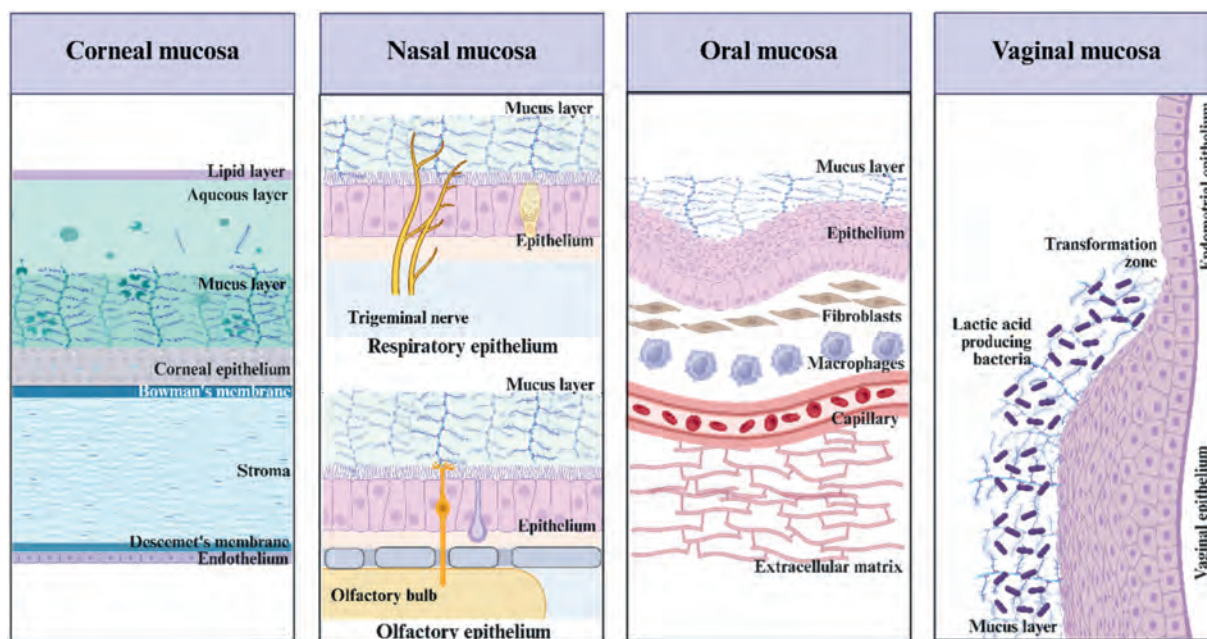


Figure 1 Visual representation of the corneal-, nasal-, oral-, and vaginal mucosa. Created with BioRender.

		Pore size	pH	Clearance rate	Mucins	Temperature	Viscosity	Thickness	Microbiome
Corneal mucosa		13 550 nm $\pm$ 50 nm	13 7.3 – 7.6	53 2 – 3 min (tear film)	13, 56 MUC2 MUC5AC MUC5B MUC6	32.8 – 35.4 $^{\circ}$ C	13 0.97–2.33 mPa.s Literature is inconclusive	64 $\pm$ 3 μ m	69 Coagulase-negative <i>Staphylococcus</i> , <i>Corynebacterium</i> , <i>Propionibacterium</i> , and <i>Streptococcus</i>
Nasal mucosa		13 150 nm $\pm$ 50 nm ²	13 5.5 – 6.5	54 15.53 $\pm$ 5.57 min	13 MUC2 MUC5AC MUC5B MUC19	58 $\pm$ 34.0 $^{\circ}$ C	13 1.8 $\pm$ 1.7 Pa.s	65 10 – 15 μ m	70 <i>Actinobacteria</i> , <i>Firmicutes</i> , and <i>Proteobacteria</i>
Oral mucosa		50 1.5 – 3 nm	51 6.28 $\pm$ 0.36	55 Varies according to location in oral cavity	13 MUC5B MUC19	59 36.11 – 37.56 $^{\circ}$ C	62, 63 Impacted by age, disease, and hydration	66 Buccal: 500 – 800 μ m Mouth floor: 100 – 200 μ m	71 Over 700 bacterial species
Vaginal mucosa		13 20 – 200 nm	13, 52 3.8 – 8.2 Hormonally regulated	12 Hormonally regulated	13 MUC2 MUC5B MUC6	60, 61 $\pm$ 37.0 $^{\circ}$ C	13 Hormonally regulated	67, 68 1.18 $\pm$ 0.20 – 3.50 $\pm$ 2.06 mm Influenced by hormonal fluctuations, age, and birthing method	72 <i>Lactobacillus</i> dominated

Figure 2 Comparison of the corneal-, nasal-, oral-, and vaginal mucosa^{13,39,50–72}. Created with BioRender. Nasal mucosa image provided by Servier Medical Art (<https://smart.servier.com/>).

delivery⁷³. Additionally, mucoadhesion prolongs contact time between mucosal surfaces and drug delivery vehicles to avoid mucosal clearance and thereby increase drug absorption³⁹. Mucoadhesion is initiated during the contact stage of drug vehicle after administration by capitalizing on the repulsion or attraction facilitated between mucosal surfaces and drug delivery vehicles⁷⁴. Importantly, during the application of liquid-based systems, this is also the stage where activation of lipid-based drug delivery systems like SEDDSs occurs based on the affinity of a liquid for mucus, as this will improve spreadability across mucosal surfaces to optimize mucoadhesion or mucosal penetration^{74–76}. Improved spreadability and wetting are accompanied by enhanced swelling capacity of the mucus layer to allow increased access to the underlying epithelial surface⁷⁴.

Numerous theories are described in the literature to provide insight into mucoadhesive interactions between vehicles and mucus. The “Wettability Theory” considers wettability based on vehicle-mucus affinity during the contact stage^{77,78}. Whereas the “Diffusion Theory” considers the sufficient interpenetration of excipients like polymers to mucin chains to mediate adhesive bonds of a semi-permanent nature during the consolidation stage⁷⁸. According to the “Diffusion Theory”, sufficient solubility of liquids in mucus combined with bioadhesive properties is highly favorable for mucoadhesion. Hence, the structural similarity of liquids to mucus enforces stronger mucoadhesive bonding⁷⁸. As an additional option, the “Fracture Theory” describes the mechanical measurement of mucoadhesion by determining the force required to successfully separate two adhered surfaces. Importantly, despite the frequent use of the “Fracture Theory”, it is only suitable for investigating the resistance to rupture of rigid- or semi-rigid agents, as it does not consider phenomena such as interpenetration of polymers⁷⁸. The

“Electronic Theory” considers mucosal adhesion as mediated by the transfer of electrons between the drug delivery vehicle and mucosal surfaces. Thereby, giving rise to a double-layer maintaining electrical charges between the mucosa and vehicle, leading to attractive force formation within the double-layer⁷⁸. Lastly, the literature also considers the formation of primary and secondary surface binding interactions between drug delivery vehicles and mucus. Primary bonds are adhesions attributed to chemisorption by permanent covalent and ionic bond formation. Secondary bonds are considered more desirable due to their reversible binding capacity, such as van der Waals forces, hydrogen bonding, and hydrophobic interactions⁷⁸.

After considering the theories presented by the literature, it is important to acknowledge that these theories should be viewed as integrative processes at work to establish mucoadhesive interactions between vehicles and mucosal surfaces. For instance, wetting of the mucosal surface can be followed by excipient diffusion into mucosal structures to disrupt mucosal arrangement to allow adhesion or electronic transfer formation to optimize mucoadhesive drug delivery^{78,79}. However, it is important to consider that mucoadhesive bond formation will differ between mucosal surfaces located at different anatomical areas (*i.e.*, rate of mucus turnover, the impact of hormonal changes on cervicovaginal mucus, and ocular tear film)^{13,80–84}, the nature of the incorporated mucoadhesive excipient in the drug delivery system vehicle (*i.e.*, molecular weight, flexibility, crosslinking density, charge, hydration capacity, hydrogen bonding, and concentration)^{78,85–88}, the type of liquid-based formulation^{89,90}, and the physiological environment that the vehicle will be exposed to after attachment (*i.e.*, mucosal immune response, nasal mucociliary clearance, bleeding during the menstrual cycle, and the effect of diseases on mucosal barriers)^{78,91–95}.

2.3. Muco-penetrating vehicles as a strategy for drug molecules to cross the mucus layer

Permeation enhancement strategies to cross mucosal barriers are based on the concept of temporarily reducing the viscosity, adhesion or elastic properties of the mucus layer to create a gateway for drugs into the underlying epithelia⁹⁶. This can be achieved by inflicting dissociation of disulfide bonds, utilizing calcium chelators to counteract mucin compaction mediated by calcium, employing surfactants to impede hydrophobic cross-linking, or *via* mucus dilution mediated by osmolytes⁴⁶. In terms of liquid-based formulations, the inclusion of hydrophilic agents like PEG, mucus-penetrating peptides, ligand conjugation, and mucolytic enzymes can improve mucus layer penetration⁹⁷. However, rapid permeation into the underlying epithelia after mucus disruption is imperative since the mucus production rate will increase to repair the equilibrium between mucosal secretion and breakdown/removal⁴⁶. The inclusion of mucus production regulators has been reported to improve penetration enhancement for absorptive mucosa³⁹. As an example, niflumic acid, classified as a calcium-activated human chloride channel 1 inhibitor, has remarkably reduced mucus production within human airway mucosal cells⁹⁸. However, in terms of transmucosal drug delivery, mucus production regulators have not been extensively explored.

Muco-permeative systems can be classified as either active or passive based on their penetrating mechanism. Passive systems aim to restrict interactions between vehicles and mucus by capitalizing on the slippery surface principle^{41,99}. In the context of transmucosal drug delivery, a slippery surface vehicle refers to the ability of the vehicle to glide through mucus by minimizing adhesion to the mucus itself⁴¹. This mechanism can be described as similar to virus surface properties that mediate inert mucosal surface properties as attributed to increased density of anions and cations to facilitate gliding through mucus⁴⁵. Additionally, PEG-based carriers render mucosal migration *via* the slippery surface principle⁴¹. In contrast to the inertness of passive systems, active systems participate in

mucosal permeation by either changing the properties of the carrier upon mucus entrapment or by altering the properties of mucus. Zeta potential changing carriers are excellent examples of active systems that migrate through mucus by altering the pre-mucosal charge from negative to positive or even from negative to less negative to bypass mucosal entrapment in negatively charged mucus^{41,75}. Zeta potential transitioning from negative to positive can be established by enzymatic cleavage targeting anionic surface-impeded substructures after arriving at the underlying epithelia to prevent reuptake into mucus while ensuring cellular absorption⁴¹. The latter group that renders active permeation *via* changing the three-dimensional mucosal structure generally refers to mucolytic enzyme decorated carrier systems that destroy peptide bond formation between glycoproteins or to carriers harboring free thiol groups intended for disulfide bond cleavage^{41,79,100,101}.

Due to the disruptive nature of muco-permeative systems, it is imperative to consider the impact of these systems on mucosal barrier integrity and local immune responses, especially upon chronic use. For instance, a drug delivery vehicle that irritates the nasal mucosa due to penetrative mucosal crossing mechanisms can render excellent permeation while inflicting compensatory hypersecretion, thereby disrupting the physiological balance¹⁰². This disruption can lead to epithelial barrier dysfunction, which can trigger unwanted immune responses and unpredictable drug delivery^{102,103}. The implications of nasal mucosal barrier disruption are portrayed in Fig. 3.

Hence, it is important to trade off the advantages of rapid muco-penetration against unintended physiological responses, as these responses can negatively impact prolonged drug release at mucosal surfaces. Thereby, implying that in some cases mucoadhesion can be favored above muco-penetration as a less disruptive approach to cross mucosal barriers³⁸. Some sources even consider combining these approaches as a potential solution to optimize mucosal drug delivery^{38,116–118}. However, limited studies have explored or successfully implemented this combined approach.

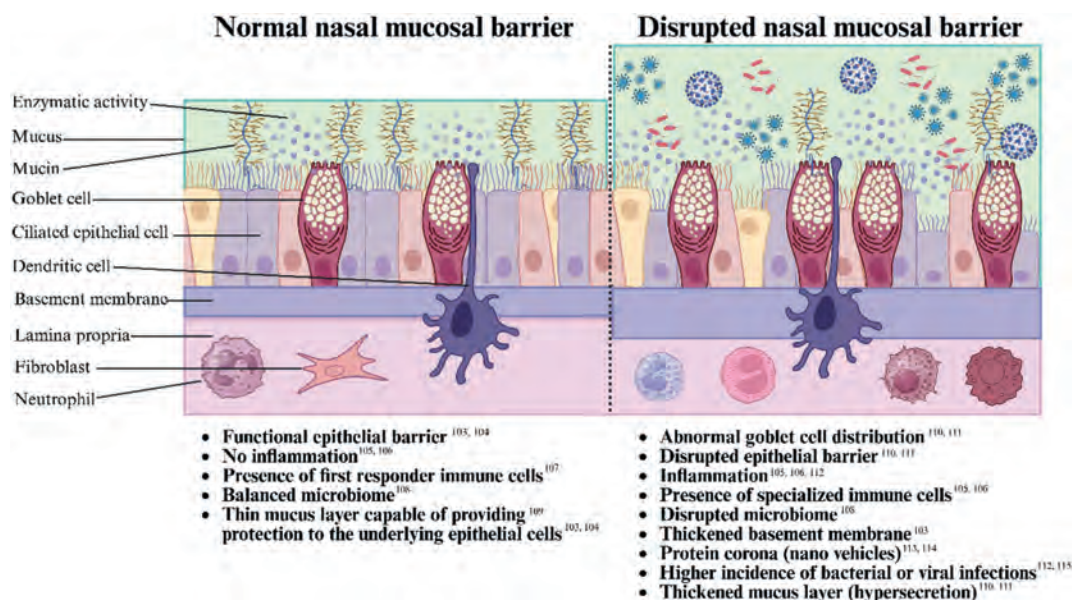


Figure 3 Schematic comparison of factors that influence drug delivery across normal and disrupted nasal mucosal barrier^{103–115}. Created with BioRender.

3. Liquid-based drug delivery approaches

Liquids can be defined as the intermediate phase presenting features that coincide with gases and solids¹¹⁹. It is well-known that scientific knowledge gaps exist regarding the forces that govern structural liquid phase changes^{119–121}. At some point, liquids were considered structurally homogenous vehicles¹¹⁹. However, the classic Bernal presentation of liquids as coherent, homogenous, and inherently irregular structures has evolved due to advances in visualization technologies (*i.e.*, electron microscopy, transmission electron microscopy, and scanning electron microscopy) to clarify experimental, computational, and theoretical discoveries describing liquid structures which is of specific interest to the nanoscale advances made regarding liquid-based drug delivery systems^{122,123}. The advanced visualization techniques confirm that liquid systems present well-defined structures in the bulk phase and at liquid interfaces allowing accurate prediction of co-solvent and surfactant behavior during dosage form development¹¹⁹. In this section, the authors will consider the principles behind individual liquid-based drug delivery systems with an emphasis on the origin of these formulations, excipients that impart specific properties to liquid-based systems, biocompatibility considerations, and safety concerns.

3.1. Evolving emulsion development

Emulsions are considered an essential platform for drug delivery within the broader field of pharmaceutical sciences due to their vast applications in enteral, topical, vaccine, parenteral, and intravenous administration¹²⁴. An emulsion is defined as a dispersion of two immiscible liquids¹²⁵. The immiscibility of liquids is linked to their differing polarities. Generally, emulsions comprise an oil and water phase where one is dispersed in the other with the help of surfactants and co-surfactants. One phase is known as the continuous/external phase, which provides a liquid medium harboring droplets referred to as the dispersed/internal phase. Emulsions can be tailored to form water-in-oil (W/O), oil-in-water (O/W), oil-in-oil (O/O), or even multiple emulsions such as O/O/W, W/O/O, O/W/O or W/O/W by carefully selecting external and internal phases^{21,124}. Apart from composition, emulsions are classified according to their droplet size (*i.e.*, nanoemulsions (<100 nm), miniemulsions (50–500 nm), microemulsions (10–200 nm), and macroemulsions (>0.1 μ m))¹²⁴.

Confusion can arise from the droplet size categorization of emulsions, as some studies classify microemulsions as nanoemulsions due to the nano-sized droplets. Therefore, it is important to emphasize that nanoemulsions differ from microemulsions in terms of droplet morphology, thermodynamic stability, and production methods, which can be taken into consideration when intending to classify an emulsion correctly¹²⁶. Nanoemulsions notoriously present as spherical droplets, whereas microemulsions can manifest droplets of spherical or irregular shapes. Importantly, nanoemulsions are thermodynamically unstable compared to microemulsions. This is attributed to the production method as nanoemulsions are prepared by high-energy procedures to enable minute droplet formation¹²⁶. In contrast, microemulsions can self-assemble despite lower energy input, thereby rendering the formation of larger droplets. However, in the absence of high-energy production methods, microemulsions usually comprise higher surfactant concentrations to achieve emulsion stabilization, leading to concerns about their biocompatibility compared to nanoemulsions that can be stabilized by reduced surfactant

concentrations¹²⁶. Importantly, despite potentially lower surfactant concentrations, due to their small droplet size, nanoemulsions can portray superior membrane permeability eliciting a higher risk of toxicity^{126,127}. Hence, it is recommended to reconcile microemulsions and nanoemulsions by considering each study individually according to the unique attributes of the emulsion and the mucus membrane target site to allow context-dependent and field-specific classification of nanoemulsions¹²⁶.

3.1.1. Nanoemulsions

The first peer-reviewed article referring to a nanoemulsion was published by Calvo and co-workers in 1996¹²⁸. Interestingly, this specific nanoemulsion had a droplet size that exceeded 100 nm. According to the current definition of nanoemulsions, this first reported nanoemulsion would not be considered a nanoemulsion because they are now expected to exhibit droplet sizes smaller than 100 nm¹²⁹. The surfactant phase in nanoemulsions must establish a sufficiently low interfacial tension between the oil and water phases to allow the formation of nanoscale droplets¹³⁰. Due to the very small droplet size of nanoemulsions, they can cross mucosal membranes with more ease compared to microemulsions, which can render improved bioavailability.

Other advantages of nanoemulsions include high drug-loading capacity, solubilizing lipophilic drugs, improved *in vivo* stability, and superior bioavailability. Nanoemulsions also exhibit exceptional targeting capacity which is imperative during the administration of chemotherapeutic drugs to target tumors *via* site-specific release mechanisms and prolonged release to reduce the incidence of side effects. Site-specific release mechanisms refer to techniques like antibody conjugation, ligand attachment, and hyaluronic acid inclusion^{131,132}.

Surface modification of nanoemulsions also presents an attractive alternative to optimize drug delivery. As an example, the addition of cationic polymers can generate a positive surface charge on the negatively charged surface of phosphatidyl-serine-containing surfaces of tumor cells. In terms of transmucosal drug delivery, the addition of the polymer chitosan is an excellent example of a biodegradable, non-toxic cationic polymer reported to enhance transmucosal permeation *via* mucoadhesion established by electrostatic interactions with negatively charged mucus¹³³. Chitosan has also been reported to optimize the targeting of the central nervous system following intranasal administration due to reduced mucociliary clearance and tight junction modulation of the olfactory epithelium¹³⁴. Importantly, the lipid component of nanoemulsions is associated with increased biodegradability and tissue compatibility. However, surfactant choice and the concentration needed to generate nanoscale droplet formation can be seen as a potential drawback of nanoemulsions.

It is documented that more than 200 studies have investigated the toxicity of nanoemulsions and solid-lipid nanoparticles¹³⁵. The overall conclusion was that nanoemulsion-inflicted toxicity was directly linked to the extent of surfactant interactions with cellular membranes instead of droplet size¹³⁵. Therefore, non-ionic surfactants or polymer-surfactant combinations like pluronics, poloxamers, and polysorbates are considered non-cytotoxic and a favorable choice for nanoemulsions¹³⁵. Some studies also reported that higher tolerability to surfactant exposure was observed when the surfactant was bound to nanoparticles compared to the free surfactant state of liquid systems¹³⁵. Moreover, the literature has reported the negative impact of constant surfactant exposure on the gut microbiome, accompanied by inflammation¹³⁶. Apart from surfactant inclusion mediating nanoscale droplet formation, the

formulation technique also contributes while researchers should consider how the technique can impact upscaling possibilities of nanoemulsions since droplet size reduction techniques can be a sluggish process^{76,130}. Nanoemulsions can be produced by high-energy techniques such as high-pressure homogenization, ultrasonication, and microfluidization¹³⁰. On the other hand, low-energy methods like solvent displacement/diffusion, phase inversion temperature technique, or self-emulsification can be used¹³⁰.

3.1.2. Self-emulsifying drug delivery systems

Colin W. Pouton first proposed the idea of SEDDSs in 1985 as a solution to oral lipophilic drug delivery despite the pioneering of spontaneous emulsions during the 1960s^{137,138}. SEDDSs are isotropic mixtures comprising oils, water, and surfactants⁷⁶. Generally, SEDDSs can be categorized according to droplet size, bioavailability, and stability¹³⁹. When the predetermined oil and surfactant mixture is introduced to water nanoscale droplet formation can occur. The initial concept of SEDDSs was to present the drug in an oily concentrate, mostly protected by capsule inclusion to avoid exposure to gastric secretion. Then when the oily concentrate reaches the intestines the peristaltic movements and aqueous environment will mediate spontaneous emulsification¹³⁹. Moreover, the scientific literature reports the utilization of SEDDSs for ophthalmic-, nasal-, oromucosal-, vaginal, and transdermal/topical drug administration^{138,140–143}. SEDDSs intended for dermal application require adding the water phase as part of the formulation process as the oily concentrate will not be exposed to large fluid volumes as in the gastrointestinal tract and secretions like sebum and sweat drastically differ between individuals, implying that dermal SEDDS delivery cannot depend on secretions to provide an aqueous self-emulsifying environment¹⁴⁴. On the other hand, SEDDSs intended for transmucosal drug administration should exhibit mucoadhesive-, mucopenetrating, or a combination of these properties to avoid clearance from mucosal sites while establishing cellular uptake^{22,75,76,142,145–147}. Rapid emulsification within small fluid volumes is also essential as the rate-limiting step for delivery of SEDDSs is the completion of spontaneous emulsification before the mucosal barriers can be crossed²¹.

A recent study investigating enhanced oil recovery in the petroleum industry, where self-emulsification is preferred over high-energy emulsification methods, confirmed that spontaneous emulsification is influenced by the pH of liquids¹⁴⁸. The results presented by Liu's group¹⁴⁸ confirm that a low pH value is associated with an increased tendency toward spontaneous emulsification. Thereby, implying that the natural pH of the specific transmucosal drug administration route can impact spontaneous emulsification. Moreover, spontaneous emulsification is governed by surfactant structure as well as concentration, oil-water phase composition, the addition of non-aqueous solvents, the inclusion of co-surfactants, and the order of excipient addition^{148–150}.

Apart from surfactant structure and concentration, the ratio of oil to water phase, adding non-polar solvents and/or co-surfactants, solution salinity, and temperature can influence spontaneous emulsification. Importantly, lower oil-to-surfactant ratios have been reported to advance spontaneous emulsification. In addition, elevated surfactant concentrations can cause tissue irritation or microbiome disruption¹⁵¹. Ideally, spontaneous emulsification should be facilitated by the lowest possible surfactant concentration. Surfactant concentration also influences droplet size. Monitoring droplet size change over time and measuring the polydispersity index (PDI) of SEDDSs is

imperative to predict ease of spontaneous emulsification, rate of successful drug release, and transport across mucosal membranes¹⁵².

Excipient choice also impacts the class of SEDDS that are produced. For instance, the inclusion of long-chain triglycerides renders the formation of a conventional SEDDS. Whereas medium- and short-chain triglyceride inclusion can give rise to self-micro-emulsifying drug delivery systems (SMEDDSs) with a droplet size range of 100–250 nm and self-nano-emulsifying drug delivery systems (SNEDDSs) formation with a droplet size <100 nm. Some sources differentiate between SEDDSs and SMEDDSs due to the translucent nature of SMEDDSs compared to SEDDSs. However, all sources do not differentiate and often use these terms interchangeably¹⁵³. The formation of SMEDDSs and SNEDDSs can be mediated by surfactants possessing elevated hydrophilic-lipophilic balance (HLB) values combined with certain co-surfactants to improve bioavailability and emulsion stability¹³⁹. SEDDSs improve drug bioavailability by presenting solubilized lipophilic drugs to reduce dissolution rates at absorption sites. Therefore, rapid spontaneous emulsification in small physiological fluid volumes can render nanosized droplet formation at mucosal surfaces to enhance drug absorption. Small droplet formation provides a large interfacial surface, allowing drugs to migrate from the lipid phase to the aqueous physiological environment. Additionally, encapsulation provided by SEDDSs can guard against enzymatic drug degradation and tissue irritation due to drug shielding in small oil droplets¹⁵⁴.

3.1.3. Double-emulsions

Multiple emulsions are a class of liquid-based dosage forms with endless pharmaceutical applications due to their liquid-in-liquid-in-liquid configuration, leading to them being described as emulsions of emulsions. These emulsions differ from conventional emulsions due to the presence of an additional dispersed phase to fulfil the role of a continuous phase. At this stage, double-emulsions are the most explored class of multiple-emulsions in the pharmaceutical sciences. William Seifriz¹⁵⁵ was the first author to report research works evaluating the properties of double-emulsions by considering the impact of oil phase density on the class of emulsion formed in 1925. Advanced visualization techniques have confirmed the formation of numerous classes of double-emulsions such as O/W/O, W/O/W, O/O/W, O/W/W, and W/O/O. In order to stabilize double-emulsions, more than one surfactant should be included to maintain emulsion stability due to the thermodynamic instability at two interfaces, in contrast to the single unstable interface present in conventional emulsions¹⁵⁶. The literature recommends that a surfactant with a high HLB value and a surfactant with a low HLB value should be included to stabilize double-emulsions. Thereby, presenting a strong contrast compared to the formulation of general emulsions stabilized by either one surfactant or a combination of surfactants to establish W/O or O/W emulsion formation based on the facilitated HLB value¹⁵⁶. Double-emulsions present numerous drug delivery possibilities as both lipophilic and hydrophilic drugs and stimuli-responsive polymers can be included^{157,158}. This also implies that co-solvent inclusion can be unnecessary since both an oil and water phase are present for drug and excipient solubilization. Similar to general emulsions, double-emulsions also guard drugs against enzymatic degradation while mediating controlled drug release upon mucosal surface application¹⁵⁹. From a cost-effective perspective, double-emulsions harboring two water phases can be more affordable to produce compared to general emulsions, as the

lipid content of these double-emulsions is reduced compared to classic O/W emulsions¹⁵⁹.

Double-emulsions can also be prepared by spontaneous emulsification if excipient selection is optimized during pre-formulation studies. This involves the preparation of a primary emulsion which is then subjected to the addition of an external phase without any excessive stirring or heat application. A recent publication by van Staden and co-workers²¹ reported capturing the mechanism of spontaneous nano double emulsification for the first time with transmission electron microscopy, as presented in Fig. 4. The authors reported an average droplet size of 92.46 ± 5.52 nm with a PDI of 0.391 ± 0.05 . However, upon transmission electron microscopy imaging, it was discovered that even smaller droplets were harbored within the larger droplets. Thereby, signifying that the dispersed droplets were smaller than 92.46 nm as dynamic light scattering could only detect the larger droplets. These W/O/O nano self-double emulsifying drug delivery systems also report the first double-emulsion intended to incorporate four drugs with strikingly different physicochemical properties²¹. Sustained release of drugs from double-emulsions has been reported¹⁶⁰. However, the study by van Staden and co-workers did not include drug release studies to confirm the sustained release of the four anti-tubercular drugs intended to provide a supplementary treatment option for cutaneous tuberculosis.

3.2. Phase transforming liquid formulations

Drug delivery systems intended for site-specific administration should control drug release and absorption within the *in vivo* situation, while maintaining physical stability during storage periods¹⁶¹. Stimuli-responsive liquid-based drug delivery systems have garnered significant scientific attention since the 1950s after a study by Kuhn and co-workers pioneered the use of smart polymers¹⁶². As an example, the physical phase of such vehicles transforms from a solution to a gel or a gel to a solution once exposed to the expected stimulus (*e.g.* a specific physiological condition) of the targeted site or if a type of external force is applied to the dosage form. External stimuli of either a chemical or physical nature can generate a rapid response by these vehicles,

leading to reversible macroscopic changes during phase transitions¹⁶³. Chemical triggers for phase transition include pH, enzyme exposure, redox potential, and ionic factors. Physical triggers used to modulate phase transition are light exposure, temperature changes, magnetic- or electric field application, and mechanical stress^{163,164}. Phase transitional changes are facilitated by polymers described in the literature as intelligent, smart, stimuli-responsive, or environmentally sensitive, attributed to altered molecular interactions following stimuli exposure^{163,164}.

As the first example, thermo-responsive polymers can be acquired from polymers exhibiting neutral amphiphilic properties like hydrophilic amide carriers, acrylics, or alcohol groups, together with hydrocarbon backbone chains known for their hydrophobicity¹⁶⁵. In terms of temperature-mediated phase transition, stimuli-responsive polymers undergo a coil-to-globule transformation¹⁶⁶. Generally, this transformation is established by the presence of hydrophilic- and hydrophobic functional units that transition from a hydrated to a dehydrated state, rendering a phase transformation from a homogeneously dissolved solution to a gel-resembling heterogeneous biphasic state due to minute temperature changes¹⁶⁵. The literature also reports that the phase transition of polymer-solvent vehicles can be influenced by hydrophobic interactions and hydrogel within the system if exposed to temperatures exceeding the critical solution temperature^{167,168}. At first, the hydrophilic groups of polymers harbored in a homogenous monophasic system swiftly interact with water molecules of mucosal surfaces, rendering the formation of hydrogen bonds to produce hydrated coils occupying random shapes. Upon exposure to temperature changes, hydrophilic functionalities are isolated from water implying that water molecules can no longer gain access to hydrophilic functionalities. This phenomenon initiates polymer re-crystallization presenting nonhomogeneous systems due to dehydration and biphasic transition¹⁶⁵.

Apart from the phase transitions presented in Fig. 5, polymer shape disruption and structure shrinkage facilitate drug release. Hence, it is important to distinguish between the two types of polymer solutions described as lower critical solution and upper critical solution polymeric solutions. Lower critical solution polymeric solutions (*i.e.*, entropy-driven systems) are preferably

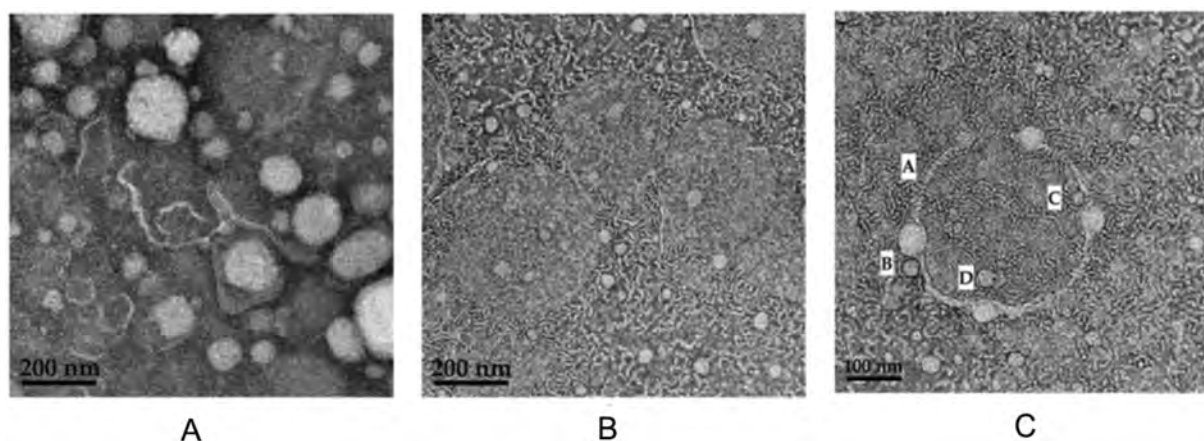


Figure 4 Transmission electron microscopy (TEM) images for (a) primary emulsion (9:9:2) comprising water to internal oil phase to surfactant phase; (b) Self-double-emulsifying drug delivery system (SDEDDS) (9:9:5) containing water to internal oil phase and surfactant phase (9:2) to external oil phase. (c) TEM image of the mechanism of self-double emulsification where the surfactant phase establishes (A) the formation of a membrane structure; (B, C) and then water is surrounded by the surfactant phase leading to small droplet formation in the internal oil phase, and (D) Demonstrates the presence of smaller droplets inside the larger droplet. Reprinted with permission from Ref. 21. Copyright   2023 MDPI.

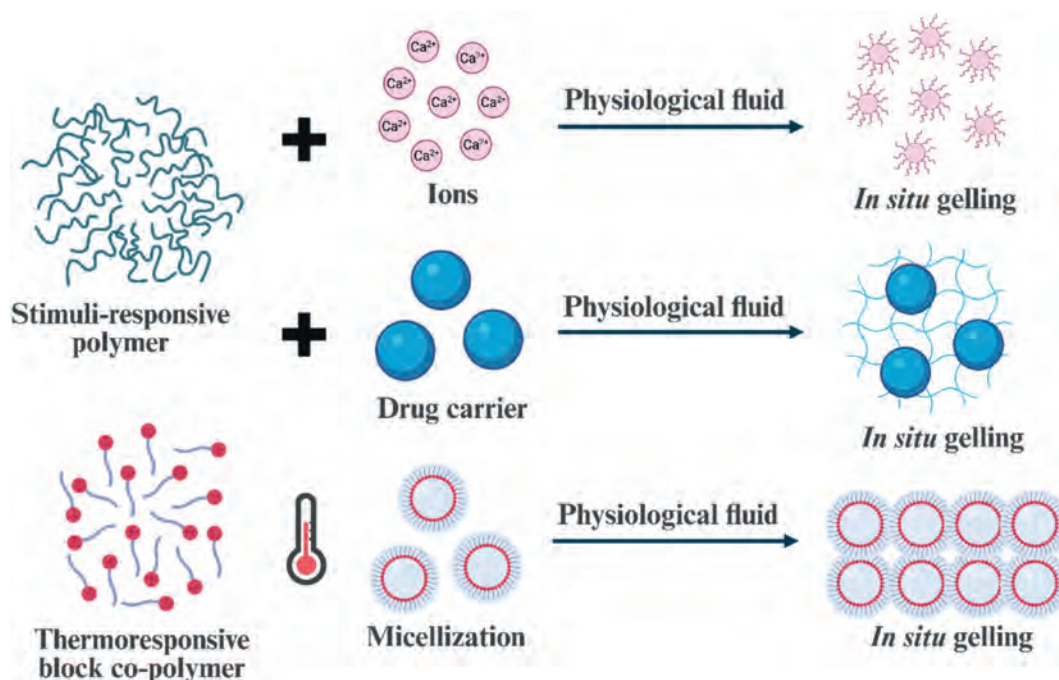


Figure 5 *In situ* gelling transition of stimuli-responsive polymers. Created with BioRender.

used over upper critical polymeric solutions (*i.e.*, enthalpy-driven systems) for phase transitioning liquid systems, as lower critical solutions have a limiting temperature where a binary phase transition occurs and hydrophobic interactions dominate¹⁶⁵. This is also described as “negative thermosensitive polymers” in contrast to “positive thermosensitive polymers” that require cooling before phase transition can occur¹⁶⁵. Importantly, thermo-responsive polymers are generally described as highly biocompatible and biodegradable, which is imperative for transmucosal drug delivery, where irritation at sensitive epithelial sites and microbiome preservation should always be considered¹⁶⁹. Moreover, polymer selection should render permeation enhancement by considering characteristics imparted by polymers such as mucoadhesion, drug release behavior, and viscosity¹⁷⁰. The literature also debates the importance of favoring natural polymers (*i.e.*, pectin, chitosan, gellan gum, and xyloglucan) that exhibit *in situ* gelation above synthetic polymers (*i.e.*, poloxamers (P407 and P188) and Carbopol) due to biocompatibility. However, it should also be considered that natural polymers generally require more than one stimulus to undergo *in situ* gelation compared to synthetic polymers that only necessitate one stimulus to establish phase transition. Thereby, implying that synthetic polymer inclusion can simplify dosage form development as mucosal surfaces can drastically differ in terms of pH and even temperature, and considering both factors can impede successful *in situ* gelation development¹⁷⁰.

Apart from polymer selection, researchers can consider physical and chemical crosslinking techniques to establish *in situ* gelation. Physical methods such as phase transition mediated by pH, temperature, or ionic mediation are preferred due to reduced mucosal irritation risk and simplified formulation techniques¹⁷⁰. Chemical crosslinking techniques involve the addition of additional excipients, apart from polymers, to enable *in situ* gelation, such as organic solvents and co-crosslinkers, which can cause mucosal irritation. An interesting example is provided by

photopolymerization, producing *in situ* gelation via an exothermic reaction occurring during polymerization rendering heat generation potentially inflicting mucosal irritation¹⁷⁰.

3.2.1. Sol-gel transition dosage forms

Regarding transmucosal drug delivery, applying gels initially presented as a solution with a planned stimuli-mediated phase transition can allow simplified handling before and during administration. On the other hand, stimuli-responsive phase transition allows increased retention time at mucosal surfaces to improve epithelial drug absorption¹⁷⁰. Stimuli-responsive polymers display pseudo-plastic rheologic behavior, implying that these excipients will improve the spreadability of sol-gel formulations across mucosal surfaces¹⁷¹. During the application of sol-gel formulations, solution viscosity is reduced as the force of application re-aligns entwined polymeric chains towards the direction of shear applied. Upon introduction to the specific conditions of the physiological environment, the original rheologic properties are re-established to mediate the formation of a viscous gel. In terms of localized, transmucosal drug delivery converting rheological properties to their original form implies that sol-gel formulations can avoid formulation leakage from body cavities like the vaginal channel and impeding mucociliary clearance from the nasal cavity^{170,171}. Thereby, allowing prolonged contact time between mucosal surfaces and liquid drug delivery systems to govern controlled and sustained drug release for localized or systemic therapeutic effects¹⁷¹.

3.2.2. Fluid gels

Fluid gels, also named shear gels, describe the suspension of gel particles in a continuous liquid phase. Shear application or physical disruption of a biopolymer solution during the cooling phase of the gelation process is known to produce fluid gels¹⁷². Thus, rendering paste or fluid resembling properties by fine gel formulations to exist as highly concentration gels suspended in a

liquid matrix. Interestingly, fluid gels and quiescent gels can have identical chemical compositions, but different drug release behavior due to the altered physical properties^{172,173}. Drug delivery from fluid gels can be optimized by exercising control over cooling and fluid rates. Additionally, the concentration and type of biopolymer selected during preformulation can also impact fluid gel properties. Fluid gels exhibit thixotropic behavior upon substantial yield stress application signifying that fluid gels can be applied as gels, but exhibit phase transition by returning to a fluid upon increased shear application followed by recovering to its fluid gel state following force removal^{174,175}. This is of particular importance in transmucosal drug delivery as fluid gels can be easily administered with devices by transpiring into a liquid phase to cover larger mucosal surface areas by spraying devices and then remarkably return to a fluid gel state to promote mucosal surface retention time and improve permeation enhancement. In contrast to delivery in body cavities, fluid gels present the option of drop administration on the corneal surface followed by the visco-elastic gelling of the drops to increase retention time¹⁷⁶. Interestingly, the literature reports that fluid gels can be employed as an emulsion encapsulating vehicle, replace the lipid component of emulsions, and mediate foam stabilization^{177,178}. Moreover, fluid gels swiftly regain their fluid gel consistency after shaking and are known for superior stability properties¹⁷⁵.

3.3. Ionic liquid-based drug delivery systems

3.3.1. Ionic liquids

Ethyl ammonium nitrate was reported by Paul Walden as the first discovered ionic liquid in 1947¹⁷⁹. Despite approximately a century of ionic liquid research, a real interest in ionic liquids only emerged during the past two decades¹⁸⁰. Ionic liquids are a molten salt class harboring cations provided by an organic source and anions from an organic or inorganic source at a low temperature (<100 °C). Ionic liquids possess unique physicochemical properties such as high conductivity, superior chemical and thermal stability properties, and minimal vapor pressure, signifying their clear difference from conventionally used solvents and salts¹⁴. Due to their unique properties, ionic liquids can be used to fulfil the water or oil phase role of liquid vehicles. Moreover, ionic liquids can also be used as structural components to mediate the formation of micelles, microparticles, nanoparticles, and emulsions¹⁸¹. According to the macrocyclic structure of ionic liquids, these liquids can generally be divided into four primary groups (*i.e.*, *N*-alkyl-pyridinium, phosphonium, alkyl-ammonium cations, dialkyl-imidazolium)¹⁸¹. Apart from their macrocyclic structure, ionic liquids can also be distinguished based on their chronological order describing their discovery providing three generations of ionic liquids.

The first generation of ionic liquids comprises cations paired with halide anions. These liquids are known for low melting points, superior thermal stability, and retaining their liquid form upon exposure to a broad temperature range. First-generation ionic liquids are expensive to produce and prone to participating in oxidation and hydrolysis reactions¹⁸¹. On the other hand, cations for second-generation ionic liquids are provided by dialkylimidazolium, alkyl-pyridinium, ammonium, and phosphonium with anions obtained from tetra-fluoroborate and hexa-fluorophosphate inclusion. Second-generation ionic liquids are less susceptible to oxidation and hydrolysis. The second-generation ionic liquids also expanded the customization of ionic liquids to alter properties such as thermal stability, solubility, biodegradability, viscosity, melting point, and hydrophobicity to

obtain ideal physicochemical properties for unique purposes¹⁸¹. Hence, the second-generation ionic liquids evolved into third-generation ionic liquids often used in green chemistry to produce ionic liquids to perform a specific function^{181–183}. Due to the involvement of green chemistry, anions are provided by natural sources like fatty acids and amino acids while cations are mainly obtained from choline to mediate transmucosal drug delivery¹⁴.

Choline-based ionic liquids are known to improve mucoadhesion by shielding mucin glycoproteins. The isolation of negative mucin domains leads to a reduced mucosal viscosity without causing structural damage to mediate mucosal permeation¹⁸⁴. The biological origin of third-generation ionic liquids allows physical strength impediment of mucus by altering the viscosity and elasticity of the mucosal layer to enable the epithelial crossing of drugs. The increased viscosity of ionic liquids not only guides mucosal modulation but also prolongs contact time between mucosal surfaces and ionic liquid-based carriers⁷³. Pairing choline with appropriate anions can also advance mucosal modulation by tailoring the pH and conductivity of liquid formulations⁷³. Ionic liquids can also enhance the solubilization of polymers to furnish vehicles with additional mucoadhesive properties allowing controlled drug release¹⁸⁵. Moreover, innovative dosage forms like ionogels have also been produced *via* the incorporation of ionic liquids¹⁸⁵. Ionogels are described as combining an ionic liquid and a solid continuous phase. These systems can be classified as inorganic, organic, or hybrid inorganic–organic systems as defined by their synthetic route¹⁸¹. Another advantage of ionic liquids is the inherent antibacterial and antifungal capacity provided by anionic groups⁷³. Moreover, drug release within divergent pH environments can be controlled *via* ionic liquid inclusion. Drugs or drug combinations can also be converted into ionic liquids to expertly modulate their physicochemical properties by eliminating the possibility of polymorphism, enhancing aqueous solubility, improving thermal stability, and tailoring dissolution rates¹⁸⁶. Moreover, as demonstrated by Fig. 6, ionic liquids can be encapsulated *via* solvent evaporation, interfacial polymerization, impregnation, and emulsion polymerization¹⁸⁷.

The development of ionic liquids and their applications have surpassed toxicity evaluations as ionic liquids are often branded as “green solvents” generating the misconceived perception that all ionic liquids are safe¹⁸⁸. The literature reports that increased cation alkyl chain length holds a higher risk of toxicity^{188–190}. The mechanism of toxicity is based on alkyl chain insertion, ultimately impeding cellular membrane formation and inflicting apoptosis. Therefore, increased alkyl chain polarity can restrict their capacity to participate in hydrophobic interactions regarding cellular membranes¹⁸⁸. Another possibility includes zwitterionization, where an anionic moiety is inserted at the final section of the cationic alkyl chain^{188,191}. Alternatively, bionic liquids such as choline cations are employed to reduce toxicity profiles¹⁸⁸. As a metabolite of the neurotransmitter acetylcholine and the presence of phosphatidylcholine as a cellular membrane polar head group, choline is considered less toxic compared to the initial cationic sources of ionic liquids. Research also revealed that choline ionic liquids paired with anions provided by acetate, aspartate, chloride, and glutamate produce safer toxicity profiles. Therefore, the selection of the anionic source is imperative when producing choline-based ionic liquids¹⁸⁸.

3.3.2. Liquid crystals

An Austrian botanical physiologist, Friedrich Reinitzer, discovered the liquid crystal phase in 1888. Reinitzer reported two

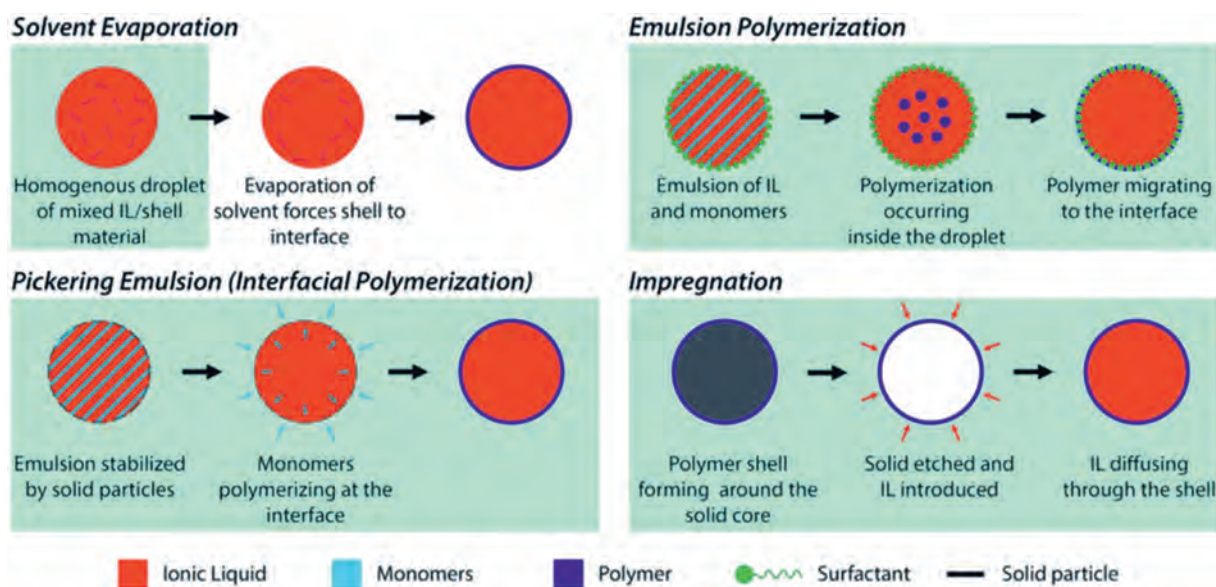


Figure 6 Schematic representations of the different methods used for the preparation of encapsulated ionic liquids. Reprinted with the permission from Ref. 187. Copyright © 2021 American Chemical Society.

melting points for cholesteryl benzoate that he successfully extracted from the root of a carrot^{192,193}. He observed that the solid form of cholesteryl benzoate at 145 °C transpired into a turbid liquid and became transparent at a temperature of 178.5 °C. Thereby, discovering the transition from the solid phase to liquid crystal formation followed by its metamorphosis to the liquid phase¹⁹⁴. These liquid systems simultaneously combine solid and liquid state characteristics by generating a mesomorphic state^{195,196}. In this mesomorphic state, the solid can be described as positively disordered to generate properties such as electrical and optical anisotropy. While the liquid is considered orientally ordered to provide mobility comparable to fluids. Liquid crystalline systems are biocompatible, stable systems that can be prepared cost-effectively to improve aqueous solubility and control the rate of drug release. Additionally, liquid crystalline systems can increase drug potency and therefore enhance pharmacological effects¹⁹⁵. The mesomorphic state can be classified as thermotropic liquid crystals or lyotropic liquid crystals. Thermotropic liquid crystal formation is temperature-dependent and subdivided into cholesteric, nematic, and smectic liquid crystal systems. On the other hand, lyotropic liquid crystals are produced *via* the liquefaction of a drug in numerous solvents¹⁹⁵.

These systems comprise self-assembled amphiphilic molecules generating structures such as cubic, hexagonal, and lamellar mesophases. Regarding drug delivery lyotropic liquid crystals have garnered significant scientific attention as a tailorable drug delivery carrier as the liquid crystallinity of lyotropic liquid crystals is a function of solvent concentration¹⁹⁶. Lyotropic is derived from *lyo* indicating “solvent” and *tropic* is a synonym for “forms” signifying that lyotropic refers to solvent formation. This is attributed to the self-assembling capacity of lyotropic systems as influenced by the concentration-dependent addition of solvents. As a one-dimensional type of lyotropic system, lamellar phases comprise amphiphilic molecule bilayers, thereby separating water and oil phases¹⁹⁷. Hence, interactions between water and oil are prohibited which can improve drug encapsulation. Moreover, bilayer thickness can be controlled by considering the partitioning between different oil phases and water as well as water or drug concentrations present

in the liquid crystalline system. As the second type of lyotropic system, hexagonal liquid crystals refer to surfactant-generated micelles transpiring into advanced cylindrical micelles¹⁹⁶. Aggregation of these cylindrical micelles then facilitates two-dimensional higher-order structures known as the hexagonal phase. The water phase concentration can impact the formation of reverse hexagonal phase or general hexagonal phase formation, similar to the formation of reversed micelles in oil-rich formulations compared to general micelle formation in water-rich formulations¹⁹⁶. Interestingly, polarized light microscopy investigation of hexagonal liquid crystals described a fan-resembling structure. The fan-resembling structure is mediated by the hexagonal arrangement of each cylindrical micelle encircled by six additional cylindrical micelles, where the radius of the divided cross-section is related to the molecular length of the incorporated surfactant¹⁹⁸. Hence, providing a favorable structure to maintain *in vivo* stability, enhancing drug loading, and advancing the delivery of macromolecules. The third type of lyotropic system, known as cubic liquid crystals, refers to crystals formed by a lipid bilayer, with an approximate thickness of 3.5 nm, continuously curving to create channels that avoid aqueous contact¹⁹⁶. Thereby, generating a large interfacial area of approximately 400 m²/g. This liquid crystalline phase is hallmarked as a more rigid structure able to optimize drug release while presenting exceptional drug loading capacity. Moreover, different cubic liquid crystalline phases (*i.e.*, double diamond-, body-orientated-, and gyroid lattice cubic phase) are considered more resistant to instability upon dilution compared to other liquid crystalline phases. However, cubic phases are considered more suitable for lipid drug delivery due to a reduced water presence¹⁹⁶. The multi-dimensional structures of the different lyotropic liquid crystalline phases are presented in Fig. 7¹⁹⁷.

Apart from the multi-dimensional structures of liquid crystalline drug delivery structures, the storage stability of liquid crystals remains imperative as these systems are influenced by pH and temperature changes¹⁹⁷. Normal liquid crystal storage conditions specified at 4 °C where the transition of liquid crystals upon storage has been reported^{197,199}. Emulsions harboring lyotropic liquid crystals have been reported to maintain stability for a 1-year

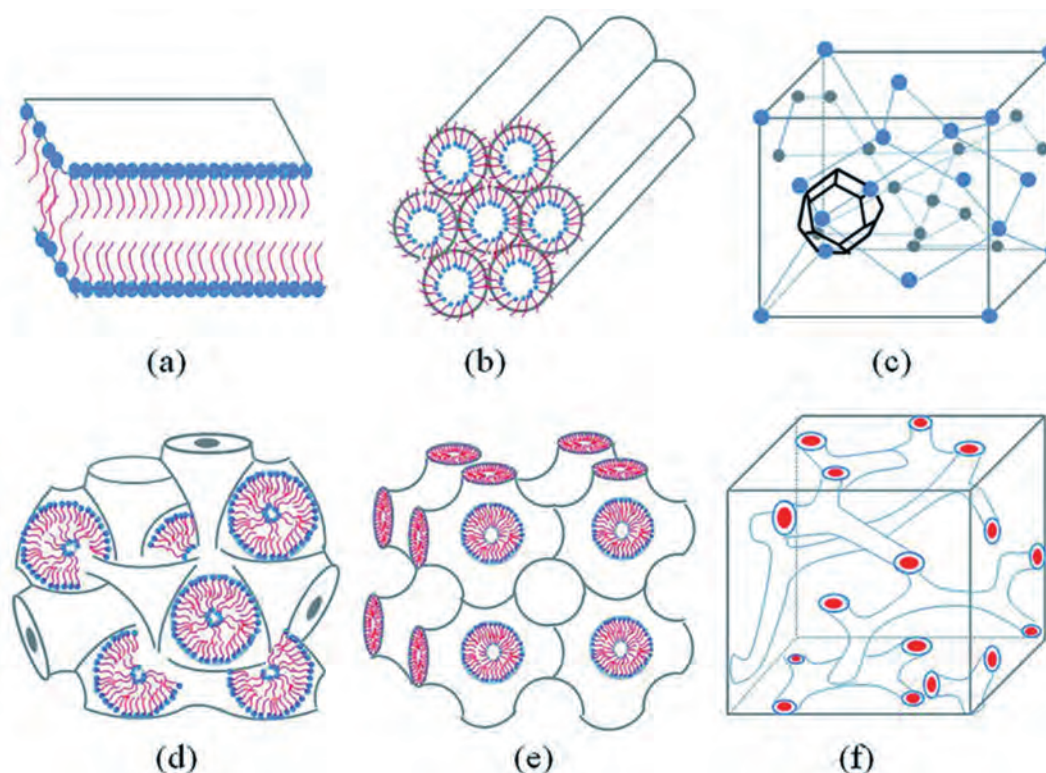


Figure 7 Schematic representation of the lyotropic liquid crystalline phases commonly found in neutral lipid/water systems. (a) Lamellar phase (b) reverse hexagonal phase (c) reversed micellar cubic (d) reversed bicontinuous cubic (e) reversed bicontinuous cubic (f) reversed bicontinuous cubic. Reprinted with permission from Ref. 197. Copyright   2018 American Chemical Society.

storage period with an expected structure decomposition following 18 months of storage¹⁹⁹. A study has demonstrated that storage times of hexasomes generated by liquid crystalline systems can be lengthened in decreased pH conditions^{196,200}. Another challenge accompanying drug delivery from liquid crystalline systems is the initial rapid drug release that can lead to either systemic or local toxicity depending on the route of administration^{196,201}. Generally, rapid drug release is overcome *via* structural drug modification¹⁹⁶. Additionally, the introduction of alkyl chain linkage in the liquid crystalline structures can prolong drug release¹⁹⁶. Regarding toxicity, the excessive use of surfactants, co-surfactants and liquid crystalline systems as a combination presents a higher risk for toxicity¹⁹⁶. However, surfactants are essential components of these systems to improve drug loading capacity. As a potential solution, the literature reports that the inclusion of amphiphilic lipids of a biodegradable and biocompatible nature can potentially decrease the incidence of toxicity¹⁹⁶.

3.3.3. Eutectic systems

Frederick Guthrie named the first eutectic system *eut  xia*, a Greek word meaning “well-melting”, in 1884^{202,203}. Eutectics are notoriously used by various industries with each field naming eutectics differently. Within the thermal energy field, eutectics are known as phase change materials, the metallurgy industry refers to eutectic alloys, green chemistry practices investigate deep eutectic solvents and low transition temperature mixtures, and the pharmaceutical sciences refer to eutectic mixtures²⁰³. Eutectic mixtures present a lower melting point for a homogenous mixture of solids compared to the melting point of individual solid components added in a specific molar ratio as attributed to non-covalent interactions²⁰².

Eutectic systems provide an alternative option for enhancing the aqueous solubility of drugs. Deep eutectic systems are a modern version of eutectic systems that pertain to an advanced melting point depression capacity known for not only lowering the melting point of the mixture to a lower melting point than individual components, but to a melting point lower than room temperature²⁰⁴. The literature describes deep eutectic systems as analogous to ionic liquids as both liquid-based vehicles can be classed as designer solvents with minimal vapor pressure²⁰⁵. However, deep eutectic systems are considered more cost-effective and require simplified preparation techniques while potentially being less toxic than ionic liquids²⁰⁶.

Interestingly, deep eutectic mixtures have superior thermodynamic stability compared to amorphous drug delivery systems²⁰⁴. Eutectic systems are established on the principle of size or shape mismatching between components mediated by weak adhesive interactions between materials of a non-isomorphous nature, rendering similar packing patterns and lattice arrangements to those of parent molecules. Thermodynamically speaking, eutectics can form upon a greater entropy gain compared to an enthalpy gain during molecular ordering. The equilibrium between entropy and enthalpy partially depends on the structure of parent molecules. Additionally, close contact between potential eutectic-forming solid components should be paired with the ability of these components to present mutual solubility when presented in their molten state²⁰⁴. The use of non-isomorphous materials in eutectics is known to be accompanied by a mutual solubility threshold as the added compound can only fit into the lattice of the first agent to a certain extent. When the respective threshold of solubility is reached strain within the corresponding solid solution lattice causes

disorganization. A eutectic phase is then formed by generating an inter-phase boundary between two phases produced during reorganization. Redistribution of atoms occurs at the formed inter-phase boundary of solid solutions. The strain provided by non-isomorphous solids is further enhanced by defective anatomic rearrangement and weak bonding interactions at the inter-phase boundary. Therefore, this process lowers the melting point of eutectic systems due to the heightened thermodynamic functions²⁰⁴.

For example, EMLA® a marketed eutectic system combining prilocaine and lidocaine as local anesthetics drastically improves transdermal permeability and aqueous solubility of the incorporated drugs, demonstrating the potential of these systems for permeation enhancement²⁰⁷. An article by Chu and Yalkowsky investigated the impact of melting point on drug absorption¹⁷⁹. Their findings concluded that the absorption of drugs possessing a lower melting point is less influenced by the incorporated drug dose, revealing that reduced melting points can improve drug solubility and partitioning values²⁰⁸. Hence, eutectic systems can also be considered a potential alternative to supersaturated drug delivery systems. Supersaturated solutions significantly increase flux due to an enhanced driving force which renders permeation enhancement without the inclusion of permeation enhancers. However, supersaturated drug delivery systems are in a metastable thermodynamic state as precipitation of the supersaturated drug can occur²⁰⁹. Hence, by eliminating the risk of drug precipitation,

eutectic systems can present a more stable alternative to supersaturated drug delivery systems.

In terms of the safety and potential toxicity of eutectic systems, there is a general perception that eutectics can be considered non-toxic mixtures as the parent molecules are regarded as green or safe to use. Pharmaceutical regulatory bodies therefore do not consider eutectic systems novel polymorphs and do not require new clinical trials to investigate their toxicity. However, safety concerns involving eutectic systems are being raised in the literature. Specifically, due to the lack of knowledge regarding the type of toxicity that may arise due to the instability observed during the formation of these systems. Moreover, research also revealed that high hygroscopicity of eutectic systems can lead to oxidation or chemical degradation of parent molecules^{210,211}.

3.4. Combining liquid-based drug delivery approaches

Combining liquid-based transmucosal drug delivery approaches has been reported as a strategy to enhance vehicle ability to cross mucosal barriers^{212–215}. However, various physiological factors at specific mucosal sites often hinder mucosal barrier crossing, thereby creating obstacles to effective transmucosal drug delivery⁷³. These challenges are summarized in Table 1^{12,13,17,51,91,216–239} to compare the advantages to the reported challenges/disadvantages of site-specific transmucosal drug delivery.

Table 1 Advantages and challenges/disadvantages of ocular, nasal, oromucosal, and vaginal drug delivery.

Transmucosal route	Advantage	Challenge/disadvantage	Ref.
Ocular	Ease of administration	Blinking reflex	216,218
	Targeting of specific ocular sites	Drug binding to tear proteins	217,218
	Reduced risk of side effects	Drug dilution by tears	51,217
	Increased stability of drugs	Short contact time	217,219
	Reduced dosing intervals	Delivery to posterior sections requires crossing of numerous physiological barriers	217,220
	Improved mucosal retention	Avoiding systemic side-effects	217,221
		Cornea is a static barrier to drug diffusion	51
		Temporary vision impairment	91
		Rapid mucociliary clearance	222,225
Nasal	Viable option for systemic drug delivery	Potential enzymatic degradation	223
	Direct nose-to-brain delivery by bypassing the blood–brain barrier	Precisely targeting the drug to specific regions of the nasal cavity	223
	Localized drug delivery	Potential for irritation or damage to the nasal mucosa with repeated use	222,226
	Bypassing hepatic metabolism	Low drug retention time	223
	High epithelial permeability	Often involves a drug/device combination	222,225
	Swift onset of action		223,225
	Simplified administration		223,227
	Relatively low enzymatic activity		224
	Non-invasive vaccine administration site		223
Oromucosal	Direct delivery to the systemic circulation	Limited surface area	228,229,234
	Swift onset of action	Salivary washout	228,229,235
	Relatively low enzymatic activity	Potential for enzymatic degradation	228,229,234
	Suitable for patients with swallowing difficulties	Short residence time (swallowing)	230,236
	Excellent route for emergency treatments	Establishing sustained release	231,237
	Vaccination target site	Tongue movement	232,237
	Epithelial permeability	Eating and drinking can interfere with drug absorption	233,235
		Permeability varies according to stage of menstrual cycle	13,238
Vaginal	Large surface area for drug absorption	pH influenced by stage of menstrual cycle	13,239
	Bypassing liver metabolism	Intercourse alters pH	77,238
	Reduced occurrence of side-effects	Menstruation washes drug away	12,238
	Dense network of blood vessels	Limited drug retention time	238,239
	Access to the uterine first-pass effect	Leakage of formulations	12,238
	Localized treatment during pregnancy		

To capitalize on the advantages listed in Table 1, combined liquid-based drug delivery approaches advance the capacity of established formulations to optimize their targeting potential and subsequent therapeutic efficacy. Examples include simple strategies such as mixing ionic liquids to impart improved physical characteristics to tailor conductivity, viscosity, and polarity as a means to improve drug solubility and mediate the crossing of mucosal barriers^{73,240}. Advanced approaches are also reported such as the work of Lin and co-workers exploring the incorporation of oil-in-ionic liquid-based nanoemulsions intended for intranasal influenza split-virus vaccine delivery²¹⁵. Their study involved mixing squalene (oil phase), Tween[®] 80 (surfactant), choline, and niacin (ionic liquid combination replacing traditional water phase) in a single-step ultrasonic method. The ionic liquid combination drastically improved emulsion stability compared to a conventional O/W counterpart emulsion. Moreover, their results revealed a 25-fold higher secretory IgA titers *via* adjuvant antigen induction compared to antigens that were not delivered *via* a combined liquid-based drug delivery approach. Additionally, the oil-in-ionic liquid-based nanoemulsion rendered a 5.8-fold superior antigen induction, systemic humoral, mucosal, and cellular immune response, compared to the commercially available MF59-adjuvant vaccination²¹⁵. These favorable results were attributed to prolonged nasal mucosa retention mediated by the combined liquid-based drug delivery vehicle. This led to superior paracellular transport, expertly delivering antigens to the underlying submucosa. This success can also be attributed to the uniform nano-sized droplets mediating mucosal crossing of the incorporated antigens²¹⁵. When considering the mucoadhesion theories, prolonged mucosal retention at the nasal mucosa could have been mediated by ionic liquid inclusion adhering to the mucus layer *via* electrostatic interactions to prolong mucosal retention time^{73,241,242}. Moreover, ionic liquids can also improve wetting of the mucosal surface as promoted by the improved spreadability of the ionic liquid-nanoemulsion combination to optimize mucoadhesion^{243,244}. The uniform, small droplet size of the nanoemulsion component can also promote diffusion through the mesh-resembling mucin network of nasal mucus²⁴⁵. Thus, demonstrating the genius of combining liquid-based drug delivery approaches to capitalize on multiple theories of mucoadhesion to render superior transmucosal drug delivery.

Combining liquid-based drug delivery systems can inspire innovative approaches that mimic viruses, which have naturally evolved to cross mucosal barriers through mechanisms of mucoadhesion and mucosal penetration^{87,246,247}. By integrating these liquid-based formulations, it can be possible to create an effective platform that utilizes both mucoadhesion and mucosal penetration. However, before moving toward combined strategies, it is essential to have a comprehensive understanding of each liquid-based drug delivery system. Key considerations of the individual liquid-based drug delivery systems presented by this review are summarized in Table 2^{21,73,75,138,142-144,152,170,171,174,175,181,196,207,248-286} to understand the targeting mechanism of these dosage forms when used individually or in combination to mediate the crossing of mucosal barriers guarding the ocular-, nasal-, oromucosal-, and vaginal epithelia.

4. Liquid-based drug delivery approaches for ophthalmic administration

The eyes can be described as specialized organs permitting sensory awareness *via* impulse transmission processed into vision

within the brain²⁸⁷. The eyes present a highly sophisticated physiology which can be divided into posterior and anterior segments²⁸⁸. The anterior segment occupies approximately one-third of the eye and includes the following ophthalmic structures: conjunctiva, cornea, aqueous humor, ciliary body, iris, and crystalline lens. The posterior segment refers to the remaining ocular structures (*i.e.*, sclera, Bruch's membrane, choroid, retinal pigment epithelium, vitreous humor and neutral retina)²⁸⁹. Pharmaceutical scientists are still challenged by ocular drug delivery due to various barriers provided by the complex ocular structures such as static barriers referring to the corneal stroma, stratified corneal epithelium, and sclera. Moreover, dynamic barriers like lymphatic clearance, conjunctival blood flow, and tear turnover rate combined with metabolic barriers such as enzymatic action and efflux pumps can impede ophthalmic drug delivery²⁹⁰.

The main challenge for ophthalmic delivery systems is to reach an optimum drug concentration at the corneal surface to establish localized therapeutic efficacy²⁹¹. Almost 95% of the ocular products on the market are developed for the topical route of administration^{291,292}. The bioavailability of this route is as low as 5%, caused by various factors like tear/solution drainage, blinking, lacrimation, and tear dilution^{291,292}. The cornea is the primary ocular transmucosal route for topical drug absorption²⁹³. The literature describes the cornea as a collagenous structure providing mechanical strength to the eyeball. The cornea presents a transparent nature to fulfil its light-focusing function on the retina²⁹⁰. The multilayered nature of the cornea is enforced by the presence of lipophilic and hydrophilic layers, which makes it relatively impermeable²⁹³. After crossing the tear film, the cornea presents five layers²⁹⁴. These layers are predominantly lipophilic, implying that the absorption of hydrophilic drugs can be difficult²⁹³. At a physiological pH the cornea presents acidic groups on the ocular surface and therefore negatively charged drugs may exhibit a slower absorption rate²⁹³. The barriers to ocular drug permeation are presented in Fig. 8.

Understandably, more than 95% of ocular products on the market are eye drops, with advantages like easy administration and efficient stability²⁹⁵. The majority of eye drops are aqueous based, with common negatives including rapid clearance combined with the failure to be retained at the corneal mucosa¹⁷⁶. Simplicity of the products, ease of administration, convenience, good corneal penetration, patient compliance, lower toxicity and reduced side effects, are some of the advantages of ophthalmic drug administration^{291,293}. Simplified administration is a necessity as geriatric patients are at higher risk of suffering from ocular conditions implying that formulators should avoid complex drug administration strategies to ensure patient adherence. Examples of ocular conditions include cataract formation, myopia, age-related macular degeneration, scarring, chronic dry eyes, diabetic neuropathy, post-operative endophthalmitis, proliferative vitreoretinopathy, inflammation, allergies, glaucoma, and fungal/bacterial keratitis²⁹⁶.

Developing new drug delivery systems other than conventional ocular therapy to improve efficacy and bioavailability can present challenges²⁹¹. Importantly, ideal ocular drug delivery vehicles should demonstrate good corneal penetration, prolong drug retention within the ocular mucosa, target specific areas of the eye to mediate a localized therapeutic effect, render sufficient bioavailability, and avoid vision impairment^{296,297}. Most of the recent ophthalmic research attempts to increase the residence time of drug delivery systems²⁹⁸. The majority of research work is still in the experimental phases, and not commercially available yet. In

Table 2 Key considerations of transmucosal liquid-based drug delivery systems.

Drug delivery system	Key excipients	Targeting mechanisms	Outcomes	Ref
Nanoemulsions	Oil phase	Mucoadhesion	Increased bioactive stability of drugs	248,249
	Water phase	Reduced droplet size	Reported inability to cross corneal barrier	248,250,252
	Surfactant/co-surfactant	Site-specific drug release	Superior brain targeting following intranasal administration	251
	Polymers			248,252
SEDDSs	Oil phase	Mucoadhesion	Prolonged drug stability	253
	Water phase	Muco-penetration	Enhanced cellular uptake	144,143
	Surfactant/co-surfactant	Enzymatic encapsulation	Enhanced drug solubility	75,144,254,256
		Enhanced drug solubility	Superior spreadability	144,255
		Rapid emulsification in small fluid volumes	Increased permeation	138,254
				142
Double-emulsions	Oil phase	Encapsulating hydrophilic and lipophilic drugs	Improved sustained release	152,254
	Water phase	Enhanced drug solubility	Enhanced corneal permeability	21
	Surfactant			21,258
	Co-surfactant			21,257
Sol-gel transition dosage forms	Thermo-responsive polymers	Increased spreadability	Superior reduction of intraocular pressure	21,258
	Water	Mucoadhesion	Enhanced corneal permeability	21,257
		Prolonged drug retention	Enhanced nasal mucosa permeation	21,258
		Reduced leakage	Increased nasal retention time	171
		Stimuli dependent site-specific drug release	Sustained release (intranasal)	170,260,261
			Enhanced nasal mucosa permeation	170,259
			Increased nasal retention time	170,171,262
			Sustained oromucosal release of multiple drugs	170,260,262
Fluidgels	Biopolymers	Mucoadhesion	Superior reduction of intraocular pressure	260
	Water	Increased spreadability	Increased mucosal wound healing due to shielding	263
		Increased surface retention	Superior vaginal mucosa coverage	264
			Enhanced sprayability capacity (vaginal administration)	174,175,265
Ionic liquids	Can fulfill role of: oil phase	Enhanced drug solubility	Effective nose-to-brain delivery	175,265
	water phase	Mucoadhesion	Enhanced sprayability capacity (vaginal administration)	265
	Structural component	Shielding mucin glycoproteins	Good biocompatibility with oral mucosa	266
		Altering viscosity and elasticity of mucus	Increased dissolution rate in synthetic saliva	181,267,268
		Prolongs contact time	Increased dissolution rate in synthetic saliva	181,267,268
			Superior mucoadhesion in saliva	73,181,269
Liquid crystals	Surfactants	Superior drug loading	Superior mucoadhesion in saliva	73
	Polymers	Optimized drug release		73,270
	Co-solvents	Mucoadhesion		271
	Amphiphilic lipids		Prolonged drug bioavailability (corneal application)	196,272
			Superior nose-to-brain and systemic targeting (lipid molecules) following intranasal administration	196,273
			Prolonged oromucosal retention	274,276,277,278
			Enhanced oromucosal permeability	275
			Sustained release/prolonged retention (vaginal mucosa)	279
Eutectic systems	Fatty acids	Enhanced aqueous solubility	Controlled release (vaginal mucosa)	280,281
	Alcohols	Permeation enhancement		280,281
	Ionic liquids			282

this review, we will consider examples of experimental work to demonstrate the advances of liquid-based ophthalmic drug delivery systems.

4.1. Nanoemulsions

A recent study by Tran and co-workers incorporated cannabidiol into a nanoemulsion for ocular administration to relieve intraocular pressure and inflammation²⁵². Cannabidiol is notorious for its chemical instability, extensive liver metabolism, and high lipophilicity impeding its clinical use. Hence, localized drug delivery in a suitable vehicle can potentially bypass first-pass metabolism and improve the stability of cannabidiol. The potential of cannabidiol micellar formulations described as nanoemulsions was

tested on three human corneal cell lines used to construct an artificial human corneal substitute. Their findings concluded that nanoemulsions drastically improved the bioactive stability of cannabidiol. However, the produced nanoemulsions were not able to cross the corneal barrier²⁵².

4.2. Double-emulsions

The objective of another study involved the development of a cationic reverse micelle-based double-emulsion to improve the therapeutic activity of betaxolol hydrochloride in anti-glaucoma therapy²⁵⁸. The hydrophilic betaxolol hydrochloride was incorporated inside the core of reverse micelles in a W/O/W double-emulsion to generate sustained drug release while improving

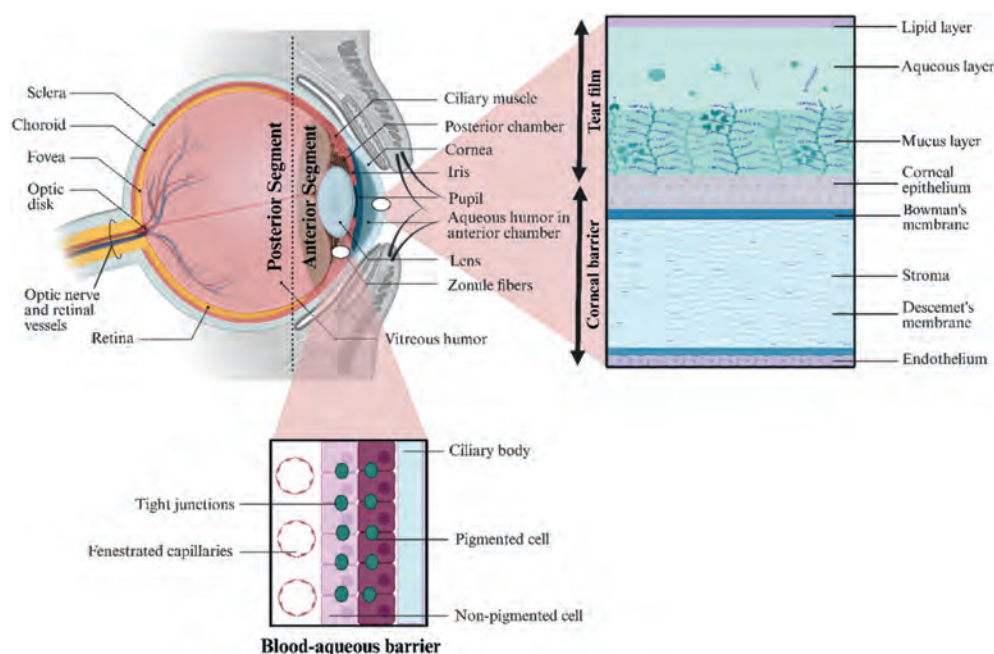


Figure 8 Barriers to ocular drug delivery²²¹. Created with BioRender.

corneal permeability. A two-step emulsification process was used to produce the primary emulsion by homogenization followed by sonication to assemble reverse micelle formation. For reference, the method and excipients used to produce the aqueous and oil phases are thoroughly described by Sakr and co-workers²⁵⁸. The optimized W/O/W double-emulsion provided a sustained release for up to 7 h following the initial rapid release of betaxolol hydrochloride. Moreover, an *ex vivo* permeation evaluation revealed that the W/O/W double-emulsion rendered 2.5-fold enhanced permeation upon comparison to a marketed eye drop²⁵².

4.3. Self-emulsifying drug delivery systems

Generally, SEDDSs are employed to improve the aqueous solubility of lipophilic drugs²⁹⁹. Interestingly, a recent study proposed utilizing self-emulsifying oils for water-soluble drugs that do not remain stable in water for long periods of time such as the antibiotic cefuroxime sodium²⁵⁶. Cefuroxime sodium was suspended in self-emulsifying oils to guard against external water exposure during storage. Self-emulsifying oil phases comprised 5% w/w of either Cremophor® EL, Span® 80 and Tween® 20 combined with Miglyol oil. Benzalkonium chloride at a concentration of 0.01% w/w was also considered as an adjuvant cationic surfactant. Results of 6-month stability testing at a temperature of 40 °C revealed less than 2% chemical degradation of cefuroxime sodium in Span® 80 and Tween® 20 formulations. Moreover, the effect of benzalkonium chloride on chemical degradation was described as negligible. The suspended cefuroxime sodium transpired into spontaneous emulsions upon exposure to small fluid volumes. Thereby, signifying its potential as a substitute for conventional antibiotic eye drops²⁵⁶.

4.4. Liquid crystals

A recent study suggested co-delivery of dorzolamide and timolol facilitated by liquid crystal nano cubosomes to mediate trans-corneal drug delivery for glaucoma treatment²⁷⁸. At this point in

time, the nanoscale co-delivery of dorzolamide and timolol is a novel scientific contribution, as the literature only reports individual nanoencapsulation of these drugs. Glycerol monooleate and poloxamer 407, together with the two drugs, were stirred, homogenized, and probed to produce liquid crystals. The cubosomes presented an average size of 143.5 ± 2.3 nm, zeta potential of 26.3 ± 1.7 mV, and PDI of 0.164 ± 0.014 . Dorzolamide and timolol were encapsulated at a concentration of $84.5 \pm 2.1\%$ and $91.3 \pm 2.7\%$, respectively. Importantly, *in vitro* evaluation revealed gradual release for both drugs providing a maximum dose 24 h after administration. Moreover, *in vivo* studies performed on New Zealand rabbits revealed reduced intraocular pressure 4 h after liquid crystal administration. Pharmacokinetic experiments depicted prolonged bioavailability compared to a commercial product. Additionally, histopathological studies identified the nano cubosome liquid crystals as optimally biocompatible with corneal epithelia²⁷⁸.

4.5. Phase transforming liquid formulations

In terms of stimuli-responsive drug delivery systems, temperature is most often used as a trigger for phase transition regarding ocular drug delivery³⁰⁰. For instance, a study attempted to minimize intraocular pressure by developing thermo-responsive gels incorporating the prodrug dipivefrin hydrochloride containing poloxamers (P407 and P188)-carbopol-934. These polymer-based gels provided simplified application due to their solution-like nature. External stimuli can transform these solutions to form a gel, which prolongs the ocular residence time due to improved viscosity and mucoadhesive properties^{301,302}. External stimuli included temperature, pH and the presence of ions³⁰¹. A thermo-responsive gel transforms through the sol-gel phases after temperature changes²⁶¹. This system shows a superior ability to reduce elevated intraocular pressure in rabbit eyes²⁶¹.

Ion-activated polymers like gellan gum and alginic acid are also used in ophthalmology formulations³⁰³. To prolong ocular

residence time, alginic acid, a biodegradable and biocompatible polysaccharide, is successfully used^{304,305}. This polysaccharide is useful since it increases the ocular drug resistance time due to its mucoadhesive and gel formation properties³⁰⁴. This can be attributed to the double helices formation *via* weak van der Waals forces produced by gellan gum at room temperature. The lacrimal fluid consists of cationic electrolytes, and upon contact between gelling agents and lacrimal fluid, these helices aggregate leading to crosslinking of the polymer. This resulted in formed complexes with the cations and hydrogen bonds of the water to improve ocular residence time^{306–308}.

4.6. Eutectic systems

A recent review article written by Sarmiento and co-workers raises the question of whether deep eutectic systems obtained from a natural origin can enhance the efficacy of ocular therapeutics³⁰⁹. This excellent review considers the different layers of the cornea that must be permeated during transmucosal drug delivery and how deep natural eutectic systems can modify viscosity, be a bioactive excipient, improve aqueous solubility, and provide superior stabilization of drug delivery systems to enhance ocular drug delivery³⁰⁹. As an example, this study referred to a study performed by Achkar and co-workers that developed novel supramolecular deep eutectic systems *via* levulinic acid and cyclodextrin inclusion³¹⁰. These unique systems provide multiple drug encapsulation due to the presence of cyclodextrin while improving the permeation of mucosal barriers³¹⁰. Thereby, signifying that natural eutectic systems might be the next hot topic in ocular drug delivery.

5. Liquid-based drug delivery approaches for nasal administration

The main function of the nose is to filter out airborne particles, to heat and humidify inhaled air and to provide a mechanism for olfaction. Although the well-perfused mucosal epithelial surfaces of the nasal cavity are designed to protect the body against the uptake of unwanted foreign materials, they also provide an opportunity for drug absorption after nasal administration³¹¹.

Evidence exists that people have administered traditional medicines into the nose since ancient times, but the use of nasal sprays for modern medicines commenced approximately in the middle of the 20th century. These original nasal spray products were mainly intended for local action in the nose (*e.g.* for the relief of nasal congestion experienced during diseases such as allergic rhinitis). However, nasal drug delivery products have since evolved tremendously and the nasal route of drug administration has become a useful alternative for the administration of drugs for systemic delivery^{312,313}. Furthermore, it was discovered that the nasal route of drug administration not only provides a viable option for systemic drug delivery, but it also provides an opportunity for delivery of drug molecules into the brain³¹⁴.

Direct nose-to-brain drug delivery after nasal administration provides an effective, advantageous and non-invasive way to treat neurological and mental diseases of the central nervous system due to avoidance of the blood–brain barrier. By deposition of the drug dose in the olfactory region of the nasal cavity during administration, the nasal route of drug administration can specifically be used to target drug delivery to the brain³¹⁵. Delivery of drug molecules into the brain after nasal administration can occur through different pathways of absorption (as presented in Fig. 9).

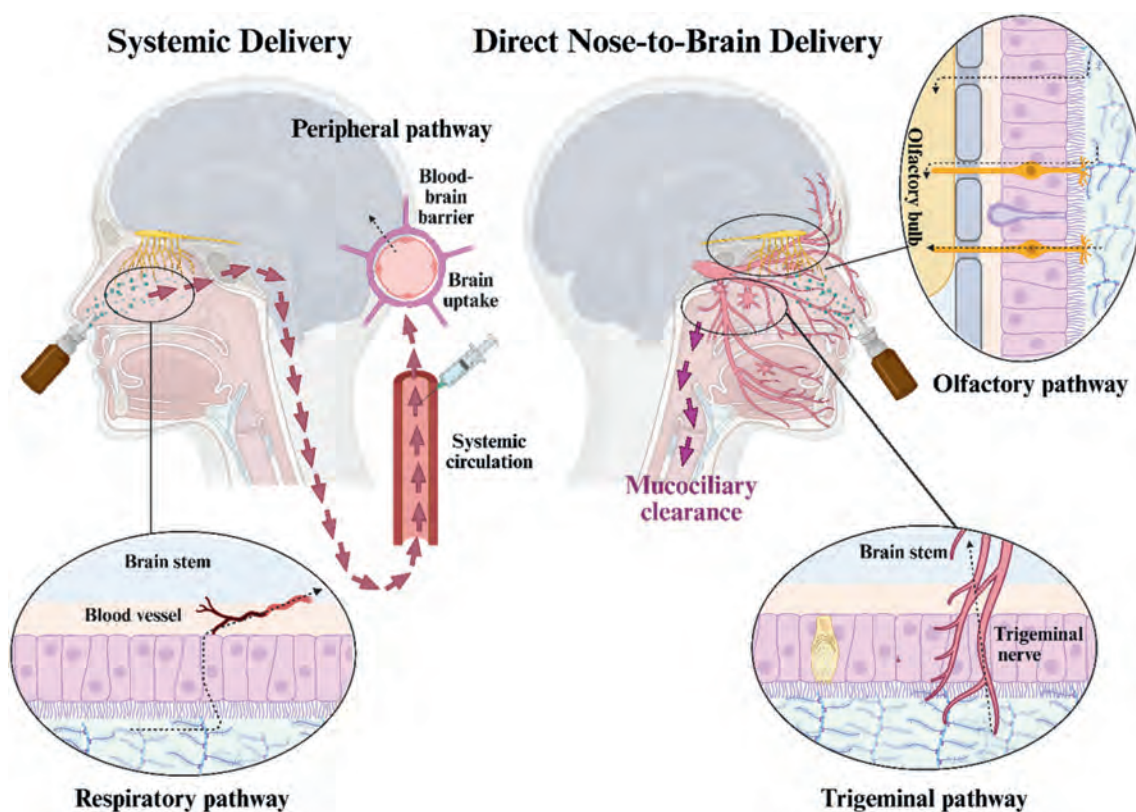


Figure 9 Schematic representation of pathways for intranasal drug delivery^{316,317}. Created with BioRender.

Namely by diffusion across the olfactory epithelium or through uptake *via* the olfactory neurons (olfactory pathway) and/or through uptake *via* the trigeminal nerve (trigeminal pathway). In addition, drug molecules can be indirectly delivered into the brain after nasal administration (respiratory pathway) *via* the systemic circulation, but then the drug molecules need to cross the blood–brain barrier (peripheral pathway)²²³.

The nasal route of drug administration is specifically suitable as an alternative to the oral route of drug administration for delivery of drugs that are active in relatively small doses and that are highly susceptible to first-pass metabolism³¹⁸. The nasal route of drug administration is also useful for targeted delivery of drugs for the treatment of central nervous system diseases, where drug levels in the brain are usually hampered by the blood–brain barrier when these drugs are administered by the oral and parenteral routes of drug administration³¹⁵. Unfortunately, the nasal route of drug administration is less suitable for treatment of chronic conditions that require multiple daily doses over an extended period of time and also conditions that are treated with drugs that require relatively large doses due to the limited volume and surface area available in the nasal cavity^{319,320}.

The application of the nasal route of drug administration will be demonstrated here by giving examples of drug products currently available on the market for treatment of various conditions. Firstly, nasal medicinal products are available for local action in the nose to treat diseases affecting the nasal cavity such as allergic and non-allergic rhinitis (*e.g.*, antihistamines and steroidal anti-inflammatory agents), nasal congestion (*e.g.*, vasoconstrictors), intranasal anesthesia (*e.g.*, local anesthetics) and nasal polyps (*e.g.*, corticosteroids). A wide variety of nasal drug products are available for nasal administration to afford a systemic action to treat diseases or conditions such as general pain (*e.g.*, opioid analgesics), diabetes insipidus (*e.g.*, hormones such as desmopressin), post-menopausal osteoporosis (*e.g.*, hormones such as calcitonin) and for smoking cessation (*e.g.*, autonomic ganglia stimulants). Nasal products are available for the treatment of neurological and mental diseases such as migraine (*e.g.*, ergot alkaloids and selective serotonin agonists), epilepsy related seizures (*e.g.*, benzodiazepines) and resistant major depression (*e.g.*, *N*-methyl-D-aspartate receptor antagonists)^{312,321,322}. Lastly, the nasal route of drug administration is also highly suitable for the administration of emergency or rescue therapeutics due to its rapid onset of pharmacological action and ease of administration. In this regard, nasal products are currently commercially available for the treatment of opioid overdose (*e.g.*, opiate antagonist such as naloxone) and treatment of seizures during epilepsy attack (*e.g.*, benzodiazepines such as midazolam)^{312,321}.

5.1. Ionic liquids

Ionic liquids can be formed through ionic interactions between anions and cations, which have been explored as functional excipients in formulations to improve drug delivery *via* different routes of administration. Choline-based ionic liquids have initially been explored as permeation enhancers for the transdermal delivery of both macromolecules and small molecules with poor water solubility^{323,324}, but later also for enhanced drug delivery *via* the nasal route of drug administration. The ability of ionic liquids to enhance the adhesion and residence time of molecules on nasal mucosal surfaces has specifically stimulated research into using this formulation technique for improving drug delivery into the systemic circulation or into the brain after nasal administration³²⁵.

Choline-based ionic liquid formulations have been investigated using both *in vitro* and *in vivo* models for the enhancement of nasal glucagon delivery. In this study, choline geranic acid and choline citric acid have been used to form ionic liquids for enhancing the permeation and prolonging the retention time of glucagon after intranasal administration. Intranasal delivery of glucagon in the form of ionic liquids elicited a rapid reversal of acute hypoglycemia in type 1 diabetic rats without leading to rebound hyperglycemia. In addition, electrostatic interaction was identified as the mechanism by which ionic liquids improve the transcellular transport of glucagon across the Calu-3 cell monolayer model, which serves as an *in vitro* model for nasal epithelial surfaces. The ionic liquids could also transiently open intercellular tight junctions to facilitate the paracellular transport of glucagon. Although continuous intranasal administration of ionic liquids led to changes such as inflammation in the epithelial cells, goblet cell overgrowth and distribution changes of the cilia, they were reversible and the nasal epithelium was able to repair itself³²⁵.

A study that investigated the effect of an ionic liquid formulation on the nose-to-brain delivery of etodolac in mice after nasal intranasal administration showed high potential of these formulations for improved drug delivery to the central nervous system. The results exhibited an approximate 7-fold increase in etodolac delivery into the brain when compared to that of etodolac solution alone. Furthermore, the solubility of etodolac was improved approximately 200-fold by ionic liquid formation in simulated nasal fluids. The increase in the nose-to-brain delivery of etodolac was also partially attributed to the enhanced retention of the compound on the nasal mucosal surface due to the higher viscosity of the ionic liquid as compared to that of the solution²⁶⁸.

5.2. Liquid crystals

Liquid crystal formulations have been investigated as a novel approach for enhanced systemic and brain delivery of drugs after nasal administration. One example of such a study is where liquid crystal formulations were prepared with monoglycerol ester and glyceryl monooleate for tranilast delivery into the systemic circulation as well as into the brain after intranasal administration in Sprague Dawley rats. The monoglycerol ester liquid crystal formulation exhibited a 10-fold increase in tranilast exposure in the whole brain ($AUC_{0-8} = 94.5 \pm 3.68$ ng h/mL) as compared to the solution formulation (control group, $AUC_{0-8} = 9.55 \pm 2.17$ ng h/mL). Furthermore, when ethanol and a non-ionic surfactant (Pluronic® F-127) were included in the liquid crystal formulation, the tranilast exposure in the whole brain ($AUC_{0-8} = 114 \pm 5.92$ ng h/mL) increased by 12-fold. Furthermore, the monoglycerol ester liquid crystal formulation containing ethanol and Pluronic® F-127 caused a 3.8-fold increase in tranilast blood plasma level ($AUC_{0-8} = 725 \pm 33.1$ ng h/mL) as compared to that of the solution formulation ($AUC_{0-8} = 189 \pm 16.2$ ng h/mL). The results from this study therefore showed that liquid crystal formulations prepared from monoglycerol ester have particular high potential for enhanced delivery of lipid drug molecules into the brain and systemic circulation after nasal administration²⁷⁹.

5.3. Nanoemulsions

Different nanocarrier type dosage forms have been investigated to improve drug delivery to the brain after nasal administration, which include lipid-based nanoparticles, polymeric-based

nanoparticles, polymeric micelles, nanosuspensions, nanovesicular spanlastics, dendrimer-based nanoparticles, nanosponges, neosomes and nanoemulsions³²⁶. The use of nanoemulsions to target brain delivery after nasal administration has been reviewed elsewhere^{253,327}. In general, it was found in several studies that nanoemulsions provide a non-toxic formulation strategy to improve nose-to-brain delivery of neurotherapeutic drugs. To demonstrate the efficacy of nano-emulsions in enhanced nose-to-brain drug delivery, some examples of nanoemulsion formulations used for investigations on drug delivery into the brain and efficacy after nasal administration will be described here.

Mucoadhesive nano-based drug delivery systems can be prepared by using chitosan as a coating material, which has proven effective in enhancing the delivery of therapeutic agents to the central nervous system after nasal administration. The *ex vivo* permeation and *in vivo* brain delivery of luteolin in Wistar rats when formulated in chitosan-coated nano-emulsion showed 6-fold and 4.4-fold increases, respectively, when compared to that of free luteolin. The improved delivery of luteolin when formulated in the nano-emulsion was attributed to the mucoadhesive properties of the chitosan-coated nanoemulsion formulation, which could improve the time of contact of the luteolin to the nasal epithelium, and the very small diameter of the nanoemulsion droplets¹³⁴. In another study, a chitosan-coated nano-emulsion was developed for the chemotherapeutic agent, temozolomide, to improve its nose-to-brain delivery for treatment of glioblastoma multiforme. This chitosan-coated nanoemulsion significantly enhanced the *ex vivo* mucosal permeation of temozolomide when compared to the free drug and the nanoemulsion drug delivery system has also exhibited enhanced *in vitro* uptake of temozolomide into brain cells³²⁸. Sildenafil formulated in a chitosan-coated nanoemulsion showed increased efficacy (as determined by oxidative stress markers) in a hyperlipidemic rat model as compared to a sildenafil suspension formulation³²⁹.

When docosahexaenoic acid concentrations are decreased in the brain, it increases the risk for development of Alzheimer's disease. It has been suggested that supplementation of the brain with docosahexaenoic acid may contribute to prevention of Alzheimer's disease. A nanoemulsion formulation containing docosahexaenoic acid did not show any detrimental signs in the viability of RPMI 2650 cells as measured with MTS assays. Intranasal administration of a nanoemulsion of docosahexaenoic acid in Hemizygous hAPPSwInd transgenic (J20) mice had positive effects on the cognitive behavior of the mice through counteracting oxidative stress, neuroinflammation and amyloid plaque formation³³⁰.

5.4. Phase transforming liquid formulations

A thermosensitive gel can be defined as a specialized type of hydrogel that transforms from a solution to a semi-solid gel in response to environmental stimuli such as temperature, pH or ionic strength. In this regard, the thermosensitive gel exists in the free-flowing liquid state at ambient temperature, but turns into a highly viscous gel at physiological temperatures (*e.g.*, 32–37 °C) by means of sol-gel transition. Although liquid-based dosage forms such as metered dose nasal sprays are the most commonly used formulations for intranasal drug administration, they exhibit short retention times in the nose due to removal by mucociliary clearance. The use of thermosensitive gels showed special application for increased nose-to-brain drug delivery after intranasal

administration due to increased residence time in the nose as compared to that of solutions³³¹. Several studies have investigated novel strategies to further improve the physical properties of thermosensitive gels for more efficient drug delivery after nasal administration. These strategies included the incorporation of functional excipients or nanocarriers into the thermosensitive gel formulations. One such study investigated the incorporation of graphene-oxide in a thermosensitive chitosan hydrogel for intranasal delivery of rasagiline mesylate. In this work, the incorporation of graphene-oxide showed a significant concentration-dependent effect on the physical and mechanical properties of the thermosensitive chitosan hydrogel such as a decrease in the gelation time, porosity, and swelling of the hydrogel, but an increase in gel strength³³². In another study, the mixing of different poloxamer polymers in several ratios as well as incorporation of hydroxypropylmethylcellulose showed improved mucoadhesion properties. The optimized thermosensitive hydrogel showed high potential for carbamazepine delivery to the brain after nasal administration²⁶⁰. Moreover, nifedipine was formulated into a nanostructured lipid carrier-based intranasal *in situ* gelling system, which showed optimal gelation time, temperature, and pseudothixotropic behavior, sustained release and enhanced permeation across the nasal mucosa. Furthermore, *in vivo* administration of the thermosensitive gel containing nifedipine in lipid nanocarriers revealed substantial improvement in cognitive ability along with reduced oxidative damage in an Alzheimer's disease Wistar rat model²⁶².

6. Liquid-based drug delivery approaches for oromucosal administration

The oromucosal, or pre-gastric, route of drug administration consists of the buccal (between the gums and cheeks) and sublingual (under the tongue) routes and has shown great promise in recent decades as an alternative route of drug administration²²⁸. Several physiological factors make the mucosal membranes of this route highly permeable, such as its low degrees of epithelial thickness and keratinization.

Concretely, the thinnest regions of the oral cavity can be found in the floor of the mouth, inner sides of the cheeks and lips, and ventral side of the tongue, *i.e.*, the very regions which constitute the buccal and sublingual routes of administration. These oromucosal regions are indicated in Fig. 10. A major reason being that these regions consist of non-keratinized stratified squamous epithelium^{228,333}. The pH of the buccal mucosa is also more neutral, with a reported average value of 6.28 ± 0.36 ⁷⁰, making it more amicable to drugs that might degrade or denature in lower pH environments. Moreover, the intercellular spaces of the epithelium in the upper third of the inner sides of the cheeks are tight junction-free. Instead, these spaces are filled with lipids, facilitating the rapid permeation of small lipophilic molecules^{334,335}.

Apart from the physiological advantages of the oromucosal route, there are also several patient or treatment related problems that can be addressed by administering drugs *via* this route. These include sub-therapeutic drug concentrations, and irregular or inappropriate drug use, potentially leading to treatment discontinuation^{338–341}. The above-mentioned negative therapeutic outcomes are more prevalent geriatric and pediatric patients, as well as dysphagic patients, highlighting the growing need for patient-centered pharmaceutical development^{231,340,342}. The oromucosal route, with its high permeability, rapid onset of action,

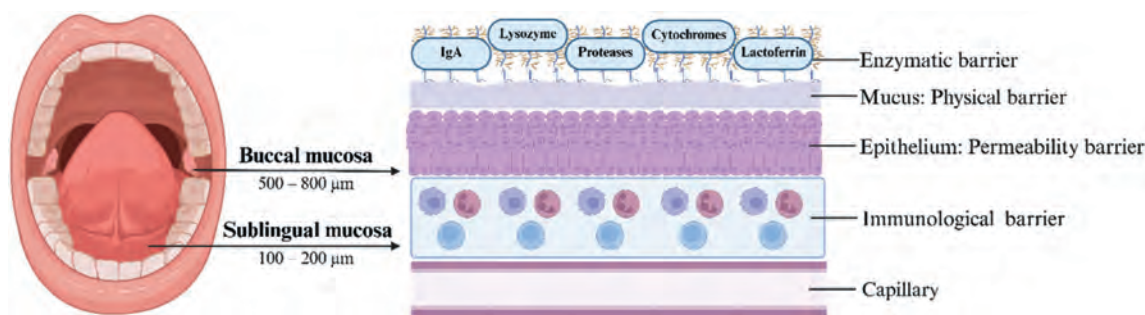


Figure 10 Mucosal barriers to oromucosal drug delivery^{247,336,337}. Created with BioRender.

and no need to swallow (for patients with dysphagia), might be an appealing route of drug administration for future pharmaceutical development.

An excellent review by Bastos and co-workers²³¹, explored the oromucosal products currently on the market, highlighting the broad range of conditions that are currently treated using this route of administration. Examples of these conditions include allergic rhinitis and asthma, chronic pain, opioid dependence, angina, recurrent herpes labialis, and allergic conjunctivitis. The dosage forms of these products mostly include sublingual tablets, wafers, powders, drops, and sprays, as well as buccal films and mucoadhesive buccal tablets²³¹. In this paper, we will explore recent examples of liquid-based formulations for oromucosal drug delivery.

6.1. Ionic liquids

Several studies have suggested that ionic liquids may offer an exciting means of delivering drugs *via* the oromucosal route. Vaidya and Mitragotri (2020) developed mucoadhesive buccal patches of insulin-containing choline and geranic acid (CAGE)-polyvinylalcohol-based ionic liquids and assessed the systemic delivery thereof in rats²⁶⁹. Not only were the insulin/CAGE ionic liquids able to give a dose-dependent reduction in blood glucose levels of up to 50%, but the patches also did not cause significant damage to the epithelial tissue, suggesting good biocompatibility^{73,269}.

Ng and Hadinoto (2024) developed gelatin films loaded with imidazolium-based ibuprofen salt ionic liquids, and assessed the effects of ionic liquid loading on the films' mechanical strength, inter-batch dosage uniformity, thermal stability, and dissolution and solubility enhancement²⁷⁰. Their results showed good inter-batch drug content uniformity, with an acceptance value (AV) of 15%. The dissolution rate of ibuprofen in simulated saliva fluid was also sufficiently enhanced, to 100% in 15 min²⁷⁰.

Although not a drug delivery study, Medeiros's group²⁷¹ developed mucoadhesive films containing chitosan and protic ionic liquids, to act as wound dressings on the oral mucosa. Their results suggested that films based on chitosan and containing 35% to 50% (w/w) 2-hydroxy diethylammonium protic ionic liquids had superior mucoadhesive properties, antimicrobial activity, and flexibility in synthetic saliva.

6.2. Self-emulsifying drug delivery systems

Another formulation strategy that has found novel application in the oromucosal route of drug administration, is that of SEDDS. Since SEDDS are known to improve the cellular uptake of entrapped drugs^{76,343}, it might be an appealing approach to

enhance the permeation of drugs with limited mucosal tissue penetration.

Friedl's group¹⁴³, investigated SEDDS as a possible enhancer of the buccal delivery of curcumin, a biopharmaceutics classification system (BCS) class IV drug. Their delivery system was a mucoadhesive patch, consisting of two intersecting fiber types. The first type of fiber consisted of thiolated polyacrylic acid, while the second type consisted of a water-insoluble polymer containing curcumin-loaded SEDDS. As saliva passes through the patch, the second fiber type releases the SEDDS as nanoemulsions. As a control, the same mucoadhesive patch was developed, but containing only curcumin (not SEDDS) in the second fiber type. The patches were then applied to excised porcine buccal tissue and studied. The results suggested that the patches containing SEDDS were able to dramatically enhance the penetration of curcumin into buccal tissue, relative to the SEDDS-free patches. These findings suggest that SEDDS formulations can be a viable drug delivery approach for the oromucosal administration route.

6.3. Liquid crystals

The use of a methylene blue-loaded mucoadhesive liquid crystal precursor system (LCPS), intended for the treatment of oral cancers *via* the buccal route of administration, was investigated by Balian and co-workers³⁴⁴. Their rationale for using LCPSs was based on the knowledge that these systems can give sustained drug release at the site of administration^{345,346}, and turn into gels upon contact with saliva, increasing the residence time in the oral cavity^{280,281}. Furthermore, their LCPS contained polyethyleneimines and chitosan as mucoadhesive polymers, further adding to the residence time of the LCPS in the oral cavity, thereby possibly enhancing the efficacy of the treatment^{276,277}. Indeed, their results showed that methylene blue in their LCPS was retained twice as long as its permeability, suggesting that adequate maintenance of therapy in the oral cavity could be achieved. The methylene blue-loaded LCPS also showed an increase in antitumor activity, compared to the control³⁴⁴.

6.4. Phase transforming liquid formulations

Bender's group²⁶³ developed a phase-transitioning sol-gel system based on Carbopol 934P NF and Pluronic F-127 blends, containing lidocaine and metronidazole. The rationale behind their formulation was to have the sol-phase make contact with the inner surfaces of the empty tooth socket, followed by a transition to the gel-phase. The resulting monolithic gel-phase then remains in the socket, preventing the ingress of debris and providing sustained release of lidocaine and metronidazole at sub-toxic levels, thereby

providing pain relief and protection from bacterial growth. Their results showed that at 23.38% Pluronic F-127, the gelation of their sol-gel started at 23 °C, and was complete at 37 °C. The gel-phase was able to give sustained release of both drugs over 24 h, at therapeutically relevant concentrations. Although not strictly oromucosal delivery, the findings presented by Bender and co-workers suggested that phase-transforming sol-gels can be used to treat wounds in the oral cavity, and can therefore possibly find future uses as oromucosal administration devices²⁶³.

Phase transforming liquid formulations were explored recently by Zhang and co-workers, who developed an oil that transitions to a gel, *in situ*, to treat oral mucositis²⁶⁴. Oral mucositis is a common complication of cancer therapy, occurring in up to 80% of patients receiving high-dose radiotherapy³⁴⁷. It is also a serious complication, possibly resulting in anorexia and significant weight loss due to dysphagia, dehydration, and dysgeusia, brought about by the severe pain experienced by patients suffering from oral mucositis³⁴⁸. The rationale behind the development of their *in situ* gel-forming oil was to provide treatment in the form of physical shielding, while α -lipoic acid (solubilized in the oil) is slowly released from the resulting gel, acting a natural antioxidant. Their results showed that, after coming into contact with saliva, their oil was able to rapidly transition into a gel, thereby adhering to the oral mucosa. Their oil also showed excellent adhesion energies of 3.9 ± 0.2 N and 60 ± 2 J/m², on excised porcine buccal tissue. In their oral mucositis mouse model, application of their oil showed improved mucosal wound healing, inhibition of pro-inflammatory cytokines, and new granulation around the wounds²⁶⁴. The results therefore suggest that an *in situ* gel-forming oil, acting as a carrier for lipophilic drugs, can be an exciting approach in the treatment of oral mucositis, and possible other wounds in the oral cavity.

7. Liquid-based drug delivery approaches for vaginal administration

The first written records of vaginal drug administration practices date back to ancient Egypt, specifically circa 1850 BCE, where the vaginal route is considered one of five main drug administration

routes^{349,350}. Vaginal drug administration is an increasingly promising route for drug delivery despite its dynamic physiological environment (as displayed by Fig. 11), particularly for drugs with unfavorable physicochemical properties, such as poor aqueous solubility, which negatively affects oral bioavailability^{239,351}.

Vaginal drug absorption into the peripheral circulation is mediated by a venous plexus directing drugs to the hemorrhoidal and internal iliac veins^{351,355,356}. Hence, vaginal drug administration can bypass first-pass hepatic metabolism, thereby enabling the use of lower drug doses while improving therapeutic efficacy^{351,355,356}. Importantly, localized drug delivery to the uterus can be facilitated by vaginal administration due to what is known as the uterine first-pass effect^{12,83,354,357}. This phenomenon facilitates targeted drug delivery to the uterus *via* a counter-current drug exchange between the arterial and venous networks involved in both vaginal and endometrial perfusion⁸³. The literature describes the uterine first-pass effect as an underutilized mechanism in the realm of female reproductive health³⁵⁴. By leveraging the uterine first-pass effect, various diseases of the upper female genital tract can be effectively treated, minimizing the side effects associated with systemic absorption^{12,83,354,357}. This approach could also allow for targeted treatment during pregnancy by managing conditions like premature birth³⁵⁸.

Egyptian gynecological records reveal that the vaginal route of administration was reserved for practices such as abortion, pregnancy prevention, and treatment of pregnancy-related health conditions implying that the Egyptians delivered therapeutics *via* the uterine first-pass effect^{349,350}. Generally, modern-day vaginal formulations are designed to elicit local effects such as antimicrobial, antifungal, or spermicide activity³⁵⁹. Additionally, there is a growing interest in using vaginal drug delivery systems for microbicides to prevent sexually transmitted infections, including those caused by the human immunodeficiency virus, human papillomavirus, and herpes simplex virus³⁵⁹. When developing liquid-based formulations for vaginal administration, it is crucial to consider how excipients might affect the physiological balance of the vaginal microbiome as the acidic pH of the vaginal environment is maintained by lactic acid-producing bacteria

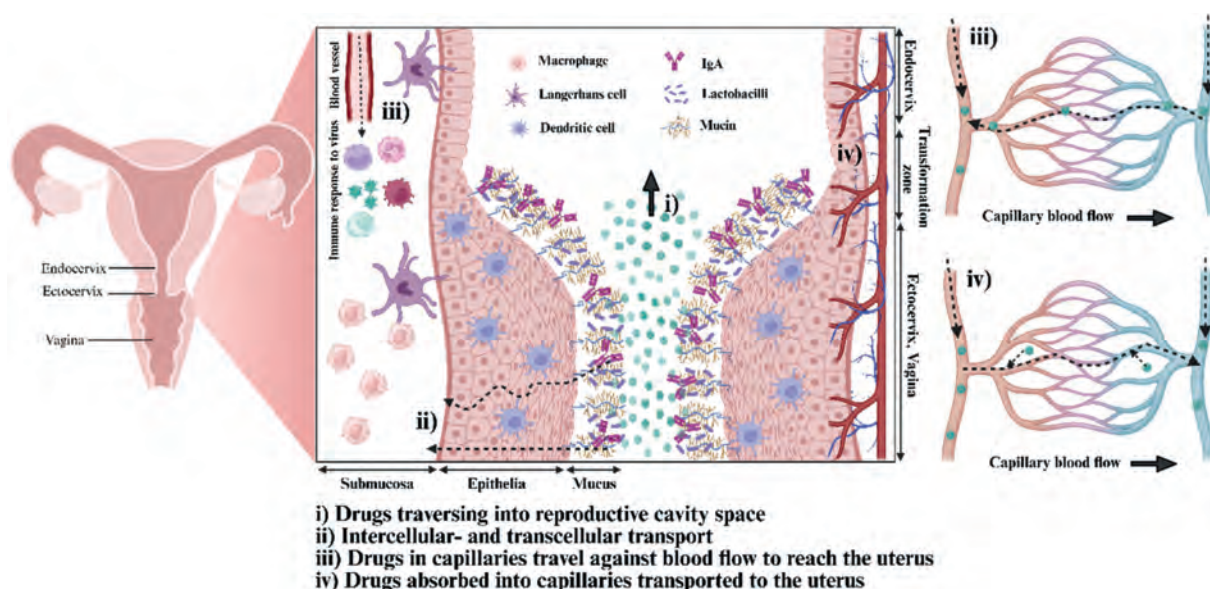


Figure 11 Mechanisms of the uterine first-pass effect following vaginal drug administration^{352–354}. Created with BioRender.

species³⁶⁰. Therefore, disrupting the natural protective microbiota can lead to infections while attempting to treat another condition. Furthermore, maximizing drug efficacy requires delivery strategies that minimize leakage, reduce user discomfort, limit local irritation, and take into account physiological changes, such as menopause or sexual intercourse, that can affect vaginal pH, subsequently impacting drug delivery^{238,361–363}.

Significant advancements have been made in the development of liquid-based formulations aimed at improving drug retention in vaginal mucosa^{118,238,346,359,364}. Retention within the vaginal epithelium can be facilitated by controlled release from liquid-based dosage forms to reduce dosing intervals while optimizing localized therapeutic efficacy. Typically, traditional semi-solid dosage forms fall short due to leakage necessitating frequent dosing²³⁸. This is especially concerning when treating localized fungal and bacterial infections as antimicrobial resistance can develop ultimately requiring systemic treatment alternatives³⁶⁵.

7.1. Emulsions

A study by Campaña-Seoane and co-workers incorporated ciprofloxacin in mucoadhesive water in silicone emulsions to study its absorption, distribution, and vaginal residence time in an *in vivo* rat model preceded by *in vitro* release experiments³⁶⁶. *In vitro* drug release profiles demonstrated zero-order kinetics with constant ciprofloxacin concentrations being released over a minimum period of 6 h. The *in vivo* data revealed significant distribution to the uterus and ovaries within 24 h of intravaginal administration. Water in silicone emulsions contains no oil phase but instead harbors water or aqueous saline solutions within a non-polar external phase comprising low molecular weight silicone-based polymers³⁶⁶. Silicone promotes tissue adhesion to withstand clearance by vaginal fluid in a dynamic physiological environment. Compared to hydrogels water in silicone emulsions provides an attractive mucoadhesive alternative since hydrogels swell following adhesion to mediate drug permeation. However, the swelling mechanism can give rise to structural deformity of the dosage form and reduced bioadhesion.

7.2. Phase transforming liquid formulations

As an additional alternative to hydrogels, the literature reports the incorporation of doxycycline in a mucoadhesive, thermos-responsive sol-gel to avoid sexually transmitted infections. The sol-gels were optimized *via* preparation in a buffered medium mimicking vaginal pH and osmolarity. The authors reported that mucoadhesion was unaffected by vaginal fluid secretions and that gelling was maintained at body temperature to ensure protection against vaginal infections by sustaining minimum inhibitory concentrations of doxycycline¹⁷¹.

Thermo-responsive gels present the opportunity for vaginal drug administration as a liquid can be inserted, but leakage is avoided due to transition to a gel after administration. Gel transitions can be mediated by including thermosensitive polymers above their critical gel concentration. Interestingly, a study by Shapiro and co-workers reported utilizing a hypotonic liquid combined with a thermosensitive polymer below its critical gelling concentration to improve the initial spreadability of a uniform and thin fluid layer³⁶⁷. As the polymer was included below the critical gelling concentration, gel formation only occurred upon water absorption as procured from the vaginal mucosa to restore the osmotic equilibrium. This study also reported reduced

cytotoxicity compared to a gel formulation and that the fluid formulations were compatible with the vaginal microbiota³⁶⁷. Spray gels have also emerged as adaptable vaginal dosage forms as thermosensitive polymer combinations can be used to capitalize on the pH influence of the vaginal environment by considering the inclusion of a pH-responsive polymer like chitosan while combining it with other thermos-responsive polymers to mediate mucoadhesion such as hydroxypropylmethylcellulose²⁶⁵. Additionally, spray gels hold the advantages of covering a larger surface area in a short time frame. A recent study revealed that a spray gel released the highest cumulative percentage of miconazole nitrate (*i.e.*, 81.72%) compared to a commercially available cream and suspension exhibiting a release percentage of 57.63% and 34.03%, respectively²⁶⁵.

In contrast to fluids transpiring into stimuli-responsive gels, vehicles like fluid gels are known to exhibit reduced viscosity upon shear application which can simplify handling due to their semi-solid nature while reversing to a fluid consistency upon vaginal application. These formulations retain their semi-solid form after the force of application has been removed to avoid leakage. Recently, Robinson and co-workers developed a fluid gel from alginate gelation while including acetic acid as an antimicrobial as well as a gelling agent³⁶⁸. These fluid gels demonstrated increased surface coverage due to their improved sprayability rendered by a discretized microstructure characterized by elevated elasticity and increased viscosity. Despite increased viscosity, the fluid gels were easily sprayed due to shear application causing fluid formation. This study investigated acetic acid fluid gels for the treatment of burn wounds³⁶⁸. However, acetic acid has been utilized to locally treat infections of the vagina and ears³⁶⁸. Signifying that this study can provide a foundation for future acetic acid fluid gels destined for vaginal administration, especially in low-resource environments.

7.3. Nanoemulsions

Natural active ingredients such as essential oils have also garnered scientific attention in transvaginal drug delivery due to their favorable antifungal and antibacterial properties³⁶⁹. Essential oils are interesting alternatives to laboratory-synthesized drugs as the literature fails to report antimicrobial resistance to these compounds due to their bioactive composition³⁷⁰. However, essential oils present stability challenges when exposed to heat, oxygen, humidity and light³⁶⁹. Moreover, these are volatile substances requiring a lipophilic vehicle to compensate for their poor aqueous solubility³⁶⁹. Interestingly, Gündel and co-workers investigated nanoemulsions as a lipophilic carrier to improve the instability of eucalyptus/lemongrass essential oils for treatment of vulvovaginal candidiasis³⁶⁹. An *in vivo* murine model demonstrated that the free form of considered essential oils exhibited no antifungal activity and the nanoemulsions harboring essential oils demonstrated superior antifungal activity compared to the miconazole treatment of the control group³⁶⁹.

Apart from utilizing nanoemulsions as a vehicle for natural ingredients, nanoemulsions also allow incorporation of natural ingredients to enhance the therapeutic action of laboratory-synthesized drugs. As an example, Ho and co-workers developed a squalene-based nanoemulsion as an adjuvant to a model immunogen vaccination in mice³⁷¹. This is an emerging field of research to induce mucosal immunity in the lower genital tract after local antigen administration to create immunity against genital infections. The squalene-based nanoemulsion increased

retention time in the vaginal mucosa while facilitating humoral and cellular immunity on an antigen-specific basis³⁷¹. Emulsions can be described as a very versatile vehicle due to their tailorability by including different stabilizing agents. Generally, surfactants are included for this purpose. However, a recent publication by Zhao and co-workers reported the innovation of graphene oxide quantum dot inclusion to replace surfactants in a pickering emulsion intended to enhance the immune response to a Pgp3 recombinant vaccine against *Chlamydia trachomatis*²⁰. The study reported no mucosal immune response in the lower genital tract. However, this study provides an interesting approach to future intravaginal drug administration by substituting surfactants, that can disrupt the vaginal microbiome, with nanocomposites offering superior biodegradability and biocompatibility. Moreover, the pickering emulsion was produced by a simple sonication method which is also beneficial for upscaling purposes²⁰.

7.4. Self-emulsifying drug delivery systems

The literature reports the development of SEDDSs for intravaginal application that have zeta potential changing behavior, enhanced self-emulsification capacity to enable spontaneous emulsification upon introduction to small vaginal fluid volumes and mucoadhesive properties²⁵⁴. Zeta potential changing SEDDSs developed by Nazir and co-workers, was achieved *via* a flip-flop mechanism. This mechanism improved mucus permeation and epithelial uptake by initially introducing SEDDSs harboring a highly negative zeta potential from surface hydrophilic phosphate substrates to accelerate mucosal permeation. Upon arriving at the epithelia phosphate substrates are cleaved by alkaline phosphatase rendering a “flip out” of amino substructures on oily droplet surfaces to amplify cellular uptake due to the slightly negative zeta potential change²⁵⁴. On the other hand, Köllner and co-workers reported curcumin containing SEDDSs having increased spreadability, mucosal permeation, and curcumin-protecting properties. These SEDDSs had slightly negative zeta potentials to mediate mucosal crossing of droplets larger than 200 nm¹⁴². Moreover, compared to a commercial O/W cream SEDDSs exhibited up to 68% mucosal coverage compared to 4% covered by the commercial cream. Swift spontaneous emulsification upon introduction to small fluid volumes by SEDDSs is imperative to ensure lipophilic drug release at mucosal surfaces¹⁴². Rohrer and co-workers evaluated 10 SEDDSs and concluded that a minimum final concentration of $\leq 40\%$ diluted SEDDS will enable self-emulsification. Emulsification time can vary from 30 min to 30 s depending on SEDDS composition¹⁵². Thereby, signifying that rapid spontaneous emulsification in small fluid volumes makes SEDDSs a promising vehicle for transvaginal drug delivery. However, the literature also reports that prolonged exposure to surfactants can disrupt the gut microbiome leading to long-term health concerns¹⁵¹. Therefore, a study by Jirwankar and co-workers investigated the effect of spontaneous nanoemulsions on *Lactobacillus acidophilus* concluding that depending on the surfactant concentration growth retardation of this probiotic bacteria could be observed. Surfactants could be ordered as Capmul® MCM > Maisine® CC > Miglyol® 810 > Kolliphor® RH40 to describe their cytotoxic impact and growth inhibition capacity³⁷².

7.5. Liquid crystals

Liquid crystals as self-assembled liquid composites are underexplored as a potential vaginal drug delivery vehicle. Jie and co-

workers incorporated sinomenine also known as the main alkaloid component present in Qingfengteng, a Chinese herb, into liquid crystal gel to target cervical cancer, provide pain relief and reduce inflammation following vaginal administration²⁸². Phytantriol, a well-known lyotropic liquid crystal material often utilized in personal care products, was included as the sinomenine-loading matrix. *In situ* cubic liquid crystal gels were developed exhibiting a gel transition time of 8.92 s within small vaginal fluid volumes. Moreover, *in vitro* release studies based on the dynamic dialysis bag method confirmed sinomenine hydrochloride sustained release ranging 144 h. Additionally, the *in vivo* experiments demonstrated that cubic liquid crystal gels were retained in vaginal mucosa of healthy adult female Sprague Dawley rats for longer than 12 h²⁸².

7.6. Eutectic systems

Limited studies have reported the application of deep eutectic systems or eutectic mixtures for vaginal administration. However, a single study reported investigating the possibility of a deep eutectic system formation between metronidazole and hydrophilic agents to enhance drug release *via* an enhanced dissolution rate in the presence of small fluid volumes³⁷³. This is an interesting concept as the idea is based on a non-mucoadhesive principle. The authors, therefore, suggested that the deep eutectic system should be incorporated into a solid vaginal insert to simplify handling and administration. Results indicated that metronidazole was released in a controlled fashion over a period of 7 days in simulated vaginal fluid. Thereby, signifying that deep eutectic systems capable of controlled release can effectively replace oral metronidazole therapy³⁷³.

8. Clinical translation of liquid-based transmucosal drug delivery systems

Practical and ethical challenges often obscure academic innovations from reaching commercialization³⁷⁴. Therefore, despite the advances made to mediate the crossing of mucosal barriers, clinical translation of these dosage forms presents numerous obstacles to overcoming bench-to-bedside transitions. The seven steps of clinical translation refer to: (a) up-scaling potential, (b) standardized characterization procedures, (c) stability analysis, (d) mode of action, (e) establishing safe and effective drug doses, (f) Good Laboratory Practices (GLP) toxicity evaluations, and (g) clinical studies involving human subjects³⁷⁴. Academic innovations are generally not constrained by the seven steps of clinical translation³⁷⁴. Thereby, implying that many significant scientific discoveries remain academic knowledge instead of changing the lives of patients in clinical settings. In most cases, liquid-based drug delivery systems lack standardized protocols considering the efficacy and safety which hinders regulatory approval due to the limited availability of extensive data on toxicity for patients, researchers involved in dosage form development as well as the impact on the environment if numerous patients are consuming these systems due to factors such as increased surfactant content and nano-sized droplets^{12,139,374–378}. In terms of scalability, nanoemulsion development remains challenging as minute process differences can drastically impact physicochemical properties³⁷⁷. Thus, compromising the safety and even quality of nano vehicles. The high-energy production methods of nanoemulsions also impede upscaling as it is not cost-

effective³⁷⁷. Apart from high-energy methods, the incorporation of surfactants can also negatively impact the long-term biological safety of these dosage forms³⁷⁷.

It is understandable that SEDDSs have garnered significant scientific attention, as no high-energy production methods are required to produce these systems and upscaling is more realistic compared to nanoemulsions. Importantly, the stability challenges of liquid SEDDSs can be conquered by solidifying them *via* melt extrusion, freeze drying, and adsorption onto solid carriers^{379–381}. This allows for the combining of liquid-based SEDDS advantages with the enhanced stability, simplified handling, and portability of solid dosage forms¹³⁹. In terms of oral drug delivery, liquid-based systems can be administered by filling soft or hard gelatine capsules as is the case with Norvir® (ritonavir), Sandimmune® (cyclosporine), and Fortovase® (saquinavir), established on liquid-based formulations^{139,382}. However, the authors suggest that the solidification of liquid-based drug delivery systems can be considered to allow reconstitution with a pre-determined water quantity prior to transmucosal administration to gain the advantages of localized liquid-based transmucosal drug delivery without compromising on dosage form stability. This can potentially simplify regulatory assessment and clinical translation of certain liquid-based dosage forms.

The literature reports numerous tools of optimization and characterization of ionic liquid-based systems^{376,383–386}. However, the reproducibility of liquid-based drug delivery systems such as liquid crystals, ionic liquids, and eutectic mixtures also causes a delay in clinical translation of these systems^{376,387–389}. This delay can be attributed to inadequate stability of formulations combined with the absence of clear guidelines for their stability. Moreover, the presence of potential impurities and unknown extent of bio-distribution also hinder their commercialization³⁷⁶. Importantly, liquid crystals are also considered as a building block for nanoparticle formulation³⁹⁰. Therefore, the clinical translation of these liquid-based systems is also restricted due to the absence of standardized guidelines by the United States Food and Drug Administration (FDA), Central Drugs Standard Control Organization, India (CDSCO), European Medicines Agency (EMA) which are the main regulatory bodies responsible for the approval of nano-therapeutics before reaching the market³⁷⁷. It should also be emphasized that tuning and production of novel ionic liquid-based systems necessitates a cumulative understanding to consider the biological implications of these systems, as exceptional safety and targeted delivery should remain imperative before allowing these systems to reach the market³⁹¹. Hence, due to the vast applications of ionic liquids in cancer, antimicrobial, adjuvant, and drug delivery innovations substantial validation of preclinical and clinical data is essential yet absent. Moreover, extrapolating data obtained from animal studies to humans also forms a complex part of clinical data collection due to interspecies variability³⁹¹. Clinical translation of these systems is also handicapped due to the absence of standardized safety and efficacy protocols despite their reported superiority to marketed products³⁷⁵. As an example, a recent randomized controlled trial considering the delivery of ketoconazole incorporated into choline geranate formulations rendered superior treatment outcomes when compared to the commercially marketed product, Daktarin®³⁹². Remarkably, the ionic liquid-based formulation incorporated a lower dose of ketoconazole while demonstrating superior efficacy to Daktarin® treatment of Tinea Pedis infection³⁹². Thereby, signifying the potential of these systems despite their scarce commercialization.

9. Conclusions and prospects

Localized, targeted drug delivery approaches provide an important alternative to oral drug delivery as these innovative drug delivery approaches can compensate for poor pharmacokinetic properties such as limited aqueous solubility to ensure controlled drug delivery at target sites^{73,357,393}. Mucosal surfaces provide unique challenges to drug delivery due to different dynamic physiological environments^{176,268,356,394}. Despite the challenging mucosal surface environments, numerous advancements have been made regarding liquid-based drug delivery from anchoring to mucosal surfaces to mediating the crossing of mucosal barriers *via* innovative drug delivery approaches^{165,170,176,269,364}. This review considered novel liquid-based drug delivery approaches involving nanoemulsions, SEDDSs, double-emulsions, phase-transforming liquid formulations, ionic liquid-based formulations, liquid crystals, and eutectic systems to cross mucosal surfaces. Although most of these drug delivery systems were first discovered during the last century, there is a scientific upsurge regarding liquid-based drug delivery systems, especially in terms of transmucosal drug delivery as liquids provide improved spreadability and adhesion at mucosal surfaces. Moreover, presenting a drug in solubilized form at mucosal surfaces can generate rapid uptake as the rate-limiting step of dissolution in small fluid volumes can be bypassed. Liquid-based drug delivery systems are also advancing due to innovative vaccine developments intended for mucosal administration or as vaccine adjuvants to generate mucosal immunity^{20,31,94,223,320}.

Another advantage of liquid-based drug delivery systems is the possibility of solidifying these vehicles to improve stability, prolong shelf-life, and simplify upscaling by subjecting formulations like SEDDSs to freeze drying, hot-melt extrusion, spray drying, wet granulation, and adsorption to solid carriers^{380,382}. Another interesting future application for liquid-based drug delivery systems is to use liquid vehicles as materials for three-dimensional dosage form printing²⁹⁷. Recent advances in the deposition of liquids in orodispersible film development describe utilizing semi-solid formulations or liquids exhibiting high viscosity that are subjected to material extrusion to produce orodispersible films²⁹⁷. Additionally, recent fluid-based three-dimensional printing of contact lenses has also garnered scientific attention as contact lenses can provide sustained release in ophthalmic therapy³⁹⁵. Currently, gelatine methacrylate combined with PEG diacrylate or hyaluronic acid is the material of choice then processed *via* material extrusion to provide a base for anti-inflammatory, local antibiotic, and beta antagonist administration²⁹⁷. The main challenge here lies in creating biocompatible contact lenses without causing vision impairment²⁹⁷.

Apart from three-dimensional dosage form printing, magneto-responsive ferrofluids are also emerging as colloidal suspensions harboring magnetic nanoparticles in an oil or water medium while stabilized against nanoparticle agglomeration by surfactant inclusion³⁹⁶. The magnetic manipulation potential of ferrofluids can especially benefit targeted drug delivery during cancer therapy. Limited studies report using ferrofluids in transmucosal dosage forms due to potential mucosal irritation. However, the literature does report the intranasal administration of magnetic nanoparticles and separate publications report utilizing ferrofluids during nanoparticle formulation to improve nanoparticle targeting capacity^{397,398}. Moreover, a recent study reported the stabilization of a double-emulsion comprising ferrofluids by permanent magnet application³⁹⁹. Ferrofluids are also employed in liquid robotics as their morphology and multimodal locomotion can be controlled

via external stimuli to guide liquid-based drug delivery⁴⁰⁰. This review mentions the potential of ferrofluids as a future prospect for liquid-based drug delivery system development, but also contemplates that the material properties such as viscosity, polarity, and magnetic responsiveness of these systems can drastically impact up-scaling and long-term stability. Moreover, these magnetic fluid systems lack extensive safety evaluations, similar to more established liquid-based drug delivery systems, which implies that these systems are even further from successful clinical translation compared to dosage forms such as SEDDSs and eutectic systems.

The most notorious shortfall of liquid-based drug delivery systems is their slow progress regarding clinical translation despite advanced academic innovations. Hence, meaningful directions for future studies include comparing the efficacy of novel innovations to marketed products to demonstrate their superiority³⁹². Moreover, one must bear in mind the importance of studies that evaluate the safety, toxicity, and environmental impact of liquid-based drug delivery systems as extensive data is needed to assist regulatory bodies such as the FDA, CDSCO, and EMA in establishing guidelines and protocols for their clinical translation.

Acknowledgements

The authors would like to thank the Centre of Excellence for Pharmaceutical Sciences, Faculty of Health Sciences, North-West University, South Africa, for the financial contribution to this project.

Author contributions

Daniëlle van Staden: Writing - original draft, Writing, review and editing, Visualization, Project administration, Funding; Hendrik J.R. Lemmer: Writing - original draft, Writing, review and editing, Funding; Wilna Liebenberg: Writing - original draft, Writing, review and editing, Funding; Josias H. Hamman: Writing - original draft, Writing, review and editing, Conceptualization, Funding. All authors have read and approved the final manuscript.

Conflicts of interest

The authors have no conflicts of interest to declare.

References

- Zhao Z, Ukidve A, Kim J, Mitragotri S. Targeting strategies for tissue-specific drug delivery. *Cell* 2020;**181**:151–67.
- Ramström O, Lehn J-M. Drug discovery by dynamic combinatorial libraries. *Nat Rev Drug Discov* 2002;**1**:26–36.
- Topol EJ. High-performance medicine: the convergence of human and artificial intelligence. *Nat Med* 2019;**25**:44–56.
- Macarron R, Banks MN, Bojanic D, Burns DJ, Cirovic DA, Garyantes T, et al. Impact of high-throughput screening in biomedical research. *Nat Rev Drug Discov* 2011;**10**:188–95.
- Sun D, Gao W, Hu H, Zhou S. Why 90% of clinical drug development fails and how to improve it?. *Acta Pharm Sin B* 2022;**12**:3049–62.
- Lin A, Giuliano CJ, Palladino A, John KM, Abramowicz C, Yuan M Lou, et al. Off-target toxicity is a common mechanism of action of cancer drugs undergoing clinical trials. *Sci Transl Med* 2019;**11**:eaaw8412.
- Liu R, Zuo R, Hudalla GA. Harnessing molecular recognition for localized drug delivery. *Adv Drug Deliv Rev* 2021;**170**:238–60.
- Yang R, Wei T, Goldberg H, Wang W, Cullion K, Kohane DS. Getting drugs across biological barriers. *Adv Mater* 2017;**29**:1606596.
- Yu L, Liu S, Jia S, Xu F. Emerging frontiers in drug delivery with special focus on novel techniques for targeted therapies. *Biomed Pharmacother* 2023;**165**:115049.
- Lock JY, Carlson TL, Carrier RL. Mucus models to evaluate the diffusion of drugs and particles. *Adv Drug Deliv Rev* 2018;**124**:34–49.
- Cu Y, Saltzman WM. Mathematical modeling of molecular diffusion through mucus. *Adv Drug Deliv Rev* 2009;**61**:101–14.
- van Staden D, Gerber M, Lemmer HJR. The application of nano drug delivery systems in female upper genital tract disorders. *Pharmaceutics* 2024;**16**:1475.
- Leal J, Smyth HDC, Ghosh D. Physicochemical properties of mucus and their impact on transmucosal drug delivery. *Int J Pharm* 2017;**532**:555–72.
- Zhang W-X, Gao Y-R, Xue R, Nguyen W, Chen W, Wang JH, et al. Liquid formulations based on ionic liquids in biomedicine. *Mater Today Phys* 2023;**30**:100925.
- Moreira M, Sarraça M. How can oral paediatric formulations be improved? A challenge for the XXI century. *Int J Pharm* 2020;**590**:119905.
- Khan D, Kirby D, Bryson S, Shah M, Rahman Mohammed A. Paediatric specific dosage forms: patient and formulation considerations. *Int J Pharm* 2022;**616**:121501.
- Racaniello GF, Silvestri T, Pistone M, D'Amico V, Arduino I, Denora N, et al. Innovative pharmaceutical techniques for paediatric dosage forms: a systematic review on 3D printing, prilling/vibration and microfluidic platform. *J Pharmacol Sci* 2024;**113**:1726–48.
- Mushtaq A, Mohd Wani S, Malik AR, Gull A, Ramniwas S, Ahmad Nayik G, et al. Recent insights into nanoemulsions: their preparation, properties and applications. *Food Chem X* 2023;**18**:100684.
- Tenorio-Garcia E, Araiza-Calahorra A, Simone E, Sarkar A. Recent advances in design and stability of double emulsions: trends in pickering stabilization. *Food Hydrocoll* 2022;**128**:107601.
- Zhao L, Shu M, Shi K, Tang S, Li Z. Novel use of graphene oxide quantum dots in a pickering emulsion as a *Chlamydia trachomatis* vaccine adjuvant. *Int Immunopharmacol* 2023;**118**:110035.
- van Staden D, Haynes RK, Viljoen JM. The development of dermal self-double-emulsifying drug delivery systems: preformulation studies as the keys to success. *Pharmaceutics* 2023;**16**:1348.
- Efiana NA, Phan TNQ, Wicaksono AJ, Bernkop-Schnürch A. Mucus permeating self-emulsifying drug delivery systems (SEDDS): about the impact of mucolytic enzymes. *Colloids Surf B Biointerfaces* 2018;**161**:228–35.
- Shoaib A, Mangla B, Javed S, Sultan MH, Alqahtani SS, Shakeel F. Vicissitudes of liquid crystals for solubility enhancement of poorly soluble drugs. *J Mol Liq* 2021;**321**:114924.
- McCrary PD, Beasley PA, Gurau G, Narita A, Barber PS, Cojocaru OA, et al. Drug specific, tuning of an ionic liquid's hydrophilic–lipophilic balance to improve water solubility of poorly soluble active pharmaceutical ingredients. *New J Chem* 2013;**37**:2196.
- Moshikur RM, Ali MK, Moniruzzaman M, Goto M. Recent advances in surface-active ionic liquid-assisted self-assembly systems for drug delivery. *Curr Opin Colloid Interface Sci* 2021;**56**:101515.
- Tambe S, Jain D, Meruva SK, Rongala G, Juluri A, Nihalani G, et al. Recent advances in amorphous solid dispersions: preformulation, formulation strategies, technological advancements and characterization. *Pharmaceutics* 2022;**14**:2203.
- Baghel S, Cathcart H, O'Reilly NJ. Polymeric amorphous solid dispersions: a review of amorphization, crystallization, stabilization, solid-state characterization, and aqueous solubilization of biopharmaceutical classification system class II drugs. *J Pharmacol Sci* 2016;**105**:2527–44.
- França MT, Martins Marcos T, Costa PFA, Bazzo GC, Nicolay Pereira R, Gerola AP, et al. Eutectic mixture and amorphous solid

- dispersion: two different supersaturating drug delivery system strategies to improve griseofulvin release using saccharin. *Int J Pharm* 2022;**615**:121498.
29. Desu HR, Narang AS, Kumar V, Thoma LA, Mahato RI. Liquid dosage forms. In: Dash A, Singh S, editors. *Pharmaceutics*. Amsterdam: Elsevier; 2024. p. 271–318.
 30. Lu RM, Hwang YC, Liu IJ, Lee CC, Tsai HZ, Li HJ, et al. Development of therapeutic antibodies for the treatment of diseases. *J Biomed Sci* 2020;**27**:1–30.
 31. Rivera-Izquierdo M, Morales-Portillo A, Guerrero-Fernández de Alba I, Fernández-Martínez NF, Schoenenberger-Arnaiz JA, Barranco-Quintana JL, et al. Vaccination strategies for patients under monoclonal antibody and other biological treatments: an updated comprehensive review based on EMA authorisations to January 2024. *Expert Rev Vaccines* 2024;**23**:887–910.
 32. Grairi M, Le Borgne M. Antibody–drug conjugates: prospects for the next generation. *Drug Discov Today* 2024;**29**:104241.
 33. Bruschi M. Drug delivery systems. In: Bruschi M, editor. *Strategies to modify the drug release from pharmaceutical systems*. Amsterdam: Elsevier; 2015. p. 87–194.
 34. Lam JKW, Cheung CCK, Chow MYT, Harrop E, Lapwood S, Barclay SIG, et al. Transmucosal drug administration as an alternative route in palliative and end-of-life care during the COVID-19 pandemic. *Adv Drug Deliv Rev* 2020;**160**:234–43.
 35. Zhang H, Shahbazi MA, Almeida PV, Santos HA. Mucus as a barrier for biopharmaceuticals and drug delivery systems. In: das Neves J, Sarmento B, editors. *Mucosal delivery of biopharmaceuticals*. Boston: Springer US; 2014. p. 59–97.
 36. Leceta J, Del Campo R, Jordan S, Klose CSN. Editorial: immunoregulation at mucosal surfaces. *Front Immunol* 2022;**13**:983201.
 37. Bansil R, Turner BS. The biology of mucus: composition, synthesis and organization. *Adv Drug Deliv Rev* 2018;**124**:3–15.
 38. Netsomboon K, Bernkop-Schnürch A. Mucoadhesive vs. mucopetrating particulate drug delivery. *Eur J Pharm Biopharm* 2016;**98**:76–89.
 39. Bandi SP, Bhatnagar S, Venuganti VVK. Advanced materials for drug delivery across mucosal barriers. *Acta Biomater* 2021;**119**:13–29.
 40. Vinod A, Tadmor R, Katoshevski D, Gutmark EJ. Gels that serve as mucus simulants: a review. *Gels* 2023;**9**:555.
 41. Menzel C, Bernkop-Schnürch A. Enzyme decorated drug carriers: targeted swords to cleave and overcome the mucus barrier. *Adv Drug Deliv Rev* 2018;**124**:164–74.
 42. Authimoolam S, Dziubla T. Biopolymeric mucin and synthetic polymer analogs: their structure, function and role in biomedical applications. *Polymers (Basel)* 2016;**8**:71.
 43. Lai SK, O'Hanlon DE, Harrold S, Man ST, Wang Y-Y, Cone R, et al. Rapid transport of large polymeric nanoparticles in fresh undiluted human mucus. *Proc Natl Acad Sci U S A* 2007;**104**:1482–7.
 44. McGuckin MA, Thornton DJ, Whitsett JA. Mucins and mucus. In: Mestecky J, Strober W, Russell M, Kelsall B, Cheroutre H, Lambrecht B, editors. *Mucosal immunology*. Amsterdam: Elsevier; 2015. p. 231–50.
 45. Laffleur F, Hintzen F, Shahnaz G, Rahmat D, Leithner K, Bernkop-Schnürch A. Development and *in vitro* evaluation of slippery nanoparticles for enhanced diffusion through native mucus. *Nanomedicine* 2014;**9**:387–96.
 46. Witten J, Samad T, Ribbeck K. Selective permeability of mucus barriers. *Curr Opin Biotechnol* 2018;**52**:124–33.
 47. Zhou X, Wu Y, Zhu Z, Lu C, Zhang C, Zeng L, et al. Mucosal immune response in biology, disease prevention and treatment. *Signal Transduct Targeted Ther* 2025;**10**:7.
 48. Zierden HC, Josyula A, Shapiro RL, Hsueh HT, Hanes J, Ensign LM. Avoiding a sticky situation: bypassing the mucus barrier for improved local drug delivery. *Trends Mol Med* 2021;**27**:436–50.
 49. Dong W, Ye J, Zhou J, Wang W, Wang H, Zheng X, et al. Comparative study of mucoadhesive and mucus-penetrative nanoparticles based on phospholipid complex to overcome the mucus barrier for inhaled delivery of baicalein. *Acta Pharm Sin B* 2020;**10**:1576–85.
 50. Wanasathop A, Patel PB, Choi HA, Li SK. Permeability of buccal mucosa. *Pharmaceutics* 2021;**13**:1814.
 51. Aframian D, Davidowitz T, Benoliel R. The distribution of oral mucosal pH values in healthy saliva secretors. *Oral Dis* 2006;**12**:420–3.
 52. Lin YP, Chen WC, Cheng CM, Shen CJ. Vaginal pH value for clinical diagnosis and treatment of common vaginitis. *Diagnostics* 2021;**11**:1996.
 53. Gaudana R, Ananthula HK, Parenky A, Mitra AK. Ocular drug delivery. *AAPS J* 2010;**12**:348–60.
 54. Koparal M, Kurt E, Altuntas EE, Dogan F. Assessment of mucociliary clearance as an indicator of nasal function in patients with COVID-19: a cross-sectional study. *Eur Arch Otorhinolaryngol* 2021;**278**:1863–8.
 55. Watanabe S. Salivary clearance from different regions of the mouth in children. *Caries Res* 1992;**26**:423–7.
 56. Argüeso P. Human ocular mucins: the endowed guardians of sight. *Adv Drug Deliv Rev* 2022;**180**:114074.
 57. Efron N, Young G, Brennan NA. Ocular surface temperature. *Curr Eye Res* 1989;**8**:901–6.
 58. Keck T, Leiacker R, Riechelmann H, Rettinger G. Temperature profile in the nasal cavity. *Laryngoscope* 2000;**110**:651–4.
 59. Malamed SF. Physical and psychological evaluation. In: Malamed S, editor. *Sedation*. Amsterdam: Elsevier; 2010. p. 23–62.
 60. Pandya AK, Vora LK, Umeyor C, Surve D, Patel A, Biswas S, et al. Polymeric *in situ* forming depots for long-acting drug delivery systems. *Adv Drug Deliv Rev* 2023;**200**:115003.
 61. Thapa R, Pandey P, Parat M-O, Gurung S, Parekh HS. Phase transforming *in situ* gels for sustained and controlled transmucosal drug delivery via the intravaginal route. *Int J Pharm* 2024;**655**:124054.
 62. Moradi S, Bikker FJ, Hesse D. Saliva composition from birth to adolescence: a systematic review of the literature. *J Oral Biosci* 2025;**67**:100661.
 63. Lacoste-Ferré M-H, Ober C, Samouillan V. Viscoelastic behavior of oral mucosa. A rheological study using small-amplitude oscillatory shear tests. *J Mech Behav Biomed Mater* 2023;**143**:105898.
 64. Nichols BA, Chiappino ML, Dowson CW, Proctor FI. Demonstration of the mucous layer of the tear film by electron microscopy. *Investig Ophthalmol Vis Sci* 1985;**26**:464–73.
 65. Beule AG. Physiology and pathophysiology of respiratory mucosa of the nose and the paranasal sinuses. *GMS Curr Top Otorhinolaryngol Head Neck Surg* 2010;**9**:Doc07.
 66. Di Stasio D, Lauritano D, Iquebal H, Romano A, Gentile E, Lucchese A. Measurement of oral epithelial thickness by optical coherence tomography. *Diagnostics* 2019;**9**:90.
 67. Cui C, Xiao Y, Lin E, Luo L, Sun X, Zeng J, et al. Precise measurement of the thickness of vaginal intraepithelial neoplasia. *J Low Genit Tract Dis* 2022;**26**:245–9.
 68. Wang X, Liu J, Zou J, Luo C, Wei D. The study of vaginal wall thickness in adults based on histopathological measurements. *Sci Rep* 2024;**14**:18644.
 69. Zilliox MJ, Bouchard CS. The microbiome, ocular surface, and corneal disorders. *Am J Pathol* 2023;**193**:1648–61.
 70. Tai J, Han MS, Kwak J, Kim TH. Association between microbiota and nasal mucosal diseases in terms of immunity. *Int J Mol Sci* 2021;**22**:4744.
 71. Pathak JL, Yan Y, Zhang Q, Wang L, Ge L. The role of oral microbiome in respiratory health and diseases. *Respir Med* 2021;**185**:106475.
 72. Chen X, Lu Y, Chen T, Li R. The female vaginal microbiome in health and bacterial vaginosis. *Front Cell Infect Microbiol* 2021;**11**:631972.
 73. Khan O, Bhawale R, Vasave R, Mehra NK. Ionic liquid-based formulation approaches for enhanced transmucosal drug delivery. *Drug Discov Today* 2024;**29**:104109.

74. Hazt B, Read DJ, Harlen OG, Poon WCK, O'Connell A, Sarkar A. Mucoadhesion across scales: towards the design of protein-based adhesives. *Adv Colloid Interface Sci* 2024;**334**:103322.
75. Rohrer J, Partenhauser A, Hauptstein S, Gallati CM, Matuszczak B, Abdulkarim M, et al. Mucus permeating thiolated self-emulsifying drug delivery systems. *Eur J Pharm Biopharm* 2016;**98**:90–7.
76. Abdulkarim M, Sharma PK, Gumbleton M. Self-emulsifying drug delivery system: mucus permeation and innovative quantification technologies. *Adv Drug Deliv Rev* 2019;**142**:62–74.
77. Krause M, Wheeler TL, Richter HE, Snyder TE. Systemic effects of vaginally administered estrogen therapy. *Female Pelvic Med Reconstr Surg* 2010;**16**:188–95.
78. Boddupalli B, Mohammed Zulkar NK, Nath R, Banji D. Mucoadhesive drug delivery system: an overview. *J Adv Pharm Technol Res* 2010;**1**:381.
79. Subramanian DA, Langer R, Traverso G. Mucus interaction to improve gastrointestinal retention and pharmacokinetics of orally administered nano-drug delivery systems. *J Nanobiotechnol* 2022;**20**:362.
80. Szentkuti L, Lorenz K. The thickness of the mucus layer in different segments of the rat intestine. *Histochem J* 1995;**27**:466–72.
81. Abrami M, Biasin A, Tescione F, Tierno D, Dapas B, Carbone A, et al. Mucus structure, viscoelastic properties, and composition in chronic respiratory diseases. *Int J Mol Sci* 2024;**25**:1933.
82. Jory M, Bellouma K, Blanc C, Casanellas L, Petit A, Reynaud P, et al. Mucus microrheology measured on human bronchial epithelium culture. *Front Phys* 2019;**7**:19.
83. Patel SK, Valicherla GR, Micklo AC, Rohan LC. Drug delivery strategies for management of women's health issues in the upper genital tract. *Adv Drug Deliv Rev* 2021;**177**:113955.
84. Wang TZ, Liu XX, Wang SY, Liu Y, Pan XY, Wang JJ, et al. Engineering advanced drug delivery systems for dry eye: a review. *Bioengineering* 2022;**10**:53.
85. Surapaneni MS, Das SK, Das NG. Effect of excipient and processing variables on adhesive properties and release profile of pentoxifylline from mucoadhesive tablets. *Drug Dev Ind Pharm* 2006;**32**:377–87.
86. Cook MT, Shorthouse D. Reconceptualising mucoadhesion for future medicines. *RSC Pharmaceutics* 2024;**1**:949–57.
87. Bayer IS. Recent advances in mucoadhesive interface materials, mucoadhesion characterization, and technologies. *Adv Mater Interfac* 2022;**9**:2200211.
88. Bartkowiak A, Lewandowicz J, Rojewska M, Krüger K, Lulek J, Prochaska K. Study of viscoelastic, sorption and mucoadhesive properties of selected polymer blends for biomedical applications. *J Mol Liq* 2022;**361**:119623.
89. Shaikh R, Raj Singh T, Garland M, Woolfson Ad, Donnelly R. Mucoadhesive drug delivery systems. *J Pharm BioAllied Sci* 2011;**3**:89.
90. Santos KP, Rodero CF, Ribeiro CM, Gremião MPD, Peccinini RG, Pavan FR, et al. Development of a mucoadhesive liquid crystal system for the administration of rifampicin applicable in tuberculosis therapy. *Life* 2022;**12**:1138.
91. Ahmed S, Amin MM, Sayed S. Ocular drug delivery: a comprehensive review. *AAPS PharmSciTech* 2023;**24**:66.
92. Hussain A, Ahsan F. The vagina as a route for systemic drug delivery. *J Control Release* 2005;**103**:301–13.
93. Biswas M, Nurunnabi M, Khatun Z. Understanding mucosal physiology and rationale of formulation design for improved mucosal immunity. *ACS Appl Bio Mater* 2024;**7**:5037–56.
94. Huang M, Zhang M, Zhu H, Du X, Wang J. Mucosal vaccine delivery: a focus on the breakthrough of specific barriers. *Acta Pharm Sin B* 2022;**12**:3456–74.
95. Jiao J, Zhang L. Influence of intranasal drugs on human nasal mucociliary clearance and ciliary beat frequency. *Allergy Asthma Immunol Res* 2019;**11**:306–19.
96. Beaven E, Kumar R, An JM, Mendoza H, Sutradhar SC, Choi W, et al. Potentials of ionic liquids to overcome physical and biological barriers. *Adv Drug Deliv Rev* 2024;**204**:115157.
97. Pangua C, Espuelas S, Martínez-Ohárriz MC, Vizmanos JL, Irache JM. Mucus-penetrating and permeation enhancer albumin-based nanoparticles for oral delivery of macromolecules: application to bevacizumab. *Drug Deliv Transl Res* 2024;**14**:1189–205.
98. Hauber HP, Daigneault P, Frenkiel S, Lavigne F, Hung HL, Levitt RC, et al. Niflumic acid and MSI-2216 reduce TNF- α -induced mucin expression in human airway mucosa. *J Allergy Clin Immunol* 2005;**115**:266–71.
99. Nafee N, Forier K, Braeckmans K, Schneider M. Mucus-penetrating solid lipid nanoparticles for the treatment of cystic fibrosis: proof of concept, challenges and pitfalls. *Eur J Pharm Biopharm* 2018;**124**:125–37.
100. Udabe J, Muñoz-Juan A, Tafech B, Orellano MS, Hedtrich S, Laromaine A, et al. Modulating the mucosal drug delivery efficiency of polymeric nanogels tuning their redox response and surface charge. *Adv Funct Mater* 2024;**34**:2407044.
101. Bej R, Haag R. Mucus-inspired dynamic hydrogels: synthesis and future perspectives. *J Am Chem Soc* 2022;**144**:20137–52.
102. Zhang R, Zhang L, Li P, Pang K, Liu H, Tian L. Epithelial barrier in the nasal mucosa, related risk factors and diseases. *Int Arch Allergy Immunol* 2023;**184**:481–501.
103. Hua T, Li S, Han B. Nanomedicines for intranasal delivery: understanding the nano-bio interactions at the nasal mucus-mucosal barrier. *Expert Opin Drug Deliv* 2024;**21**:553–72.
104. Zhang N, Van Crombruggen K, Gevaert E, Bachert C. Barrier function of the nasal mucosa in health and type-2 biased airway diseases. *Allergy* 2016;**71**:295–307.
105. Nur Husna SM, Tan H-TT, Md Shukri N, Mohd Ashari NS, Wong KK. Nasal epithelial barrier integrity and tight junctions disruption in allergic rhinitis: overview and pathogenic insights. *Front Immunol* 2021;**12**:663626.
106. Brunner J, Ragupathy S, Borchard G. Target specific tight junction modulators. *Adv Drug Deliv Rev* 2021;**171**:266–88.
107. Ramirez SI, Faraji F, Hills LB, Lopez PG, Goodwin B, Stacey HD, et al. Immunological memory diversity in the human upper airway. *Nature* 2024;**632**:630–6.
108. Johansson MEV. Fast renewal of the distal colonic mucus layers by the surface goblet cells as measured by *in vivo* labeling of mucin glycoproteins. *PLoS One* 2012;**7**:e41009.
109. Ozsoy Y, Gungor S, Cevher E. Nasal delivery of high molecular weight drugs. *Molecules* 2009;**14**:3754–79.
110. Tomazic PV, Darnhofer B, Birner-Gruenberger R. Nasal mucus proteome and its involvement in allergic rhinitis. *Expert Rev Proteomics* 2020;**17**:191–9.
111. Tong J, Gu Q. Expression and clinical significance of mucin gene in chronic rhinosinusitis. *Curr Allergy Asthma Rep* 2020;**20**:63.
112. Huang F, Liu F, Zhen X, Gong S, Chen W, Song Z. Pathogenesis, diagnosis, and treatment of infectious rhinosinusitis. *Microorganisms* 2024;**12**:1690.
113. Fasoli E. Protein corona: dr. Jekyll and mr. Hyde of nanomedicine. *Biotechnol Appl Biochem* 2020;**68**:1139–52.
114. Digiacoimo L, Pozzi D, Palchetti S, Zingoni A, Caracciolo G. Impact of the protein Corona on nanomaterial immune response and targeting ability. *Wiley Interdiscip Rev Nanomed Nanobiotechnol* 2020;**12**:e1615.
115. Rawlings BA, Higgins TS, Han JK. Bacterial pathogens in the nasopharynx, nasal cavity, and osteomeatal complex during wellness and viral infection. *Am J Rhinol Allergy* 2013;**27**:39–42.
116. Zheng B, Liu D, Qin X, Zhang D, Zhang P. Mucoadhesive-to-mucopenetrating nanoparticles for mucosal drug delivery: a mini review. *Int J Nanomed* 2025;**20**:2241–52.
117. Paul S, Bhuyan S, Balasoupramanien DD, Palaniappan A. Mucoadhesive and muco-penetrative formulations for the oral delivery of insulin. *ACS Omega* 2024;**9**:24121–41.
118. Rossi S, Vigani B, Sandri G, Bonferoni MC, Caramella CM, Ferrari F. Recent advances in the mucus-interacting approach for vaginal drug delivery: from mucoadhesive to mucus-penetrating nanoparticles. *Expert Opin Drug Deliv* 2019;**16**:777–81.

119. Hayes R, Warr GG, Atkin R. Structure and nanostructure in ionic liquids. *Chem Rev* 2015;**115**:6357–426.
120. Ludwig M, von Klitzing R. Recent progress in measurements of oscillatory forces and liquid properties under confinement. *Curr Opin Colloid Interface Sci* 2020;**47**:137–52.
121. Donaldson SH, Røyne A, Kristiansen K, Rapp MV, Das S, Gebbie MA, et al. Developing a general interaction potential for hydrophobic and hydrophilic interactions. *Langmuir* 2015;**31**:2051–64.
122. Hu YT, Ting Y, Hu JY, Hsieh SC. Techniques and methods to study functional characteristics of emulsion systems. *J Food Drug Anal* 2017;**25**:16–26.
123. Finney JL. Bernal's road to random packing and the structure of liquids. *Philos Mag* 2013;**93**:3940–69.
124. Madawi EA, Manaa HM, Alatrach DG, Al Mogharbel ZA, Hussain Z, Ahmed IS. Dry emulsions as a promising adaptation in pharmaceutical dosage formulations: a review of recent developments and biopharmaceutical significance. *J Drug Deliv Sci Technol* 2024;**96**:105712.
125. Medeleanu ML, Fărcaș AC, Coman C, Leopold L, Diaconeasa Z, Socaci SA. Citrus essential oils – based nano-emulsions: functional properties and potential applications. *Food Chem X* 2023;**20**:100960.
126. Pavoni L, Perinelli DR, Bonacucina G, Cespi M, Palmieri GF. An overview of micro- and nanoemulsions as vehicles for essential oils: formulation, preparation and stability. *Nanomaterials* 2020;**10**:135.
127. Madni A, Jameel F, Bajwa SZ, Rehman A, Khan WS. Toxicity, biological fate, and bioavailability of nanoemulsion formulations. In: Abd-El Salam KA, Murugan K, editors. *Bio-Based nanoemulsions for agri-food applications*. Amsterdam: Elsevier; 2022. p. 91–104.
128. Calvo P, Vila-Jato JL, Alonso MJ. Comparative *in vitro* evaluation of several colloidal systems, nanoparticles, nanocapsules, and nanoemulsions, as ocular drug carriers. *J Pharmacol Sci* 1996;**85**:530–6.
129. Mason TG. Emulsification and the emergence of nanoemulsions. *Matter* 2019;**1**:542–6.
130. Chagediya V. Nanoemulsions: a recent drug delivery tool. In: Mejuto J, Poša J, editors. *Design and applications of self-assembly aggregates - from micelles to nanoemulsions*. London: IntechOpen Limited; 2024.
131. da Silva Tinoco LM, da Silva FLO, Ferreira LAM, Leite EA, Carneiro G. Hyaluronic acid-coated nanoemulsions loaded with a hydrophobic ion pair of all-trans retinoic acid for improving the anticancer activity. *Braz J Pharm Sci* 2018;**54**:e17361.
132. Sánchez-López E, Guerra M, Dias-Ferreira J, Lopez-Machado A, Ettcheto M, Cano A, et al. Current applications of nanoemulsions in cancer therapeutics. *Nanomaterials* 2019;**9**:821.
133. Alghareeb S, Ekenna I, Asare-Addo K, Conway BR, Adebisi AO. Chitosan nanoparticles for nasal drug delivery. *J Drug Deliv Sci Technol* 2025;**105**:106623.
134. Diedrich C, Camargo Zittlau I, Schineider Machado C, Taise Fin M, Maissar Khalil N, Badea I, et al. Mucoadhesive nanoemulsion enhances brain bioavailability of luteolin after intranasal administration and induces apoptosis in SH-SY5Y neuroblastoma cells. *Int J Pharm* 2022;**626**:122142.
135. Wooster TJ, Moore SC, Chen W, Andrews H, Addepalli R, Seymour RB, et al. Biological fate of food nanoemulsions and the nutrients they carry – internalisation, transport and cytotoxicity of edible nanoemulsions in caco-2 intestinal cells. *RSC Adv* 2017;**7**:40053–66.
136. Naimi S, Viennois E, Gewirtz AT, Chassaing B. Direct impact of commonly used dietary emulsifiers on human gut microbiota. *Microbiome* 2021;**9**:66.
137. Pouton CW. Self-emulsifying drug delivery systems: assessment of the efficiency of emulsification. *Int J Pharm* 1985;**27**:335–48.
138. van Staden D, du Plessis J, Viljoen J. Development of a self-emulsifying drug delivery system for optimized topical delivery of clofazimine. *Pharmaceutics* 2020;**12**:523.
139. Uttreja P, Karnik I, Adel Ali Youssef A, Narala N, Elkanayati RM, Baisa S, et al. Self-emulsifying drug delivery systems (SEDDS): transition from liquid to solid—a comprehensive review of formulation, characterization, applications, and future trends. *Pharmaceutics* 2025;**17**:63.
140. Elbahwy IA, Lupo N, Ibrahim HM, Ismael HR, Kasem AA, Caliskan C, et al. Mucoadhesive self-emulsifying delivery systems for ocular administration of econazole. *Int J Pharm* 2018;**541**:72–80.
141. Chen YS, Chiu YH, Li YS, Lin EY, Hsieh DK, Lee CH, et al. Integration of PEG 400 into a self-nanoemulsifying drug delivery system improves drug loading capacity and nasal mucosa permeability and prolongs the survival of rats with malignant brain tumors. *Int J Nanomed* 2019;**14**:3601–13.
142. Köllner S, Nardin I, Markt R, Griesser J, Prüfert F, Bernkop-Schnürch A. Self-emulsifying drug delivery systems: design of a novel vaginal delivery system for curcumin. *Eur J Pharm Biopharm* 2017;**115**:268–75.
143. Friedl JD, Walther M, Vestweber PK, Wächter J, Knoll P, Jörgensen AM, et al. SEDDS-loaded mucoadhesive fiber patches for advanced oromucosal delivery of poorly soluble drugs. *J Control Release* 2022;**348**:692–705.
144. van Staden D, du Plessis J, Viljoen J. Development of topical/transdermal self-emulsifying drug delivery systems, not as simple as expected. *Sci Pharm* 2020;**88**:17.
145. Lam HT, Le NMN, Phan TNQ, Bernkop-Schnürch A. Mucolytic self-emulsifying drug delivery systems (SEDDS) containing a hydrophobic ion-pair of proteinase. *Eur J Pharmaceut Sci* 2021;**162**:105658.
146. Salimi E, Le-Vinh B, Zahir-Jouzani F, Matuszczak B, Ghasee A, Bernkop-Schnürch A. Self-emulsifying drug delivery systems changing their zeta potential via a flip-flop mechanism. *Int J Pharm* 2018;**550**:200–6.
147. Leonaviciute G, Adamovic NT, Lam HT, Rohrer J, Partenhauser A, Bernkop-Schnürch A. Self-emulsifying drug delivery systems (SEDDS): proof-of-concept how to make them mucoadhesive. *Eur J Pharm Biopharm* 2017;**112**:51–7.
148. Liu J, Li Y, Lun Z, Zhang Y, Yang P, Tang X, et al. Factors, mechanisms, and kinetics of spontaneous emulsification for heavy oil-in-water emulsions. *Molecules* 2024;**29**:2998.
149. Azodi M, Solaimany Nazar AR. An experimental study on factors affecting the heavy crude oil in water emulsions viscosity. *J Pet Sci Eng* 2013;**106**:1–8.
150. Bouchemal K, Briançon S, Perrier E, Fessi H. Nano-emulsion formulation using spontaneous emulsification: solvent, oil and surfactant optimisation. *Int J Pharm* 2004;**280**:241–51.
151. Subramaniam S, Elz A, Wignall A, Kamath S, Ariee A, Hunter A, et al. Self-emulsifying drug delivery systems (SEDDS) disrupt the gut microbiota and trigger an intestinal inflammatory response in rats. *Int J Pharm* 2023;**648**:123614.
152. Rohrer J, Zupančič O, Hetényi G, Kurpiers M, Bernkop-Schnürch A. Design and evaluation of SEDDS exhibiting high emulsifying properties. *J Drug Deliv Sci Technol* 2018;**44**:366–72.
153. Maji I, Mahajan S, Sriram A, Medtiya P, Vasave R, Khatri DK, et al. Solid self emulsifying drug delivery system: superior mode for oral delivery of hydrophobic cargos. *J Control Release* 2021;**337**:646–60.
154. Meirinho S, Rodrigues M, Santos AO, Falcão A, Alves G. Self-emulsifying drug delivery systems: an alternative approach to improve brain bioavailability of poorly water-soluble drugs through intranasal administration. *Pharmaceutics* 2022;**14**:1487.
155. Seifriz V. Studies in emulsions. I-II. *J Phys Chem* 1925;**29**:587–600.
156. Ding S, Serra CA, Vandamme TF, Yu W, Anton N. Double emulsions prepared by two-step emulsification: history, state-of-the-art and perspective. *J Control Release* 2019;**295**:31–49.
157. Saffarianpour S. One-step preparation of double emulsions stabilized with amphiphilic and stimuli-responsive block copolymers and nanoparticles for nutraceuticals and drug delivery. *JCI Open* 2021;**3**:100020.
158. Iqbal M, Zafar N, Fessi H, Elaissari A. Double emulsion solvent evaporation techniques used for drug encapsulation. *Int J Pharm* 2015;**496**:173–90.

159. Bako HK, Ibeogu HI, Bassey AP, Yar MS, Zhou T, Li C. Optimisation and characterization of double emulsion derived from rice starch, rice protein isolates and rice bran oil. *Int J Biol Macromol* 2024;**258**:128966.
160. Hu C, Wang Q, Ma C, Xia Q. Non-aqueous self-double-emulsifying drug delivery system: a new approach to enhance resveratrol solubility for effective transdermal delivery. *Colloids Surf A Physicochem Eng Asp* 2016;**489**:360–9.
161. Lin X, Wu X, Chen X, Wang B, Xu W. Intellectual and stimuli-responsive drug delivery systems in eyes. *Int J Pharm* 2021;**602**:120591.
162. Kuhn W, Hartgitay B, Katchalsky A, Eisenberg H. Reversible dilation and contraction by changing the state of ionization of high-polymer acid networks. *Nature* 1950;**165**:514–6.
163. García MC. Ionic-strength-responsive polymers for drug delivery applications. In: Makhlof A, Abu-Thabit N, editors. *Stimuli responsive polymeric nanocarriers for drug delivery applications*. Amsterdam: Elsevier; 2019. p. 393–409.
164. Gol E, Hudson S. Stimuli-responsive polymers and their bioconjugates. *Prog Polym Sci* 2004;**29**:1173–222.
165. Abuwatfa WH, Awad NS, Pitt WG, Hussein GA. Thermosensitive polymers and thermo-responsive liposomal drug delivery systems. *Polymers (Basel)* 2022;**14**:925.
166. Tavagnacco L, Zaccarelli E, Chiessi E. Molecular description of the coil-to-globule transition of poly(N-isopropylacrylamide) in water/ethanol mixture at low alcohol concentration. *J Mol Liq* 2020;**297**:111928.
167. Lin S-Y, Chen K-S, Liang R-C. Thermal micro ATR/FT-IR spectroscopic system for quantitative study of the molecular structure of poly(N-isopropylacrylamide) in water. *Polymer (Guildf)* 1999;**40**:2619–24.
168. Volpert E, Selb J, Candau F. Associating behaviour of polyacrylamides hydrophobically modified with dihexylacrylamide. *Polymer (Guildf)* 1998;**39**:1025–33.
169. Madu SJ, Hassan D, Igbokwe N, Orugun OA, Muazu J. Temperature-sensitive polymers for biomaterials for drug delivery, gene delivery, and tissue engineering. In: Varaprasad K, editor. *Polymeric biomaterials for healthcare applications*. Amsterdam: Elsevier; 2022. p. 335–67.
170. Agrawal M, Saraf S, Saraf S, Dubey SK, Puri A, Gupta U, et al. Stimuli-responsive *in situ* gelling system for nose-to-brain drug delivery. *J Control Release* 2020;**327**:235–65.
171. Thapa R, Gurung S, Parat M-O, Parekh HS, Pandey P. Application of sol-gels for treatment of gynaecological conditions—physiological perspectives and emerging concepts in intravaginal drug delivery. *Gels* 2022;**8**:99.
172. Norton IT, Jarvis DA, Foster TJ. A molecular model for the formation and properties of fluid gels. *Int J Biol Macromol* 1999;**26**:255–61.
173. Gabriele A, Spyropoulos F, Norton IT. Kinetic study of fluid gel formation and viscoelastic response with kappa-carrageenan. *Food Hydrocoll* 2009;**23**:2054–61.
174. Pandey M, Choudhury H, Abdul-Aziz A, Bhattamisra SK, Gorain B, Carine T, et al. Promising drug delivery approaches to treat microbial infections in the vagina: a recent update. *Polymers (Basel)* 2020;**13**:26.
175. Mohd Shahrizan MS, Abd Aziz ZH, Katas H. Fluid gels: a systematic review towards their application in pharmaceutical dosage forms and drug delivery systems. *J Drug Deliv Sci Technol* 2022;**67**:102947.
176. M Grover L, Moakes R, Rauz S. Innovations in fluid-gel eye drops for treating disease of the eye: prospects for enhancing drug retention and reducing corneal scarring. *Expet Rev Ophthalmol* 2022;**17**:175–81.
177. Garrec DA, Frasc-Melnik S, Henry JVL, Spyropoulos F, Norton IT. Designing colloidal structures for micro and macro nutrient content and release in foods. *Faraday Discuss* 2012;**158**:37.
178. Lazidis A, Hancocks RD, Spyropoulos F, Kreuß M, Berrocal R, Norton IT. Whey protein fluid gels for the stabilisation of foams. *Food Hydrocoll* 2016;**53**:209–17.
179. Walden P. Über die Molekulargröße und elektrische Leitfähigkeit einiger geschmolzener Salze. *Bull Acad Imper Sci (St Petersburg)* 1914;**8**:405–22.
180. Lei Z, Chen B, Koo YM, MacFarlane DR. Introduction: ionic liquids. *Chem Rev* 2017;**117**:6633–5.
181. Kuddushi M, Xu B Bin, Malek N, Zhang X. Review of ionic liquid and ionogel-based biomaterials for advanced drug delivery. *Adv Colloid Interface Sci* 2024;**331**:103244.
182. Llaver M, Fiorentini EF, Quintas PY, Oviedo MN, Botella Arenas MB, Wuilloud RG. Task-specific ionic liquids: applications in sample preparation and the chemistry behind their selectivity. *Adv Sample Prep* 2022;**1**:100004.
183. Huang W, Wu X, Qi J, Zhu Q, Wu W, Lu Y, et al. Ionic liquids: green and tailor-made solvents in drug delivery. *Drug Discov Today* 2020;**25**:901–8.
184. Peng K, Gao Y, Angsantikul P, LaBarbiera A, Goetz M, Curreri AM, et al. Modulation of gastrointestinal mucus properties with ionic liquids for drug delivery. *Adv Healthcare Mater* 2021;**10**:2002192.
185. Shukla MK, Tiwari H, Verma R, Dong W-L, Azizov S, Kumar B, et al. Role and recent advancements of ionic liquids in drug delivery systems. *Pharmaceutics* 2023;**15**:702.
186. Wu X, Zhu Q, Chen Z, Wu W, Lu Y, Qi J. Ionic liquids as a useful tool for tailoring active pharmaceutical ingredients. *J Control Release* 2021;**338**:268–83.
187. Yan J, Mangolini F. Engineering encapsulated ionic liquids for next-generation applications. *RSC Adv* 2021;**11**:36273–88.
188. Kuroda K. A simple overview of toxicity of ionic liquids and designs of biocompatible ionic liquids. *New J Chem* 2022;**46**:20047–52.
189. Jing B, Lan N, Qiu J, Zhu Y. Interaction of ionic liquids with a lipid bilayer: a biophysical study of ionic liquid cytotoxicity. *J Phys Chem B* 2016;**120**:2781–9.
190. Benedetto A, Heinrich F, Gonzalez MA, Fragneto G, Watkins E, Ballone P. Structure and stability of phospholipid bilayers hydrated by a room-temperature ionic liquid/water solution: a neutron reflectometry study. *J Phys Chem B* 2014;**118**:12192–206.
191. Kuroda K, Satria H, Miyamura K, Tsuge Y, Ninomiya K, Takahashi K. Design of wall-destructive but membrane-compatible solvents. *J Am Chem Soc* 2017;**139**:16052–5.
192. Reinitzer F. Liquid crystals. *Monatsh Chem* 1888:421–41.
193. Mitov M. Liquid-crystal science from 1888 to 1922: building a revolution. *ChemPhysChem* 2014;**15**:1245–50.
194. Chen J. Current advances in anisotropic structures for enhanced osteogenesis. *Colloids Surf B Biointerfaces* 2023;**231**:113566.
195. Nalone L, Marques C, Costa S, Souto EB, Severino P. Liquid crystalline drug delivery systems. In: Shegokar R, editor. *Drug delivery trends*. Amsterdam: Elsevier; 2020. p. 141–9.
196. Chavda VP, Dawre S, Pandya A, Vora LK, Modh DH, Shah V, et al. Lyotropic liquid crystals for parenteral drug delivery. *J Control Release* 2022;**349**:533–49.
197. Huang Y, Gui S. Factors affecting the structure of lyotropic liquid crystals and the correlation between structure and drug diffusion. *RSC Adv* 2018;**8**:6978–87.
198. Tadros T. Liquid crystalline phase. In: Tadros T, editor. *Encyclopedia of Colloid and interface science*. Berlin: Springer Berlin Heidelberg; 2013. p. 682–3.
199. Zhang W, Liu L. Study on the formation and properties of liquid crystal emulsion in cosmetic. *J Cosmet Dermatol Sci Appl* 2013;**3**:139–44.
200. Yaghmur A, Kriechbaum M, Amenitsch H, Steinhart M, Laggner P, Rappolt M. Effects of pressure and temperature on the self-assembled fully hydrated nanostructures of monoolein–oil systems. *Langmuir* 2010;**26**:1177–85.
201. Ristroph K, Salim M, Wilson BK, Clulow AJ, Boyd BJ, Prud'homme RK. Internal liquid crystal structures in nanocarriers containing drug hydrophobic ion pairs dictate drug release. *J Colloid Interface Sci* 2021;**582**:815–24.
202. Guthrie F. On eutexia. *Proc Phys Soc Lond* 1884;(6):124.
203. Rahman MS, Roy R, Jadhav B, Hossain MN, Halim MA, Raynie DE. Formulation, structure, and applications of therapeutic and amino

- acid-based deep eutectic solvents: an overview. *J Mol Liq* 2021;**321**: 114745.
204. Chakraborty S, Chormale JH, Bansal AK. Deep eutectic systems: an overview of fundamental aspects, current understanding and drug delivery applications. *Int J Pharm* 2021;**610**:121203.
 205. Jisha KJ, Athira KK, Priyanka VP, Gardas RL. Liquid-liquid extraction. In: Verma C, Verma DK, editors. *Handbook of bio-molecules*. Amsterdam: Elsevier; 2023. p. 227–39.
 206. Alhadid A, Mokrushina L, Minceva M. Design of deep eutectic systems: a simple approach for preselecting eutectic mixture constituents. *Molecules* 2020;**25**:1077.
 207. Ehrenström Reiz GM, Reiz SL. EMLA – a eutectic mixture of local anaesthetics for topical anaesthesia. *Acta Anaesthesiol Scand* 1982; **26**:596–8.
 208. Chu KA, Yalkowsky SH. An interesting relationship between drug absorption and melting point. *Int J Pharm* 2009;**373**:24–40.
 209. Gan Y, Baak JPA, Chen T, Ye H, Liao W, Lv H, et al. Supersaturation and precipitation applicated in drug delivery systems: development strategies and evaluation approaches. *Molecules* 2023;**28**:2212.
 210. Chen Y, Mu T. Revisiting greenness of ionic liquids and deep eutectic solvents. *Green Chem Eng* 2021;**2**:174–86.
 211. Abdelquader MM, Li S, Andrews GP, Jones DS. Therapeutic deep eutectic solvents: a comprehensive review of their thermodynamics, microstructure and drug delivery applications. *Eur J Pharm Biopharm* 2023;**186**:85–104.
 212. Bo R, Wu J, Tao Y, Hong H, Peng W, Wang W, et al. Characterization of chitosan-coated PLGA nanoemulsion loaded with cepharanthine and inhibitory effect on *Staphylococcus aureus* pneumonia of mice. *Int J Pharm* 2025;**673**:125396.
 213. Kang JH, Yang MS, Kwon TK, Kim DW, Park CW. Inhaled deep eutectic solvent based-nanoemulsion of pirfenidone in idiopathic pulmonary fibrosis. *J Control Release* 2022;**352**:570–85.
 214. Najafabadi AP, Pourmadadi M, Yazdian F, Rashedi H, Rahdar A, Díez-Pascual AM. pH-sensitive ameliorated quercetin delivery using graphene oxide nanocarriers coated with potential anticancer gelatin-polyvinylpyrrolidone nanoemulsion with bitter almond oil. *J Drug Deliv Sci Technol* 2023;**82**:104339.
 215. Lin X, Sheng Y, Zhang X, Li Z, Yang Y, Wu J, et al. Oil-in-ionic liquid nanoemulsion-based intranasal delivery system for influenza split-virus vaccine. *J Control Release* 2022;**346**:380–91.
 216. Jumelle C, Gholizadeh S, Annabi N, Dana R. Advances and limitations of drug delivery systems formulated as eye drops. *J Control Release* 2020;**321**:1–22.
 217. Li S, Chen L, Fu Y. Nanotechnology-based ocular drug delivery systems: recent advances and future prospects. *J Nanobiotechnol* 2023;**21**:232.
 218. Forrester JV, Dick AD, McMenamin PG, Roberts F, Pearlman E. General and ocular pharmacology. In: Forrester JV, editor. *The eye*. Philadelphia: Elsevier; 2016. p. 338–69.
 219. El-Gendy MA, Mansour M, El-Assal MIA, Ishak RAH, Mortada ND. Delineating penetration enhancer-enriched liquid crystalline nanostructures as novel platforms for improved ophthalmic delivery. *Int J Pharm* 2020;**582**:119313.
 220. Akhter MH, Ahmad I, Alshahrani MY, Al-Harbi AI, Khalilullah H, Afzal O, et al. Drug delivery challenges and current progress in nanocarrier-based ocular therapeutic system. *Gels* 2022;**8**:82.
 221. Macwan JS, Hirani A, Pathak Y. Challenges in ocular pharmacokinetics and drug delivery. In: Pathak Y, Sutariya V, Hirani AA, editors. *Nano-Biomaterials for ophthalmic drug delivery*. Cham: Springer International Publishing; 2016. p. 593–611.
 222. Laffleur F, Bauer B. Progress in nasal drug delivery systems. *Int J Pharm* 2021;**607**:120994.
 223. Tai J, Han M, Lee D, Park IH, Lee SH, Kim TH. Different methods and formulations of drugs and vaccines for nasal administration. *Pharmaceutics* 2022;**14**:1073.
 224. Zia H, Dondeti P, Needham TE. Intranasal drug delivery. *Clin Res Regul Aff* 1993;**10**:99–135.
 225. Du L, Chen L, Liu F, Wang W, Huang H. Nose-to-brain drug delivery for the treatment of CNS disease: new development and strategies. *Int Rev Neurobiol* 2023;**171**:255–97.
 226. Williams G, Suman JD. *In vitro* anatomical models for nasal drug delivery. *Pharmaceutics* 2022;**14**:1353.
 227. Tiozzo Fasiolo L, Manniello MD, Tratta E, Buttini F, Rossi A, Sonvico F, et al. Opportunity and challenges of nasal powders: drug formulation and delivery. *Eur J Pharmaceut Sci* 2018;**113**:2–17.
 228. Rawas-Qalaji M, Thu HE, Hussain Z. Oromucosal delivery of macromolecules: challenges and recent developments to improve bioavailability. *J Control Release* 2022;**352**:726–46.
 229. Nayak BS, Sourajit S, Palo M, Behera S. Sublingual drug delivery system: a novel approach. *Int J Pharm Drug Anal* 2017;**5**:399–405.
 230. Brandl M, Bauer-Brandl A. Oromucosal drug delivery: trends in *in-vitro* biopharmaceutical assessment of new chemical entities and formulations. *Eur J Pharmaceut Sci* 2019;**128**:112–7.
 231. Bastos F, Pinto AC, Nunes A, Simões S. Oromucosal products – market landscape and innovative technologies: a review. *J Control Release* 2022;**348**:305–20.
 232. Esih H, Mezgec K, Billmeier M, Malenšek Š, Benčina M, Grilc B, et al. Mucoadhesive film for oral delivery of vaccines for protection of the respiratory tract. *J Control Release* 2024;**371**:179–92.
 233. Mazzinelli E, Favuzzi I, Arcovito A, Castagnola R, Fratoocchi G, Mordente A, et al. Oral mucosa models to evaluate drug permeability. *Pharmaceutics* 2023;**15**:1559.
 234. Pandey M, Choudhury H, Ying JNS, Ling JFS, Ting J, Ting JSS, et al. Mucoadhesive nanocarriers as a promising strategy to enhance intracellular delivery against oral cavity carcinoma. *Pharmaceutics* 2022;**14**:795.
 235. Silva BMA, Borges AF, Silva C, Coelho JFJ, Simões S. Mucoadhesive oral films: the potential for unmet needs. *Int J Pharm* 2015; **494**:537–51.
 236. Bredenberg S, Duberg M, Lennernäs B, Lennernäs H, Pettersson A, Westerberg M, et al. *In vitro* and *in vivo* evaluation of a new sublingual tablet system for rapid oromucosal absorption using fentanyl citrate as the active substance. *Eur J Pharmaceut Sci* 2003;**20**: 327–34.
 237. Guo Y, Pratap Singh A. Emerging strategies for enhancing buccal and sublingual administration of nutraceuticals and pharmaceuticals. *J Drug Deliv Sci Technol* 2019;**52**:440–51.
 238. Valamla B, Thakor P, Phuse R, Dalvi M, Kharat P, Kumar A, et al. Engineering drug delivery systems to overcome the vaginal mucosal barrier: current understanding and research agenda of mucoadhesive formulations of vaginal delivery. *J Drug Deliv Sci Technol* 2022;**70**: 103162.
 239. Yang Z, Wu X, Wang H, Zhou J, Lin X, Yang P. Vagina, a promising route for drug delivery. *J Drug Deliv Sci Technol* 2024;**93**:105397.
 240. Elstone NS, Shimizu K, Shaw EV, Lane PD, D'Andrea L, Demé B, et al. Understanding the liquid structure in mixtures of ionic liquids with semiperfluoroalkyl or alkyl chains. *J Phys Chem B* 2023;**127**: 7394–407.
 241. Kakati N, Paul N, Dubey S, Mahanta J, Lakshmi AR, Banerjee T, et al. Microrheology of ionic liquid doped mucus for an efficient delivery of protein-based oral drugs. *Small* 2025;**21**:2500403.
 242. Eades W, Abdolmohammadpourbonab S, Dinh L, Yan B. Ionic liquids and their potential use in development and improvement of drug delivery systems: evidence of their tendency to promote drug accumulation in the brain. *Pharmaceut Dev Technol* 2024;**29**:1065–74.
 243. Donthi MR, Munnangi SR, Krishna KV, Saha RN, Singhvi G, Dubey SK. Nanoemulgel: a novel nano carrier as a tool for topical drug delivery. *Pharmaceutics* 2023;**15**:164.
 244. Pereira MM, Kurnia KA, Sousa FL, Silva NJO, Lopes-da-Silva JA, Coutinho JAP, et al. Contact angles and wettability of ionic liquids on polar and non-polar surfaces. *Phys Chem Chem Phys* 2015;**17**: 31653–61.
 245. Colombo M, Figueiró F, de Fraga Dias A, Teixeira HF, Battastini AMO, Koester LS. Kaempferol-loaded mucoadhesive

- nanoemulsion for intranasal administration reduces glioma growth *in vitro*. *Int J Pharm* 2018;**543**:214–23.
246. Zhang J-Y, Liu X-X, Lin J-Y, Bao X-Y, Peng J-Q, Gong Z-P, et al. Biomimetic engineered nanocarriers inspired by viruses for oral-drug delivery. *Int J Pharm* 2022;**624**:121979.
 247. Sato H, Yamada K, Miyake M, Onoue S. Recent advancements in the development of nanocarriers for mucosal drug delivery systems to control oral absorption. *Pharmaceutics* 2023;**15**:2708.
 248. Preeti R, Sambhakar S, Malik R, Bhatia S, Al Harrasi A, Rani C, et al. Nanoemulsion: an emerging novel technology for improving the bioavailability of drugs. *Scientifica (Cairo)* 2023;**2023**:6640103.
 249. Sipos B, Csóka I, Szivacsik N, Budai-Szűcs M, Schelcz Z, Zupkó I, et al. Mucoadhesive meloxicam-loaded nanoemulsions: development, characterization and nasal applicability studies. *Eur J Pharmaceut Sci* 2022;**175**:106229.
 250. Jaiswal M, Dudhe R, Sharma PK. Nanoemulsion: an advanced mode of drug delivery system. *3 Biotech* 2015;**5**:123–7.
 251. Wilson RJ, Li Y, Yang G, Zhao C-X. Nanoemulsions for drug delivery. *Particuology* 2022;**64**:85–97.
 252. Tran VN, Strnad O, Šuman J, Veverková T, Sukupová A, Cejnar P, et al. Cannabidiol nanoemulsion for eye treatment – anti-inflammatory, wound healing activity and its bioavailability using *in vitro* human corneal substitute. *Int J Pharm* 2023;**643**:123202.
 253. Bonferoni MC, Rossi S, Sandri G, Ferrari F, Gavini E, Rassu G, et al. Nanoemulsions for “nose-to-brain” drug delivery. *Pharmaceutics* 2019;**11**:84.
 254. Nazir I, Fürst A, Lupo N, Hupfau A, Gust R, Bernkop-Schnürch A. Zeta potential changing self-emulsifying drug delivery systems: a promising strategy to sequentially overcome mucus and epithelial barrier. *Eur J Pharm Biopharm* 2019;**144**:40–9.
 255. Lupo N, Tkadlečková VN, Jelkmann M, Laffleur F, Hetényi G, Kubová K, et al. Self-emulsifying drug delivery systems: *in vivo* evaluation of their potential for oral vaccination. *Acta Biomater* 2019;**94**:425–34.
 256. Krzemińska K, Sznitowska M. Development of self-emulsifying oils for ophthalmic delivery of antibiotics instable in water. *J Drug Deliv Sci Technol* 2023;**84**:104423.
 257. Qi X, Wang L, Zhu J, Hu Z, Zhang J. Self-double-emulsifying drug delivery system (SDEDDS): a new way for oral delivery of drugs with high solubility and low permeability. *Int J Pharm* 2011;**409**:245–51.
 258. Sakr MG, El-Zahaby SA, Al-Mahallawi AM, Ghorab DM. A novel reverse micelle based cationic double nanoemulsion as a potential nanoplatform for enhancing the anitglucomal activity of betaxolol hydrochloride; formulation, *in vitro* characterization, *ex vivo* permeation and *in vivo* pharmacodynamic evaluation in glaucomatous rabbits’ eyes. *J Drug Deliv Sci Technol* 2023;**90**:105112.
 259. Raghunandan S, Kumar RS, Kamaraj M, Gandhi AS. Role of water in the sol-gel synthesis of yttrium monosilicate. *Ceram Int* 2019;**45**:4487–92.
 260. Corazza E, di Cagno MP, Bauer-Brandl A, Abruzzo A, Cerchiara T, Bigucci F, et al. Drug delivery to the brain: *in situ* gelling formulation enhances carbamazepine diffusion through nasal mucosa models with mucin. *Eur J Pharmaceut Sci* 2022;**179**:106294.
 261. Alkholief M, Kalam MA, Almomen A, Alshememry A, Alshamsan A. Thermoresponsive sol-gel improves ocular bioavailability of dipivefrin hydrochloride and potentially reduces the elevated intraocular pressure *in vivo*. *Saudi Pharm J* 2020;**28**:1019–29.
 262. Dighe S, Lonkar S, Jog S, Mangrulkar SV, Sawarkar SP. Thermo-sensitive *in-situ* gel of nifedipine: a strategy to offset cognition impairment in Alzheimer’s disease. *J Drug Deliv Sci Technol* 2024;**102**:106393.
 263. Bender L, Boostrom HM, Varricchio C, Zuanon M, Celiksoy V, Sloan A, et al. A novel dual action monolithic thermosetting hydrogel loaded with lidocaine and metronidazole as a potential treatment for alveolar osteitis. *Eur J Pharm Biopharm* 2020;**149**:85–94.
 264. Zhang Y, Jiang Z, Lu K, Ding B, Wang J, Wang N, et al. *In situ* gel-forming oil solubilizing α -lipoic acid as a physical shielding alleviated chemotherapy-induced oral mucositis *via* inhibiting oxidative stress. *Int J Pharm* 2024;**665**:124714.
 265. Hsin YK, Thangarajoo T, Choudhury H, Pandey M, Meng LW, Gorain B. Stimuli-Responsive *in situ* spray gel of miconazole nitrate for vaginal candidiasis. *J Pharmacol Sci* 2023;**112**:562–72.
 266. El-Kamel A, Ashour A, Shebl N, Heikal L. Ionic liquids: an innovative approach in drug delivery. *Matter* 2024;**7**:2660–2.
 267. Peng K, Shi Y, LaBarbiera A, Mitragotri S. Mucoadhesive ionic liquid gel patches for oral delivery. *ACS Biomater Sci Eng* 2023;**9**:2838–45.
 268. Tanigawa H, Suzuki N, Suzuki T. Application of ionic liquid to enhance the nose-to-brain delivery of etodolac. *Eur J Pharmaceut Sci* 2022;**178**:106290.
 269. Vaidya A, Mitragotri S. Ionic liquid-mediated delivery of insulin to buccal mucosa. *J Control Release* 2020;**327**:26–34.
 270. Ng LH, Hadinoto K. Buccal delivery system of active pharmaceutical ingredients-ionic liquid (API-IL): effects of API-IL loading and gelatin film concentration. *Chem Eng Res Des* 2024;**202**:115–25.
 271. Medeiros L, dos Santos RF, da Rolt Nervis B, Jacobi M, Hashizume LN, Gazzi RP, et al. Synthesis of films based on chitosan and protic ionic liquids to be used as wound dressing on the oral mucosa. *Int J Biol Macromol* 2023;**253**:127134.
 272. Goodby JW. Microscopy applications | Liquid crystals. In: Worsfold PJ, Townshend A, Poole CF, editors. *Encyclopedia of analytical science*. Amsterdam: Elsevier; 2005. p. 74–84.
 273. Greene JP. *Automotive plastics and composites*. 1st ed. Amsterdam: Elsevier; 2021.
 274. Ivanova R, Lindman B, Alexandridis P. Modification of the lyotropic liquid crystalline microstructure of amphiphilic block copolymers in the presence of cosolvents. *Adv Colloid Interface Sci* 2001;**89–90**:351–82.
 275. Blanco-Fernández G, Blanco-Fernandez B, Fernández-Ferreiro A, Otero-Espinar FJ. Lipidic lyotropic liquid crystals: insights on biomedical applications. *Adv Colloid Interface Sci* 2023;**313**:102867.
 276. Calixto GMF, Vettorelli FD, Dovigo LN, Chorilli M. Polyethyleneimine and chitosan polymer-based mucoadhesive liquid crystalline systems intended for buccal drug delivery. *AAPS PharmSciTech* 2018;**19**:820–36.
 277. Jere D, Jiang HL, Kim YK, Arote R, Choi YJ, Yun CH, et al. Chitosan-graft-polyethyleneimine for Akt1 siRNA delivery to lung cancer cells. *Int J Pharm* 2009;**378**:194–200.
 278. Nezhad NM, Kamali H, Sarchahi AA, Jalalifar S, Amani A, Oroojalian F. Investigating the efficacy of liquid crystal nano cubosomes containing dorzolamide and timolol for drug delivery to the cornea. *J Drug Deliv Sci Technol* 2025;**106**:106724.
 279. See GL, Arce F, Dahlizar S, Okada A, Fadli MFBM, Hijikuro I, et al. Enhanced nose-to-brain delivery of tranilast using liquid crystal formulations. *J Control Release* 2020;**325**:1–9.
 280. Marena GD, Fonseca-Santos B, Matheus Aparecido dos Santos R, dos Santos KC, Bauab TM, Chorilli M. Incorporation of ursolic acid in liquid crystalline systems improves the antifungal activity against *Candida* sp. *J Pharm Innov* 2021;**16**:576–86.
 281. Borgheti-Cardoso LN, Depieri LV, Kooijmans SAA, Diniz H, Calzzani RAJ, Vicentini FTM de Carvalho, et al. An *in situ* gelling liquid crystalline system based on monoglycerides and polyethyleneimine for local delivery of siRNAs. *Eur J Pharmaceut Sci* 2015;**74**:103–17.
 282. Jie H, Liu L, Shuangying G, Xingqi W, Rongfeng H, Yong Z, et al. A novel phytantriol-based *in situ* liquid crystal gel for vaginal delivery. *AAPS PharmSciTech* 2019;**20**:185.
 283. Wolbert F, Brandenbusch C, Sadowski G. Selecting excipients forming therapeutic deep eutectic systems—a mechanistic approach. *Mol Pharm* 2019;**16**:3091–9.
 284. Ribeiro BD, Florindo C, Iff LC, Coelho MAZ, Marrucho IM. Menthol-based eutectic mixtures: hydrophobic low viscosity solvents. *ACS Sustainable Chem Eng* 2015;**3**:2469–77.

285. Stolarska O, Soto A, Rodríguez H, Smiglak M. Properties modification by eutectic formation in mixtures of ionic liquids. *RSC Adv* 2015;**5**:22178–87.
286. Bazzo GC, Pezzini BR, Stulzer HK. Eutectic mixtures as an approach to enhance solubility, dissolution rate and oral bioavailability of poorly water-soluble drugs. *Int J Pharm* 2020;**588**:119741.
287. de Paiva CS, St Leger AJ, Caspi RR. Mucosal immunology of the ocular surface. *Mucosal Immunol* 2022;**15**:1143–57.
288. Arribada RG, Rodrigues-Braz D, Silva-Cunha A, Behar-Cohen F. Excipients in drug delivery systems: a comprehensive review of approved inactive ingredients for human ophthalmic formulations. *Eur J Pharm Biopharm* 2025;**208**:114637.
289. Cholkar K, Dasari SR, Pal D, Mitra AK. Eye: anatomy, physiology and barriers to drug delivery. In: Mitra AK, editor. *Ocular transporters and receptors*. Swastan: Woodhead Publishing; 2013. p. 1–36.
290. Suri R, Beg S, Kohli K. Target strategies for drug delivery bypassing ocular barriers. *J Drug Deliv Sci Technol* 2020;**55**:101389.
291. Patel PB, Shastri DH, Shelat PK, Shukla AK. Ophthalmic drug delivery system: challenges and approaches. *Sys Rev Pharm* 2010;**1**: 113–20.
292. Elsayed I, Sayed S. Tailored nanostructured platforms for boosting transcorneal permeation: Box-Behnken statistical optimization, comprehensive *in vitro*, *ex vivo* and *in vivo* characterization. *Int J Nanomed* 2017;**12**:7947–62.
293. Onugwu AL, Nwagwu CS, Onugwu OS, Echezona AC, Agbo CP, Ihim SA, et al. Nanotechnology based drug delivery systems for the treatment of anterior segment eye diseases. *J Control Release* 2023;**354**:465–88.
294. Meek KM, Knupp C. Corneal structure and transparency. *Prog Retin Eye Res* 2015;**49**:1–16.
295. Maulvi FA, Soni TG, Shah DO. A review on therapeutic contact lenses for ocular drug delivery. *Drug Deliv* 2016;**23**:3017–26.
296. Shafiq M, Rafique M, Cui Y, Pan L, Do CW, Ho EA. An insight on ophthalmic drug delivery systems: focus on polymeric biomaterials-based carriers. *J Control Release* 2023;**362**:446–67.
297. Auel T, Mentrup AFC, Oldfield LR, Seidlitz A. 3D printing of pharmaceutical dosage forms: recent advances and applications. *Adv Drug Deliv Rev* 2025;**217**:115504.
298. Cassano R, Di Gioia ML, Trombino S. Gel-based materials for ophthalmic drug delivery. *Gels* 2021;**7**:130.
299. Salawi A. Self-emulsifying drug delivery systems: a novel approach to deliver drugs. *Drug Deliv* 2022;**29**:1811–23.
300. Nezhad NM, Rahimi M, Gheybi F, Kesharwani P, Oroojalian F, Sahebkar A. Navigating the evolution of ophthalmic drug delivery and ocular regenerative medicine from conventional to cutting-edge treatments. *Appl Mater Today* 2025;**42**:102602.
301. Agrawal AK, Das M, Jain S. *In situ* gel systems as 'smart' carriers for sustained ocular drug delivery. *Expet Opin Drug Deliv* 2012;**9**: 383–402.
302. Geethalakshmi A, Karki R, Sagi P, Jha SK, Venkatesh DP. Temperature triggered *in situ* gelling system for betaxolol in glaucoma. *J Appl Pharmaceut Sci* 2013;**3**:153–9.
303. Al-Kinani AA, Zidan G, Elsaid N, Seyfoddin A, Alani AWG, Alany RG. Ophthalmic gels: past, present and future. *Adv Drug Deliv Rev* 2018;**126**:113–26.
304. Karmakar S, Manna S, Kabiraj S, Jana S. Recent progress in alginate-based carriers for ocular targeting of therapeutics. *Food Hydrocoll Health* 2022;**2**:100071.
305. Grant GT, Morris ER, Rees DA, Smith PJC, Thom D. Biological interactions between polysaccharides and divalent cations: the egg-box model. *FEBS Lett* 1973;**32**:195–8.
306. Grasdalen H, Smidsrød O. Gelation of gellan gum. *Carbohydr Polym* 1987;**7**:371–93.
307. Chandrasekaran R, Puigjaner LC, Joyce KL, Arnott S. Cation interactions in gellan: an x-ray study of the potassium salt. *Carbohydr Res* 1988;**181**:23–40.
308. Chandrasekaran R, Thailambal VG. The influence of calcium ions, acetate and l-glycerate groups on the gellan double-helix. *Carbohydr Polym* 1990;**12**:431–42.
309. Sarmiento C, Duarte ARC, Rita Jesus A. Can (natural) deep eutectic systems increase the efficacy of ocular therapeutics? *Eur J Pharm Biopharm* 2024;**198**:114276.
310. El Achkar T, Moufawad T, Ruellan S, Landy D, Greige-Gerges H, Fourmentin S. Cyclodextrins: from solute to solvent. *ChemComm* 2020;**56**:3385–8.
311. Türker S, Onur E, Özer Y. Nasal route and drug delivery systems. *Pharm World Sci* 2004;**26**:137–42.
312. Ignaczak M. Nasal delivery a galaxy not so far away. a brief market history of the nasal delivery landscape. (industry projections of nasal products) *Drug Deliv*. Published 23 November 2022. p. 6–8. Accessed 20 November 2025. Available form: <https://www.ondrugdelivery.com/nasal-delivery-a-galaxy-not-so-far-away-a-brief-market-history-of-the-nasal-delivery-landscape/>.
313. Chung S, Peters JM, Detynecki K, Tatum W, Rabinowicz AL, Carrazana E. The nose has it: opportunities and challenges for intranasal drug administration for neurologic conditions including seizure clusters. *Epilepsy Behav Rep* 2023;**21**:100581.
314. Costantino HR, Illum L, Brandt G, Johnson PH, Quay SC. Intranasal delivery: physicochemical and therapeutic aspects. *Int J Pharm* 2007;**337**:1–24.
315. Raghav M, Gupta V, Awasthi R, Singh A, Kulkarni GT. Nose-to-brain drug delivery: challenges and progress towards brain targeting in the treatment of neurological disorders. *J Drug Deliv Sci Technol* 2023;**86**:104756.
316. Inoue D, Furubayashi T, Tanaka A, Sakane T, Sugano K. Effect of cerebrospinal fluid circulation on nose-to-brain direct delivery and distribution of caffeine in rats. *Mol Pharm* 2020;**17**:4067–76.
317. Jeong S-H, Jang J-H, Lee Y-B. Drug delivery to the brain via the nasal route of administration: exploration of key targets and major consideration factors. *J Pharm Investig* 2023;**53**:119–52.
318. Cho HJ, Ku WS, Termsarasab U, Yoon I, Chung CW, Moon HT, et al. Development of udenafil-loaded microemulsions for intranasal delivery: *in vitro* and *in vivo* evaluations. *Int J Pharm* 2012;**423**: 153–60.
319. Fortuna A, Alves G, Serralheiro A, Sousa J, Falcão A. Intranasal delivery of systemic-acting drugs: small-molecules and biomacromolecules. *Eur J Pharm Biopharm* 2014;**88**:8–27.
320. Lobaina Mato Y. Nasal route for vaccine and drug delivery: features and current opportunities. *Int J Pharm* 2019;**572**:118813.
321. Keller LA, Merkel O, Popp A. Intranasal drug delivery: opportunities and toxicologic challenges during drug development. *Drug Deliv Transl Res* 2022;**12**:735–57.
322. de Barros C, Portugal I, Batain F, Portella D, Severino P, Cardoso J, et al. Formulation, design and strategies for efficient nanotechnology-based nasal delivery systems. *RPSPPR* 2022;**1**:rqac003.
323. Yang D, Liu C, Piao H, Quan P, Fang L. Enhanced drug loading in the drug-in-adhesive transdermal patch utilizing a drug-ionic liquid strategy: insight into the role of ionic hydrogen bonding. *Mol Pharm* 2021;**18**:1157–66.
324. Wu X, Zhang H, He S, Yu Q, Lu Y, Wu W, et al. Improving dermal delivery of hyaluronic acid by ionic liquids for attenuating skin dehydration. *Int J Biol Macromol* 2020;**150**:528–35.
325. Dong Z, Zhang L, Li G, Li Y, He H, Lu Y, et al. Mechanism and performance of choline-based ionic liquids in enhancing nasal delivery of glucagon. *J Control Release* 2024;**375**:812–28.
326. Upadhyay R, Ghosh P, Desavathu M. Advancement in the nose-to-brain drug delivery of FDA-approved drugs for the better management of depression and psychiatric disorders. *Int J Pharm* 2024;**667**: 124866.
327. Chatterjee B, Gorain B, Mohananaidu K, Sengupta P, Mandal UK, Choudhury H. Targeted drug delivery to the brain via intranasal nanoemulsion: available proof of concept and existing challenges. *Int J Pharm* 2019;**565**:258–68.

328. Duarte JL, Di Filippo LD, Azevedo Vilella KJ, Paes Dutra JA, Ribeiro DM, Freitas da Silva M, et al. Chitosan-coated nanoemulsion for intranasal administration increases temozolomide mucosal permeation, cellular uptake, and *in vitro* cytotoxicity in glioblastoma multiforme cells. *J Drug Deliv Sci Technol* 2024;**102**:106390.
329. Salama A, Soliman GM, Elsherbiny N, Safwat MA. Chitosan-coated nanoemulsion for the direct nose-to-brain delivery of sildenafil: development and *in vivo* evaluation in a brain oxidative stress and inflammation model. *J Drug Deliv Sci Technol* 2024;**98**:105842.
330. Otaegui L, Urgin T, Zaiter T, Zussy C, Vitalis M, Pellequer Y, et al. Nose-to-brain delivery of DHA-loaded nanoemulsions: a promising approach against Alzheimer's disease. *Int J Pharm* 2025;**670**:125125.
331. Wang Q, Zuo Z, Cheung CKC, Leung SSY. Updates on thermosensitive hydrogel for nasal, ocular and cutaneous delivery. *Int J Pharm* 2019;**559**:86–101.
332. Mahajan S, Nangare S, Chaudhari A, Patil G. Synthesis of chitosan-graphene oxide thermosensitive *in situ* hydrogel for nasal delivery of rasagiline mesylate: *in vitro*–*ex vivo* characterization. *J Drug Deliv Sci Technol* 2024;**95**:105549.
333. Bierbaumer L, Schwarze UY, Gruber R, Neuhaus W. Cell culture models of oral mucosal barriers: a review with a focus on applications, culture conditions and barrier properties. *Tissue Barriers* 2018;**6**:1479568.
334. Elsayed A, Al-Remawi M, Jaber N, Abu-Salah KM. Advances in buccal and oral delivery of insulin. *Int J Pharm* 2023;**633**:122623.
335. Morales JO, Brayden DJ. Buccal delivery of small molecules and biologics: of mucoadhesive polymers, films, and nanoparticles. *Curr Opin Pharmacol* 2017;**36**:22–8.
336. Kraan H, Vrieling H, Czerkinsky C, Jiskoot W, Kersten G, Amorij J-P. Buccal and sublingual vaccine delivery. *J Control Release* 2014;**190**:580–92.
337. Bisen AC, Biswas A, Dubey A, Sanap SN, Agrawal S, Yadav KS, et al. A review on polymers in ocular drug delivery systems. *MedComm – Biomater Appl* 2024;**3**:e77.
338. Madhav NVS, Shakya AK, Shakya P, Singh K. Orotransmucosal drug delivery systems: a review. *J Control Release* 2009;**140**:2–11.
339. Stegemann S, Ternik RL, Onder G, Khan MA, van Riet-Nales DA. Defining patient centric pharmaceutical drug product design. *AAPS J* 2016;**18**:1047–55.
340. Wasilewska K, Winnicka K. How to assess orodispersible film quality? A review of applied methods and their modifications. *Acta Pharm* 2019;**69**:155–76.
341. Patel VF, Liu F, Brown MB. Advances in oral transmucosal drug delivery. *J Control Release* 2011;**153**:106–16.
342. Krampe R, Visser JC, Frijlink HW, Breikreutz J, Woerdenbag HJ, Preis M. Oromucosal film preparations: points to consider for patient centricity and manufacturing processes. *Expet Opin Drug Deliv* 2016;**13**:493–506.
343. Sandhu PS, Kumar R, Beg S, Jain S, Kushwah V, Katore OP, et al. Natural lipids enriched self-nano-emulsifying systems for effective co-delivery of tamoxifen and naringenin: systematic approach for improved breast cancer therapeutics. *Nanomedicine* 2017;**13**:1703–13.
344. Balian GMFC, Luiz MT, Di Filippo LD, Chorilli M. Mucoadhesive liquid crystal precursor system for photodynamic therapy of oral cancer mediated by methylene blue. *Photodiagnosis Photodyn Ther* 2023;**44**:103739.
345. dos Santos Ramos MA, Calixto GMF, Gaspar de Toledo L, Vidal Bonifácio B, Campaner dos Santos L, Gottardo de Almeida MT, et al. Liquid crystal precursor mucoadhesive system as a strategy to improve the prophylactic action of *Syngonanthus nitens* (Bong.) ruhlmann against infection by *Candida krusei*. *Int J Nanomed* 2015;**16**:7455–66.
346. de Araújo PR, Calixto GMF, da Silva IC, de Paula Zago LH, Oshiro Junior JA, Pavan FR, et al. Mucoadhesive *in situ* gelling liquid crystalline precursor system to improve the vaginal administration of drugs. *AAPS PharmSciTech* 2019;**20**:225.
347. Liu Y, Qi X, Wang Y, Li M, Yuan Q, Zhao Z. Inflammation-targeted cannabidiol-loaded nanomicelles for enhanced oral mucositis treatment. *Drug Deliv* 2022;**29**:1272–81.
348. Blakaj A, Bonomi M, Gamez ME, Blakaj DM. Oral mucositis in head and neck cancer: evidence-based management and review of clinical trial data. *Oral Oncol* 2019;**95**:29–34.
349. das Neves J, Notario-Pérez F, Sarmiento B. Women-specific routes of administration for drugs: a critical overview. *Adv Drug Deliv Rev* 2021;**176**:113865.
350. Stevens JM. Gynaecology from ancient Egypt: the papyrus Kahun: a translation of the oldest treatise on gynaecology that has survived from the ancient world. *MJA (Med J Aust)* 1975;**2**:949–52.
351. Subi MTM, Selvasudha N, Vasanthi HR. Vaginal drug delivery system: a promising route of drug administration for local and systemic diseases. *Drug Discov Today* 2024;**29**:1040.
352. Katz DF, Yuan A, Gao Y. Vaginal drug distribution modeling. *Adv Drug Deliv Rev* 2015;**92**:2–13.
353. Gao Y, Yuan A, Chuchuen O, Ham A, Yang KH, Katz DF. Vaginal deployment and tenofovir delivery by microbicide gels. *Drug Deliv Transl Res* 2015;**5**:279–94.
354. Xin Y, Fei W, Zhang M, Chen Y, Peng Y, Sun D, et al. Uterine first-pass effect: unlocking the potential of vaginally administered ritodrine-loaded thermosensitive gel for uterine drug delivery. *Eur J Pharmaceut Sci* 2025;**204**:106945.
355. Ensign LM, Cone R, Hanes J. Nanoparticle-based drug delivery to the vagina: a review. *J Control Release* 2014;**190**:500–14.
356. Alexander NJ, Baker E, Kaptein M, Karck U, Miller L, Zampaglione E. Why consider vaginal drug administration?. *Fertil Steril* 2004;**82**:1–12.
357. Bulletti C, de Ziegler D, Flamigni C, Giacomucci E, Polli V, Bolelli G, et al. Targeted drug delivery in gynaecology: the first uterine pass effect. *Hum Reprod* 1997;**12**:1073–9.
358. Coler BS, Shynlova O, Boros-Rausch A, Lye S, McCartney S, Leimert KB, et al. Landscape of preterm birth therapeutics and a path forward. *J Clin Med* 2021;**10**:2912.
359. Osmatek T, Froelich A, Jadach B, Tatarek A, Gadziński P, Falana A, et al. Recent advances in polymer-based vaginal drug delivery systems. *Pharmaceutics* 2021;**13**:884.
360. Plummer EL, Bradshaw CS, Doyle M, Fairley CK, Murray GL, Bateson D, et al. Lactic acid-containing products for bacterial vaginosis and their impact on the vaginal microbiota: systematic review. *PLoS One* 2021;**16**:e0246953.
361. Notario-Pérez F, Galante J, Martín-Illana A, Cazorla-Luna R, Sarmiento B, Ruiz-Caro R, et al. Development of pH-sensitive vaginal films based on methacrylate copolymers for topical HIV-1 pre-exposure prophylaxis. *Acta Biomater* 2021;**121**:316–27.
362. Abou-Taleb HA, Fathalla Z, Naguib DM, Fatease A Al, Abdelkader H. Chitosan/solid-lipid nanoparticles hybrid gels for vaginal delivery of estradiol for management of vaginal menopausal symptoms. *Pharmaceutics* 2023;**16**:1284.
363. Obuobi S, Škalko-Basnet N. Understanding vaginal biofilms: the first step in harnessing antimicrobial nanomedicine. *J Control Release* 2024;**376**:1190–208.
364. Caramella CM, Rossi S, Ferrari F, Bonferoni MC, Sandri G. Mucoadhesive and thermogelling systems for vaginal drug delivery. *Adv Drug Deliv Rev* 2015;**92**:39–52.
365. Vanić Ž, Jøraholmen MW, Škalko-Basnet N. Nanomedicines for the topical treatment of vulvovaginal infections: addressing the challenges of antimicrobial resistance. *Adv Drug Deliv Rev* 2021;**178**:113855.
366. Campaña-Seoane M, Pérez-Gago A, Vázquez G, Conde N, González P, Martínez A, et al. Vaginal residence and pharmacokinetic preclinical study of topical vaginal mucoadhesive w/s emulsions containing ciprofloxacin. *Int J Pharm* 2019;**554**:276–83.
367. Shapiro RL, Bockley KM, Hsueh HT, Appell MB, Carter DM, Ortiz J, et al. Hypotonic, gel-forming delivery system for vaginal drug administration. *J Control Release* 2024;**371**:101–10.

368. Robinson TE, Clark C, Moakes RJA, Schofield Z, Moiem N, Geoghegan JA, et al. Simultaneous viscoelasticity and sprayability in antimicrobial acetic acid-alginate fluid gels. *Biomater Adv* 2025;**166**: 214051.
369. Gündel S da S, de Godoi SN, Santos RCV, da Silva JT, Leite LB de M, Amaral AC, et al. *In vivo* antifungal activity of nanoemulsions containing eucalyptus or lemongrass essential oils in murine model of vulvovaginal candidiasis. *J Drug Deliv Sci Technol* 2020;**57**: 101762.
370. Mittal RP, Rana A, Jaitak V. Essential oils: an impending substitute of synthetic antimicrobial agents to overcome antimicrobial resistance. *Curr Drug Targets* 2019;**20**:605–24.
371. Ho HM, Huang CY, Cheng YJ, Chen IH, Liu SJ, Huang C-H, et al. Squalene nanoemulsion reinforces mucosal and immunological fingerprints following intravaginal delivery. *Biomed Pharmacother* 2021;**141**:111799.
372. Jirwankar P, Gobbooru S, Shao J. Self-emulsified nanoemulsion for vaginal administration: *in vitro* study of effect on *actobacillus acidophilus*. *J Pharmacol Sci* 2020;**109**:3145–52.
373. Li S, Culkin A, Jones DS, Andrews GP. Development of polycaprolactone-based metronidazole matrices for intravaginal extended drug delivery using a mechanochemically prepared therapeutic deep eutectic system. *Int J Pharm* 2021;**593**:120071.
374. Ko J, Mandal A, Dhawan S, Shevachman M, Mitragotri S, Joshi N. Clinical translation of choline and geranic acid deep eutectic solvent. *Bioeng Transl Med* 2021;**6**:e10191.
375. Gowda S, Gajbhiye A, Bhatt S, Padia M, Mishra R, Agrawal-Rajput R. Emerging applicative areas of ionic liquids in biomedical sciences: unveiling new biological insights. *J Mol Liq* 2025;**427**: 127454.
376. Vitoria Pupo Silvestrini A, Wender Debiasi B, Garcia Praça F, Vitoria Lopes Badra Bentley M. Progress and challenges of lyotropic liquid crystalline nanoparticles for innovative therapies. *Int J Pharm* 2022;**628**:122299.
377. Manohar G, Srivastava P. Translational potential of nanoemulsion in wound healing — from discovery to product commercialization. *Nanotechnology* 2025;**7**:100187.
378. Nardin I, Köllner S. Successful development of oral SEDDS: screening of excipients from the industrial point of view. *Adv Drug Deliv Rev* 2019;**142**:128–40.
379. Uttreja P, Youssef AAA, Karnik I, Sanil K, Narala N, Wang H, et al. Formulation development of solid self-nanoemulsifying drug delivery systems of quetiapine fumarate *via* hot-melt extrusion technology: optimization using central composite design. *Pharmaceutics* 2024;**16**:324.
380. Friedl JD, Jörgensen AM, Le-Vinh B, Braun DE, Tribus M, Bernkop-Schnürch A. Solidification of self-emulsifying drug delivery systems (SEDDS): impact on storage stability of a therapeutic protein. *J Colloid Interface Sci* 2021;**584**:684–97.
381. Kumar M, Chawla PA, Faruk A, Chawla V. Spray drying as an effective method in the development of solid self-emulsifying drug delivery systems. *Curr Drug Deliv* 2023;**20**:508–25.
382. Mandić J, Zvonar Pobirk A, Vrečer F, Gašperlin M. Overview of solidification techniques for self-emulsifying drug delivery systems from industrial perspective. *Int J Pharm* 2017;**533**:335–45.
383. Silvestrini AVP, Caron AL, Viegas J, Praça FG, Bentley MVLB. Advances in lyotropic liquid crystal systems for skin drug delivery. *Expet Opin Drug Deliv* 2020;**17**:1781–805.
384. Ferreira DA, Bentley MVLB, Karlsson G, Edwards K. Cryo-TEM investigation of phase behaviour and aggregate structure in dilute dispersions of monoolein and oleic acid. *Int J Pharm* 2006;**310**: 203–12.
385. Lin P-C, Lin S, Wang PC, Sridhar R. Techniques for physicochemical characterization of nanomaterials. *Biotechnol Adv* 2014;**32**: 711–26.
386. Dong Y-D, Boyd BJ. Applications of x-ray scattering in pharmaceutical science. *Int J Pharm* 2011;**417**:101–11.
387. Heubach M-K, Schuett FM, Mayer J, Elkhafif OW, Engstfeld AK, Jacob T. Reproducibility of electrochemical measurements with ionic liquids: role of supplied batches, water and adventitious oxygen. *Top Catal* 2025;**68**:1910–23.
388. Mayrand-Provencher L, Rochefort D. Origin and effect of impurities in protic ionic liquids based on 2-methylpyridine and trifluoroacetic acid for applications in electrochemistry. *Electrochim Acta* 2009;**54**: 7422–8.
389. Valenti S, Cazorla C, Romanini M, Tamarit JL, Macovez R. Eutectic mixture formation and relaxation dynamics of coamorphous mixtures of two benzodiazepine drugs. *Pharmaceutics* 2023;**15**:196.
390. Rapalli VK, Waghule T, Hans N, Mahmood A, Gorantla S, Dubey SK, et al. Insights of lyotropic liquid crystals in topical drug delivery for targeting various skin disorders. *J Mol Liq* 2020;**315**: 113771.
391. Amorim AMB, Piochi LF, Gaspar AT, Preto AJ, Rosário-Ferreira N, Moreira IS. Advancing drug safety in drug development: bridging computational predictions for enhanced toxicity prediction. *Chem Res Toxicol* 2024;**37**:827–49.
392. Wu X, Shen M, Wang H, He X, Tan J, Wang R, et al. Evaluation of the efficacy and safety of ionic liquids containing ketoconazole in patients with tinea pedis: a randomized controlled clinical trial. *Bioeng Transl Med* 2023;**8**:e10463.
393. Pinal R. Enhancing the bioavailability of poorly soluble drugs. *Pharmaceutics* 2024;**16**:758.
394. Macedo AS, Castro PM, Roque L, Thomé NG, Reis CP, Pintado ME, et al. Novel and revisited approaches in nanoparticle systems for buccal drug delivery. *J Control Release* 2020;**320**:125–41.
395. Zidan G, Greene CA, Etxabide A, Rupenthal ID, Seyfoddin A. Gelatine-based drug-eluting bandage contact lenses: effect of PEGDA concentration and manufacturing technique. *Int J Pharm* 2021;**599**:120452.
396. Bijarchi MA, Favakeh A, Mohammadi K, Akbari A, Shafii MB. Ferrofluid droplet breakup process and neck evolution under steady and pulse-width modulated magnetic fields. *J Mol Liq* 2021;**343**: 117536.
397. Seino S, Ikehata H, Tanabe M, Umeda T, Tomiyama T, Tanaka A, et al. Investigating the efficacy of nasal administration for delivering magnetic nanoparticles into the brain for magnetic particle imaging. *J Control Release* 2024;**367**:515–21.
398. Xie J, Shen Z, Anraku Y, Kataoka K, Chen X. Nanomaterial-based blood–brain-barrier (BBB) crossing strategies. *Biomaterials* 2019;**224**:119491.
399. Mohseni A, Azimi AA, Bijarchi MA. Formation of magnetic double emulsions under steady and variable magnetic fields from a 3D-printed coaxial capillary device. *Anal Chim Acta* 2024;**1309**:342573.
400. Chen J, Xia L, Wu X, Du L, Liu R, Liu J, et al. The exploration of multifunctional liquid robotics with ferrofluids: fabrication, control, operation and sensing. *Nano Energy* 2024;**131**:110169.